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ORIGINAL ARTICLE

# Characterization of Chemical Composition and Antioxidant Properties of *Trachyspermum ammi* Seed as a Potential Medicinal Plant

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## KEYWORDS

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**ABSTRACT:** In this study, the essential oil obtained by hydrodistillation of the seed of *Trachyspermum ammi* (Umbelliferae), growing wild in Sabzevar, Khorasan Razavi province (Iran), were analyzed by GC and GC/MS. The yield of total volatiles was 2.3% (w/w). Forty four compounds representing 91.6% of the aerial parts oil were identified. The main components of the oil were Hexadecanoic acid (27.5%), ethyl linoleate (8.5%), 6-methyl- $\alpha$ -ionone (8.0%), isobutyl phthalate (5.8%),  $\alpha$ -cadinol (4.7%), germacrene D (4.3%) and  $\delta$ -cadinene (3.5%). The oil was rich in nonterpenoids (56.0%). The total flavonoid content of different extracts of the plant was found to be in the range 53.2-164.5 mg/g while the maximum amount concern to methanol extract. The antioxidant activity of the extracts was also measured by radical scavenging activity of antioxidants against free radical 1, 1-diphenyl-2-picrylhydrazyl (DPPH) method. Results showed that IC<sub>50</sub> of the extracts of *Trachyspermum ammi* seed are higher than the standard synthetic antioxidants, BHT, ascorbic acid and gallic acid.

## INTRODUCTION

All plants synthesize and accumulate secondary metabolites such as alkaloids, sterols, terpenes,

flavonoids, saponins, glycosides, volatile oils, etc., many of which have medicinal value. Since ancient times, nearly all cultures have used medicinal plants for the treatment of a variety of maladies and diseases.

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Furthermore, practitioners in many of these cultures often use the same or similar plants, which suggest that the active components of these plants have been found to be generally effective [1]. A plant used traditionally in Iran is *Trachyspermum ammi* L. (= *Trachyspermum copticum*=*Carum copticum*) [2]. As a part of on-going work on the medicinal plants of Iran, we decided to investigate the oil and antioxidant properties from *Trachyspermum ammi* seed.

*Trachyspermum ammi* (Umbelliferae) is an annual herb growing wild or planted in Iran [3, 4]. *Trachyspermum ammi*, commonly referred as bishop's weed, bullwort, omum plant and ajowan caraway (English names). Some biological effects of ajowan such as antiviral [5], anti-inflammatory [6], antifungal [7], analgesic [8] and antioxidant activity [9] have been reported.

In this study, the hydrodistilled volatile oil from seeds of *Trachyspermum ammi* from Davarzan, Khorasan province (Iran), was studied by GC and GC/MS. Total phenolic content and radical scavenging capacity were used to determination of antioxidant activity of different extracts of the plant's seed.

Antioxidants have already been found in plant materials and supplements. Due to their natural origin, the antioxidants obtained from plants are of greater benefit in comparison to synthetic ones [10]. The use of natural antioxidants from plants does not induce side effects, while synthetic antioxidants were found to have genotoxic effect [11]. Therefore, the investigations of biological activity and chemical composition of medicinal plants as a potential source of natural antioxidants are numerous. The basic aim of the research was to determine the total phenolic content and radical scavenging capacity in various extracts of the species *Trachyspermum ammi* seed using spectrophotometric methods.

## MATERIALES AND METHODS

### Chemicals

Solvents (methanol, chloroform, ethyl acetate) were purchased from Merck (Darmstadt-Germany) products. Standards of phenolic acids (gallic acid) and 2, 2-diphenyl-1-picrylhydrazyl (DPPH) were obtained from Sigma Chemicals Co., St Louis, MO, USA. The Folin-Ciocalteu's phenol reagent and 3-tert-butyl-4-hydroxyanisole (BHA) were from FlukaChemie AG, Buchs, Switzerland. All other solvents and chemicals were of analytical grade.

### Plant Material

The plant material was collected in July 2013 from Davarzan in Khorasan Province, Iran. A voucher specimen has been deposited in the herbarium of Research Center of Natural Resources, Sabzevar, Iran. The collected plant material was air-dried in darkness at room temperature (20 °C). Seeds of dried plant were separated from plant and stored in tight-seal dark containers until needed.

### Essential oil isolation

Air-dried seeds of *Trachyspermum ammi* (100 g) were subjected to hydrodistillation in a Clevenger-type apparatus for three hours to produce oils. The yield of total volatiles was 2.3% (w/w). The oils were dried over anhydrous sodium sulfate and stored in sealed vials at 4 °C before analysis.

