Short Paper

Commercial polyester/cotton fabric (Damour): a novel prosthetic material for hernioplasty in ruminants

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Summary

The aim of this work was to study the possibility of using a new commercial polyester/cotton fabric to repair large sized umbilical and abdominal hernias in calves and small ruminants. Twenty-four animals (16 calves, 3 buffalo calves, 4 sheep, 1 goat) with umbilical or abdominal hernia were included in this study. The diameter of hernial ring ranged from (12-25) cm. An underlay technique was used for fixation of the prosthetic material that was placed under the visceral peritoneum after omentopexy and were secured to the recipient tissue with interrupted overlapped pattern using polypropylene monofilament suture material. After one year follow up, normal healing without complications in 19 animals was revealed (79.82%). Wound infection and delayed healing were recorded in 3 animals (12.5%) and wound dehiscence and re-herniation were observed in 2 cases (8.3%). In conclusion, our results suggested that commercial polyester/cotton fabric (Damour) represents a new cost effective alternative to commonly used meshes for hernioplasty of large sized hernias with relatively low rate of complications and recurrence in calves and small ruminants.

Key words: Hernioplasty, Polyester, Cotton, Calve, Small ruminants

Introduction

Surgical management of major abdominal wall defects remains a significant problem. Many abdominal wall defects can be repaired by primary closure; however, if the defect is large and there is a tension on the closure of the wound, the use of prosthetic materials is indicated (Park and Lakes, 1992). In food animals, hernias of sufficient size to necessitate prosthetic reconstruction were most commonly caused by umbilical hernias without previous repair in older heifers, umbilical hernias that had been repaired unsuccessfully one or more times, and incisional hernias following right paramedian abomasopexy or ventral midline celiotomy (Tulleners and Fretz, 1983). Surgical biomaterials may include substances that have the capacity to interact with living tissue and are not rejected by the body, and then are either absorbed naturally by the body and eliminated by biological processes or become a permanent part of the surrounding tissue (Kartinos, 2005). The frequently used biomaterials include monofilament and multifilament polypropylene meshes, multifilament polyester mesh (mersiline), multifilament polytetrafluoroethylene meshes (Teflon) and expanded polytetrafluoroethylene (ePTFE) (Amid, 1997). Shoukry et al. (1997) also investigated the use of a commercial polyester fabric for repair of abdominal hernia in different animals. Of importance, the most effective positioning of the prosthesis is also a subject of debate. Three techniques for implanting biomaterials to bridge an abdominal wall defect that cannot be primarily closed have been described. These include the onlay, inlay and underlay techniques (Larson and Vandertoll, 1984; Rainier et al., 2006). The underlay technique was used successfully with several advantages over other techniques and with relatively fewer complications and low hernia recurrence rates (Durrart et al., 2001). The high cost associated with synthetic material initiated the search for safe and cheap prosthetic material that can replace the commonly used highly expensive meshes. Therefore, the aim of the present study was to try the use of a new commercial polyester/cotton fabric as an available, cheap material and of adequate strength for repair of large sized umbilical and abdominal hernias in calves, sheep and goat.

Materials and Methods

Animals

Twenty four animals, whose signalsments are shown in Table 1 suffering umbilical or abdominal hernia with an average diameter of hernial ring ranged from (12-25) cm and that were diagnosed at the Mansoura Veterinary Teaching Hospital at Mansoura University and different farms in Dakahlia province, Egypt during the period from January 2009 to November 2013 were included in this study.

Surgical techniques

Animals were sedated using Xylazine HCL 2%
(Xyject-ADWIA-Egypt) (0.1 mg/kg for calf, 0.05 mg/kg for ewe and 0.01 mg/kg for goat) and the site of operation was infiltrated using Lidocaine HCL 2% (Debacaine-AlDebiky-Egypt) diluted to 1% solution to provide sufficient volume for injection at 1 cm intervals along the injection site. After aseptic preparation, the affected animals were controlled in lateral or dorsal recumbency and a half circle or an elliptical skin incision was performed. The fibrous adhesions were dissected and the devitalized tissues were removed to expose the hernial ring. Following proper haemostasis, the hernial sac was incised along one side of the hernial ring margin to create a fascial flap. The fibrous edge of the hernial ring was refreshed before implantation of the graft. Underlay technique for implantation (Fig. 1) was used where a sterilized piece of commercial polyester/cotton fabric produced by (Misr Spinning and Weaving Co., Mahalla al-Kubra-Egypt) was prepared according to the size of the hernial ring and fashioned to the shape of the defect allowing for 4-5 mm underlay then placed under the visceral peritoneum after omentotomy (part of omentum was grasped and loosely stitched to the implant). The fabric was secured to the recipient tissue with interrupted overlapped pattern using No. (1) Polypropylene monofilament suture material (Prolene Ethicon; Johnson and Johnson, Brussels, Belgium) (Fig. 2B). The implant was covered with a fascial flap derived from the hernial sac when possible and the skin was sutured with horizontal mattress suture using silk No. (1). Postoperative care included injection of penicillin G procaine (Norocillin L.A., Norbrook Lab. Ltd., Newry, Ireland) and Flumixin meglumine (Flamicure, Pharma Swede, 10th of Ramadan, Egypt by 2.2 mg/kg) for 5 days post-surgery. The skin wound was dressed routinely with povidone iodine (Betadine-topical antiseptic). The operated animals were followed either by regular visits or calling the owner during one year post surgery.