### GC analysis

GC analysis was performed using a Shimadzu GC-9A gas chromatograph, equipped with a HP-5MS fused silica column (30 m×0.25 mm i.d., film thickness 0.25 μm). The oven temperature was held at 50 °C for five minutes and then programmed to 250 °C at a rate of 3 °C/min. The injector and detector (FID) temperatures were 290°C. Helium was used as carrier gas with a linear velocity of 32 cm/s.

### **GC/MS analysis**

GC/MS analysis was carried out on a Hewlett-packard 6890 gas chromatograph fitted with a fused silica HP-5MS capillary column (30m×0.25 mm; film thickness 0.32 μm). The oven temperature was programmed from 60 to 220 °C at 6 °C/min. Helium was used as carrier gas at a flow rate of 1 mL/min. The chromatograph was coupled to a Hewlett-Packard 5973 mass selective detector with an ionization voltage of 70 eV.

### **Qualitative and quantitative analyses**

Constituents of the volatile oils were identified by comparison of their retention indices relative to C9-C21 n-alkanes and of their mass spectral fragmentation pattern with those reported in the literature [12] and stored in a MS library (Wiley 275). The quantification of the components was performed on the basis of their GC peak area data from the HP-5MS column separation.

### **Preparation of the extracts**

The conventional maceration method was used for preparing the extracts. Dried and powdered aerial parts of *Trachyspermum ammi* (50 g) extracted by 400 mL of solvents. This process replicated three times with the same volume of fresh solvent. Three solvents having different polarities (methanol, ethyl acetate, chloroform) were used. An overhead stirrer mixed the materials for 24 h. All the mixtures were filtered through Whatman paper No. 41. The solvents were removed below 40°C using a rotary evaporator (Heidolph, Germany) and stored at 4 °C for further use.

### **Determination of Total Phenolic Contents**

The total phenolic content of extract was determined spectrophotometrically by Folin-Ciocalteu method according to the procedure reported by Singleton et al. [13] with some modifications. Briefly, 500 μl of (methanol, ethyl acetate, chloroform) extracts solution, 1500 μl distilled water and 500 μl of 1:10 Folin-Ciocalteu reagent were mixed for 1 minute. Then 5 minutes later, 1000 μl of sodium carbonate (5.0%) were added and shaken. After two hours in the dark of

incubation at room temperature, the absorbance at 760 nm was measured by a UV-Visible spectrophotometer, (Unico UV-2100, China). The total phenolic concentration was calculated from gallic acid (GA) calibration curve (5-100 mg/L). Total phenolic content were expressed as gallic acid equivalents (GAE)/g of extracts averaged from three replicates.

### **DPPH Radical Scavenging Activity Assay**

The ability of the plant extract to scavenge DPPH free radicals was assessed by the standard method Mensor et al. [14]. Briefly, 10 μL of the extracts was added to 2.5 mL of a 0.004% solution of DPPH in methanol. The stock solution of extracts were prepared in concentration of 1 mg/ml. Dilutions were made to obtain concentrations of 20, 40, 60 and 80 μg/ml. Diluted solutions (1 ml each) were mixed with 1 ml of methanolic solution of DPPH in concentration of 0.004%. After 30 min incubation in darkness at room temperature (23 °C), the absorbance was recorded at 517 nm. Ascorbic acid, gallic acid and BHT (20, 40, 60, 80 μg/mL) were used as positive references. Control contained all the reagents except the extract. Percentage of scavenged DPPH was calculated using equation 1. The data were presented as mean values ± standard deviation (n = 3).

$$\text{DPPH scavenging activity} = 100 \times (\text{Ac} - \text{As})/\text{Ac} \quad (1)$$

where Ac is the absorbance of the control and As is the absorbance of the sample. IC<sub>50</sub> values calculated denote the concentration of the sample required to decrease the absorbance at 517 nm by 50%.

## **RESULTS AND DISCUSSION**

As a part of on-going work on the chemical analysis of oils obtained from the wild plants of Iran, we decided to re-investigate the oils of this specific plant. Hydrodistilled volatile oils from the crushed dry seeds of *Trachyspermum ammi* (Umbelliferae) from Davarzan (Iran), was studied by GC and GC/MS. The air-dried

aerial parts of the plant yielded 2.3% (w/w) oil. The oil was clear and yellowish. Forty four components were identified in the seed's oil that contained 91.6% of the compounds. Table 1 lists formulas, percentages, and retention indices of identified compounds in the oil. As can be seen, the main components are Hexadecanoic acid (27.5%), ethyl linoleate (8.5%), 6-methyl- $\alpha$ -ionone (8.0%), isobutyl phthalate (5.8%),  $\alpha$ -cadinol (4.7%), germacrene D (4.3%) and  $\delta$ -cadinene (3.5%).