Results

Umbilical hernias were diagnosed in 17 animals (16

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Animal species</th>
<th>Sex</th>
<th>Weight (kg)</th>
<th>Age, D (M)</th>
<th>Origin</th>
<th>Character of hernial ring</th>
<th>Cause</th>
<th>Location</th>
<th>Size (cm)</th>
<th>Shape</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Calf</td>
<td>Male</td>
<td>85</td>
<td>5 M</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>12-18</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Calf</td>
<td>Female</td>
<td>35</td>
<td>6 D</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>14-16</td>
<td>Indefinite</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Calf</td>
<td>Female</td>
<td>170</td>
<td>8 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>18-22</td>
<td>Circular</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Calf</td>
<td>Male</td>
<td>80</td>
<td>5 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>15-20</td>
<td>Circular</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Calf</td>
<td>Female</td>
<td>50</td>
<td>45 D</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>13-18</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Calf</td>
<td>Female</td>
<td>60</td>
<td>6 M</td>
<td>After excision of umbilical vein abscess</td>
<td>Umbilical</td>
<td>12-15</td>
<td>Circular</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>Calf</td>
<td>Male</td>
<td>105</td>
<td>7 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>15-22</td>
<td>Semicircular</td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Calf</td>
<td>Female</td>
<td>40</td>
<td>8 D</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>14-20</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>Calf</td>
<td>Female</td>
<td>30</td>
<td>3 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>14-19</td>
<td>Circular</td>
<td></td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>Calf</td>
<td>Male</td>
<td>85</td>
<td>4 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>12-15</td>
<td>Circular</td>
<td></td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>Calf</td>
<td>Female</td>
<td>45</td>
<td>5 D</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>15-22</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>Calf</td>
<td>Female</td>
<td>40</td>
<td>4 D</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>22-23</td>
<td>Circular</td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>Calf</td>
<td>Female</td>
<td>70</td>
<td>2 M</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>12-18</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>14</td>
<td>Calf</td>
<td>Female</td>
<td>75</td>
<td>5 M</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>16-19</td>
<td>Circular</td>
<td></td>
</tr>
<tr>
<td>15</td>
<td>Calf</td>
<td>Male</td>
<td>90</td>
<td>4 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>12-18</td>
<td>Indefinite</td>
<td></td>
<td></td>
</tr>
<tr>
<td>16</td>
<td>Calf</td>
<td>Male</td>
<td>65</td>
<td>2 M</td>
<td>Congenital</td>
<td>Umbilical</td>
<td>Congenital</td>
<td>12-17</td>
<td>Indefinite</td>
<td></td>
</tr>
<tr>
<td>17</td>
<td>Buffalo calf</td>
<td>Female</td>
<td>100</td>
<td>5 M</td>
<td>Recurrent after fault operation</td>
<td>Umbilical</td>
<td>14-21</td>
<td>Oval</td>
<td></td>
<td></td>
</tr>
<tr>
<td>18</td>
<td>Buffalo calf</td>
<td>Male</td>
<td>90</td>
<td>3 M</td>
<td>Traumatic</td>
<td>At level of the left stifle fold</td>
<td>Traumatic</td>
<td>16-22</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>19</td>
<td>Buffalo calf</td>
<td>Female</td>
<td>155</td>
<td>9 M</td>
<td>Recurrent after fault operation</td>
<td>At the left ventral abdominal wall</td>
<td>Traumatic</td>
<td>13-19</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>20</td>
<td>Ewe</td>
<td>Female</td>
<td>40</td>
<td>27 M</td>
<td>Traumatic</td>
<td>Ventral hypogastrium</td>
<td>Traumatic</td>
<td>13-20</td>
<td>Indefinite</td>
<td></td>
</tr>
<tr>
<td>21</td>
<td>Ewe</td>
<td>Female</td>
<td>55</td>
<td>60 M</td>
<td>Traumatic</td>
<td>Behind the right costal arch</td>
<td>Traumatic</td>
<td>13-23</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>22</td>
<td>Ewe</td>
<td>Female</td>
<td>60</td>
<td>75 M</td>
<td>Following parturition</td>
<td>At level of the right stifle fold</td>
<td>Traumatic</td>
<td>12-20</td>
<td>Oval</td>
<td></td>
</tr>
<tr>
<td>23</td>
<td>Ewe</td>
<td>Female</td>
<td>60</td>
<td>85 M</td>
<td>Traumatic</td>
<td>Near the umbilicus</td>
<td>Traumatic</td>
<td>14-25</td>
<td>Semicircular</td>
<td></td>
</tr>
<tr>
<td>24</td>
<td>Goat</td>
<td>Female</td>
<td>30</td>
<td>29 M</td>
<td>Traumatic</td>
<td>At level of the right stifle fold</td>
<td>Traumatic</td>
<td>14-20</td>
<td>Semicircular</td>
<td></td>
</tr>
</tbody>
</table>