In this study, GC and GC/MS analysis method revealed several nonterpenoid hydrocarbons (NH), monoterpenoid hydrocarbons (MH), sesquiterpenoid hydrocarbons (SH), oxygenated sesquiterpenes (OS), oxygenated monoterpenes (OM), diterpene hydrocarbon (DH) and oxygenated diterpene (OD) in the oil from the

seed of *Trachyspermum ammi*. Onemoterpene hydrocarbons (0.1%), five oxygenated monoterpene (8.8%), thirteen sesquiterpene hydrocarbons (13.4%), six oxygenated sesquiterpene (10.4%), seventeen nonterpene hydrocarbons (56.0%), one diterpene hydrocarbon (2.5%) and one oxygenated diterpene (0.4%) were detected in this oil. The data lead to a rank order of constituent groups: NH>SH>OS>OM>DH>OD>MH for the seed oil. The main components in this oil were Hexadecanoic acid (27.5%), ethyl linoleate (8.5%), 6-methyl- $\alpha$ -ionone (8.0%), isobutyl phthalate (5.8%),  $\alpha$ -cadinol (4.7%), germacrene D (4.3%) and  $\delta$ -cadinene (3.5%).

**Table 1.** Constituents of the essential oils from seed of *Trachyspermum ammi* obtained by hydrodistillation<sup>a</sup>

No.	compound	Formula	Percentage	RRI <sup>b</sup>	Class
1	Limonene	C <sub>10</sub> H <sub>16</sub>	0.1	1031	MH <sup>c</sup>
2	4-Terpineol	C <sub>10</sub> H <sub>18</sub> O	0.1	1179	OM <sup>d</sup>
3	Fenchyl acetate	C <sub>12</sub> H <sub>20</sub> O <sub>2</sub>	0.4	1223	OM
4	Anethole	C <sub>10</sub> H <sub>12</sub> O	0.1	1285	OM
5	$\alpha$ -Copaene	C <sub>15</sub> H <sub>24</sub>	0.2	1364	SH <sup>e</sup>
6	$\beta$ - Bourbonene	C <sub>15</sub> H <sub>24</sub>	0.8	1385	SH
7	$\beta$ -Cubebene	C <sub>15</sub> H <sub>24</sub>	0.1	1390	SH
8	Tetradecane	C <sub>14</sub> H <sub>30</sub>	0.1	1400	NH <sup>f</sup>
9	$\beta$ -Caryophyllene	C <sub>15</sub> H <sub>24</sub>	0.3	1418	SH
10	$\alpha$ -Guaiene	C <sub>15</sub> H <sub>24</sub>	0.1	1439	SH
11	Aromadendrene	C <sub>15</sub> H <sub>24</sub>	0.6	1442	SH
12	$\alpha$ -Humulene	C <sub>15</sub> H <sub>24</sub>	2.0	1452	SH
13	(E)- $\beta$ -Farnesene	C <sub>15</sub> H <sub>24</sub>	0.1	1457	SH
14	Germacrene D	C <sub>15</sub> H <sub>24</sub>	4.3	1480	SH
15	$\beta$ -Ionone	C <sub>13</sub> H <sub>20</sub> O	0.2	1488	OM
16	Bicyclogermacrene	C <sub>15</sub> H <sub>24</sub>	0.7	1500	SH
17	$\gamma$ -Cadinene	C <sub>15</sub> H <sub>24</sub>	0.6	1515	SH
18	6-Methyl- $\alpha$ -ionone	C <sub>14</sub> H <sub>22</sub> O	8.0	1518	OM
19	$\delta$ -Cadinene	C <sub>15</sub> H <sub>24</sub>	3.5	1522	SH
20	Cadina-1, 4-diene	C <sub>15</sub> H <sub>24</sub>	0.1	1533	SH
21	Germacrene D-4-ol	C <sub>15</sub> H <sub>26</sub> O	0.6	1574	OS <sup>g</sup>
22	Spathulenol	C <sub>15</sub> H <sub>24</sub> O	1.5	1578	OS
23	Caryophyllene oxide	C <sub>15</sub> H <sub>24</sub> O	0.5	1583	OS
24	Humulene epoxide II	C <sub>15</sub> H <sub>24</sub> O	1.7	1608	OS
25	$\tau$ -Muurolol	C <sub>15</sub> H <sub>26</sub> O	1.4	1643	OS
26	$\alpha$ -Cadinol	C <sub>15</sub> H <sub>26</sub> O	4.7	1656	OS
27	Tetradecanoic acid	C <sub>14</sub> H <sub>28</sub> O <sub>2</sub>	1.8	1760	NH
28	Octadecane	C <sub>18</sub> H <sub>38</sub>	0.4	1800	NH

Continue of Table 1

29	6, 10, 14-Trimethyl-2-Pentadecanone,	C <sub>18</sub> H <sub>36</sub> O	1.7	1848	NH
30	2-Hydroxy-Cyclopentadecanone	C <sub>15</sub> H <sub>28</sub> O <sub>2</sub>	0.4	1853	NH
31	Pentadecanoic acid	C <sub>15</sub> H <sub>30</sub> O <sub>2</sub>	0.3	1867	NH
32	Isobutyl phthalate	C <sub>16</sub> H <sub>22</sub> O <sub>4</sub>	5.8	1877	NH
33	Cyclohexadecane	C <sub>16</sub> H <sub>32</sub>	0.3	1883	NH
34	16-methyl-Oxacyclohexadecan-2-one,	C <sub>16</sub> H <sub>30</sub> O <sub>2</sub>	0.3	1943	NH
35	Sandaracopimara-8(14), 15-diene	C <sub>20</sub> H <sub>32</sub>	2.5	1969	DH <sup>b</sup>
36	di-Butylphthalate	C <sub>16</sub> H <sub>22</sub> O <sub>4</sub>	0.9	1973	NH
37	Hexadecanoic acid	C <sub>16</sub> H <sub>32</sub> O <sub>2</sub>	27.5	1977	NH
38	Eicosane	C <sub>20</sub> H <sub>42</sub>	2.0	2000	NH
39	Heptadecanoic acid	C <sub>17</sub> H <sub>34</sub> O <sub>2</sub>	0.4	2065	NH
40	Methyl linoleate	C <sub>19</sub> H <sub>34</sub> O <sub>2</sub>	0.6	2084	NH
41	Phytol	C <sub>20</sub> H <sub>40</sub> O	0.4	2111	OD <sup>i</sup>
42	(Z, Z)-9,12-Octadecadienoic acid	C <sub>18</sub> H <sub>32</sub> O <sub>2</sub>	2.7	2136	NH
43	Ethyl linoleate	C <sub>20</sub> H <sub>36</sub> O <sub>2</sub>	8.5	2164	NH
44	Octadecanoic acid	C <sub>18</sub> H <sub>36</sub> O <sub>2</sub>	2.3	2172	NH
	Total identified			91.6	

<sup>a</sup>The compounds have been arranged according to their retention indices on an HP-5 MS capillary column

<sup>b</sup>Kovatz retention indices given in the literature

<sup>c</sup>Monoterpene hydrocarbons

<sup>d</sup>Oxygenated monoterpene

<sup>e</sup>Sesquiterpene hydrocarbons

<sup>f</sup>Nonterpene hydrocarbons

<sup>g</sup>Oxygenated sesquiterpene

<sup>h</sup>Diterpene hydrocarbon

<sup>i</sup>Oxygenatedditerpene

### Antioxidant activity

Methanol, ethyl acetate and chloroform extracts were prepared to examine the total phenolic content and antioxidant activity. The total phenolic contents in the examined plant extracts using the Folin-Ciocalteu's reagent is expressed in terms of Gallic acid equivalent (the standard curve equation:  $Y=0.0086x + 0.0175$ ,  $r^2 = 0.999$ ). The values obtained for the concentration of total phenols are expressed as mg of GA/g of dry extract (Table 2).

The total phenolic contents in the examined extracts ranged from 53.2 to 164.5 mgGA/g of dry extract.

The total phenolic contents in plant extracts of the species *Trachyspermum ammi* depends on the type of extract, i.e. the polarity of solvent used in extraction. High solubility of phenols in polar solvents provides high concentration of these compounds in the extracts obtained using polar solvents for the extraction [15].