Fig. 1: Illustration of the under lay technique

Fig. 2: Showed large umbilical hernia in a newly born calf (A) and after hernioplasty using polyester/cotton fabric (B)
calves and 1 buffalo calf) (Fig. 2A), while abdominal hernias were diagnosed in 7 animals (2 buffalo calves, 4 sheep and 1 goat) (Fig. 3A). The sex, age, weight, origin, location of abdominal hernias and characteristics of hernial rings were illustrated in Table 1.

The hernial swellings were reducible in animals presented with congenital hernias while in the acquired ones there were varying degrees of irreducibility and adhesions.

Typical signs of inflammation were noticed from the first day post-operative and the whole site of operation was painful to manual palpation in all animals. These signs were more severe in animals with abdominal recurrent or traumatic hernia. The inflammatory reaction subsided gradually within 2 weeks. Slight edematous swelling and seroma formation were developed 2 days post operation in all animals.

A slight hard mass of tissue with thickness about 1 to 2 cm and of nearly equal size to the implanted fabric was palpated at the site of operation in all animals for the first 2 weeks post operation. This mass subsided by the end of the 6th week post operation and could be hardly felt at the 8th week.

According to the gross appearance of the wound after surgery, the healing characteristics were categorized into 3 grades illustrated in Table 2. Normal healing was recorded in 19 animals (79.2%) (14 animals with umbilical hernia and 5 with abdominal hernia) in which the abdominal wall regained its integrity, skin wound was satisfactory healed and skin stitches were removed 10 days post operation (Fig. 3B).

Wound infection, stitch abscesses and presence of necrotic foci with formation of sinus tracts were recorded in 3 cases (12.5%) (two with umbilical and one with abdominal hernia). These complications responded well to topical antiseptic dressing and skin wound healing was achieved within 3 weeks.

Wound dehiscence and re-herniation were recorded in 2 cases (8.3%), one calf has developed a persistent draining sinus discharging pus; hernia was recurrent after three weeks that rendered reoperation and removal of the infected mesh. In another ewe, hernia recurrence was recorded following parturition that occurred 10 weeks post-surgery. During reoperation, implant dehiscence was found at one side, the defect was repaired with underlay technique using a new polyester/cotton fabric after removal of the old one.

All animals showed normal growth rate during the 12 months follow up and three ewes became pregnant and delivered normally without complications or hernia recurrence.

### Table 2: Animals grading according to wound healing post-surgery

<table>
<thead>
<tr>
<th>Grading</th>
<th>Umbilical hernia</th>
<th>Abdominal hernia</th>
<th>Total %</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Calf</td>
<td>Buffalo calf</td>
<td>Buffalo calf</td>
</tr>
<tr>
<td>Grade 1</td>
<td>14</td>
<td>-</td>
<td>2</td>
</tr>
<tr>
<td>Grade 2</td>
<td>1</td>
<td>1</td>
<td>-</td>
</tr>
<tr>
<td>Grade 3</td>
<td>1</td>
<td>-</td>
<td>1</td>
</tr>
</tbody>
</table>

Grade 1: Normal healing without complication, Grade 2: Wound infection and delayed healing, and Grade 3: Implantation failure and hernia recurrence