**Table 2.** Total phenolic contents in the plant extracts expressed in terms of gallic acid equivalent (mg of GA/g of extract)

Extract	Absorbance	mg of GA/g of extract <sup>1</sup>
Chloroform	1.37	82.0±1.6
Ethyl acetate	1.75	53.2±2.8
Methanol	1.51	164.5±1.3

<sup>1</sup> Each value is the average of three analyses ± standard deviation.

The antioxidant activity of different seed extracts from *Trachyspermum ammi* was determined using a methanol solution of DPPH reagent. DPPH is very stable free radical. Unlike invitro generated free radicals such as the hydroxyl radical and superoxide anion, DPPH has the advantage of being unaffected by certain side reactions, such as metal ion chelation and enzyme inhibition. A freshly prepared DPPH solution exhibits a deep purple color with an absorption maximum at 517 nm. This purple color generally fades when antioxidant molecules quench DPPH free radicals (i.e. by providing hydrogen atoms or by electron donation, conceivably via a free-radical attack on the DPPH molecule) and convert them into a colorless-bleached product (i.e. 2, 2-diphenyl-1-hydrazine, or a substituted analogous hydrazine), resulting in a decrease in absorbance at 517 nm band [16]. The antioxidant activity of five different extracts from the species *Trachyspermum ammi* is expressed in terms of percentage of inhibition (%) and IC<sub>50</sub> values (µg/ml) (Table 3). Parallel to examination of the antioxidant activity of plant extracts, the values for three standard compounds were obtained and compared to the values of the antioxidant activity. The standard substances were BHT, Gallic acid and ascorbic acid.

The examination of antioxidant activities of plant extracts from *Trachyspermum ammi* showed different

values. The obtained values varied from 9.0% to 103.8%. The largest capacity to neutralize DPPH radicals were found for methanol extract, which neutralized 50% of free radicals at the concentration of 74.5 µg/ml. A moderate activity was found for chloroform extract. The minutest capacity to inhibit DPPH radicals was determined for ethyl acetate. In comparison to IC<sub>50</sub> values of BHT, ascorbic acid and Gallic acid, methanol extract from *Trachyspermum ammi* manifested the moderate capacity for neutralization of DPPH radicals.

The extraction of antioxidant substances of different chemical structure was achieved using solvents of different polarity. Numerous investigations of qualitative composition of plant extracts revealed the presence of high concentrations of phenols in the extracts obtained using polar solvents. The extracts that perform the highest antioxidant activity (Table 3) have the highest concentration of phenols (Table 2). Between the values for the concentration of phenolic compounds (Table 2) and antioxidant activity of different plant extracts of *Trachyspermum ammi* has been proved a significant linear correlation. Methanol extract from *Trachyspermum ammi* have high concentration of total phenols (Table 2) and DPPH scavenging capacity (Table 3), which is in correlation with intense antioxidant activity of these extract.

**Table 3.** Antioxidant (DPPH scavenging) activity of investigated seed extracts and synthetic antioxidants presented as percentage of DPPH radicals inhibition and IC<sub>50</sub> values (µg/ml).

Extract	DPPH				
	20 ppm	40 ppm	60 ppm	80 ppm	IC <sub>50</sub>
Chloroform	12.3±2.3	27.0±2.1	38.2±1.5	47.1±2.1	84.6±3.4
Ethyl acetate	9.0±2.5	16.4±2.4	24.8±1.6	38.4±5.7	103.8±4.3
Methanol	17.5±1.2	23.1±2.6	32.4±2.3	59.0±4.7	74.5±4.9
Synthetic antioxidant	20 ppm	40 ppm	60 ppm	80 ppm	IC <sub>50</sub>
BHT	76.0±4.8	93.8±0.8	94.6±1.1	96.7±0.9	14.9±0.9
Ascorbic acid	34.2±5.2	51.8±2.4	67.2±0.6	85.5±1.7	40.3±1.1
Gallic acid	82.8±1.6	91.0±1.9	91.5±2.6	92.6±0.9	7.9±1.2

## CONCLUSIONS

The chemical composition of the essential oil of seed from *Trachyspermum ammi* (Umbelliferae) growing in Davarzan, northeast of Iran, was investigated. This study showed considerable amounts of Hexadecanoic acid (27.5%), ethyl linoleate (8.5%), 6-methyl- $\alpha$ -ionone (8.0%), isobutyl phthalate (5.8%),  $\alpha$ -cadinol (4.7%), germacrene D (4.3%) and  $\delta$ -cadinene (3.5%). These results demonstrated that the chemical composition of the essential oil of the same species can change depending on a variety of conditions, including climate, time of collection, and the ground composition of the sampling area besides of growth stages of plant. Also results of our study suggest the great value of the species *Trachyspermum ammi* for use in pharmacy and phytotherapy. Based on this information, it could be concluded that this plant is natural sources of antioxidant substances of high importance. It is noticed that the highest concentration of phenolic compounds in the extracts were obtained using solvents of high polarity; the methanol extract manifested greater power of extraction for phenolic compounds from *Trachyspermum ammi*. The high contents of phenolic compounds and the values of antioxidant activity indicated that these compounds contribute to the strong antioxidant activity. Further studies of this plant species should be directed to carry out in vivo studies of its medicinal active components in order to prepare natural pharmaceutical products of high value.

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## REFERENCES

1. Omidbeigi R., 2000. Production and Processing of Medicinal Plants, Vol. I, second edition. Astan Quds Razavi Publications. [In Persian]
2. Hopper D., 1937. Useful plants and drugs of Iran and Iraq. Field Museum of Natural History—Botany, Vol. IX, Field Museum, Chicago, IL, USA.
3. Rechinger K.H., 1986. *Sclerorhachis*, in: Flora Iranica, Compositae No. 158. Edits., K.H. Rechinger and I.C. Hedge, P 47, Akademische Druck and Verlagsanstalt, Graz, Austria.
4. Mozaffarian V., 1996. A Dictionary of Iranian plant names. Farhang Moaser Publishers, Tehran, 1st Edit., p 491.
5. Hussein G.H., Miyashiro H., Nakamura N., Hattori M., Kakiuchi N., Shimotohno K., 2000. Inhibitory effects of Sudanese medicinal plant extracts on hepatitis C virus (HCV) protease. *Phytother Res.*, 14, 510–516.
6. Thangam C., Dhananjayan R., 2003. Anti-inflammatory potential of the seeds of *Carum copticum* Linn. *Ind J Pharmacol.* 34, 388–391.
7. Rasooli I., Fakoor M.H., Yadegarinia D., Gachkar L., Allameh A., Rezaei M.B., 2008. Anti Mycotoxigenic characteristics of *Rosmarinus officinalis* and *Trachyspermum copticum* L. essential oils. *Int. J Food Microbiol.* 122, 135–139.
8. Dashti-Rahmatabadi M.H., Hejazian S.H., Morshedi A., Rafati A., 2007. The analgesic effect of *Carum copticum* extract and morphine on phasic pain in mice. *J Ethnopharmacol.* 109, 226–228.
9. Bera D., Lahiri D., Nag A., 2004. Novel natural antioxidant for stabilization of edible oil: the ajowan (*Carum copticum*) extract case. *Journal of the American Oil Chemists Society JAACS.* 81, 169–172.



10. Rohman A., Riyanto S., Yuniarti N., Saputra W.R., Utami R., 2010. Antioxidant activity, total phenolic, and total flavonoid of extracts and fractions of redfruit (*Pandanusconoides*Lam). *Int Food Res J.* 17, 97-106.
11. Chen C., Pearson M.A., Gray I.J., 1992. Effects of synthetic antioxidants (BHA, BHT and PG) on the mutagenicity of IQ-like compounds. *Food Chem.* 43, 177-183.
12. Adams R.P., 2007. Identification of Essential oil Components by Gas Chromatography/Mass Spectrometry. Allured Pub. Corp., Carol Stream, IL., USA.
13. Singleton V.L., Orthofer R., Lamuela-Raventos R.M., 1999. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin–Ciocalteu reagent. *Meth Enzymol.* 299, 152–178.
14. Mensor L.I, Menezes F.S., Leitao G.G., Reis A.S., Dos Santos T., Coube C.S., Leitao S.G., 2001. Screening of Brazilian plants extracts for antioxidants activity by use of DPPH free radical method. *Phytother Res.* 15, 127-130.
15. Mohsen M.S., Ammar S.M.A., 2008. Total phenolic contents and antioxidant activity of corn tassel extracts. *Food Chem.* 112, 595-598.
16. Amarowicz R., Pegg B.R., Rahimi-moghaddam P., Bar B., Weil J.A., 2003. Free-radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies. *Food Chem.* 84, 551-562.

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روش تحقیق کمی



آموزش مهارت‌های کاربردی در تدوین و چاپ مقالات ISI

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آموزش نرم افزار Word برای پژوهشگران

آموزش نرم افزار Word برای پژوهشگران