### Discussion

The good results achieved with the different prosthesis available and the reduction of the recurrence rate from up to 50% to less than 24% after repair of abdominal defects has placed prosthetic materials at the forefront with respect to the conventional suture repair techniques (Kingsnorth, 2000). However, in the developing countries, where veterinary surgery is considered of low public priority due to the usual comparison between animal value and the proposed cost of operation, the search for an ideal artificial biomaterial substitute to be used as a graft for large sized acquired or congenital hernias is a challenge. In this study a sterilized commercial polyester/cotton fabric was proven to be good hernioprosthetic material owing to its availability, flexibility, tissue compatibility and its unique cost effectiveness as a size of 1 m² of this...
material costs less than 1 American Dollar in comparison with other highly priced prosthetic materials.

Refreshment of the fibrosed edges of the hernial rings before implantation of the prosthetic material was applied to facilitate a better mesh incorporation with the host tissue. Similar advice was given by Attinger et al. (2000) who reported that removal of the hernial ring induced better mesh incorporation and stimulated wound healing in comparison with suturing the mesh to undisturbed hernial ring.

The technique of implantation of the prosthetic materials is thought to play an important role in hernia recurrence (Schumpelick et al., 2004). In the present study, prosthetic herniorrhaphy was carried out using the underlay technique with omentopexy that revealed satisfactory results in terms of low hernia recurrence (2 cases among 24 cases). Fixation of the implant by interrupted sutures in the underlay technique provided a multiple point non tension fixation which helped in dividing the stress evenly over the mesh and reduced crimping and bulging of the mesh (Parulkker et al., 1986; Ladurner et al., 2001). Also, the position of the implant behind the rectus muscles where the force of abdominal pressure hold the prosthesis against the deep surface of the abdominal muscle wall is another explanation obtained by Reilingh et al. (2004).

The type of suture material used for fixation of the fabric also has a great role in the success of the prosthetic repair. The use of non-absorbable synthetic monofilament suture (polypropylene) has high strength and low tendency to harbor microorganisms was endorsed by Kawcak and Stashak (1995).

Omentopexy was performed to decrease the direct contact between the mesh and the abdominal viscera and the subsequent inflammatory response to the prosthesis that gives rise to intra-abdominal adhesions. Similar recommendation was given by Aydinli et al. (2007) for interpositioning of the omentum to act as a physical barrier between the implant and the visera.

Covering the implanted material with a supportive flap from the hernial sac was found to strengthen the abdominal wall closure and reduce postoperative complications attributed to excessive suture tension and seroma formation. Similar advice was given by Scott (1979) in repairing of incisional hernia in horses.

Seroma formation was a major problem with the non-perforated Surgisis mesh repair (Gupta et al., 2006). Also, hernia recurrence was the major complication with alloderm mesh in addition to limited size which requires sheets to be joined together for large defects (Butler et al., 2005). Seroma formation was reported in all animals after implantation of the polyester/cotton fabric. Such complication was attributed by Amid (1997) to the size of dead space created between the implant and the host tissues, local circulatory disturbance resulting from the tight suture and the degree of haemostasis. It is necessary to avoid the contact of mesh with bowel to prevent late complications following prosthetic mesh repair (Basoglu et al., 2004).

Generally, all prosthetic materials are associated with post-repair clinical complications as wound infection, bowel fistula, erosion into abdominal viscera, increased recurrence rate (25%), repair failure and mesh extrusion even with polypropylene mesh that was universally accepted, where its implantation was associated with a high rate of adhesion formation to abdominal viscera especially small bowel (Bellon et al., 1996; Coda et al., 2003; Kingsnorth and LeBlanc, 2003).

The post-operative complications occurred in this study were within the acceptable limit. Wound infection and delayed healing were recorded in 3 animals in which the cause of hernia was recurrence after fault operation. Interestingly, the infection was cured with topical dressing without the need for mesh removal, which confirms the previous observation by Vilar et al. (2009) that it was not always necessary to remove the mesh in presence of infection.

Two cases of hernioplasty failure (2 out of 24, i.e., 8.3%) were recorded in this study. The cause of failure was wound infection in one calf with ratio (4.1%) that was considered an encouraging result in comparison with Hock et al. (1989) who previously reported that the rate of incisional hernia recurrence due to infection, ranged from 15 to 45% and explained that these procedures should be considered contaminated for surveillance and reporting purposes. The second case of hernia recurrence was recorded in a ewe following parturition which may be attributed to abdominal distension due to pregnancy and to the violent straining during parturition which lead to recurrent herniation at the same site of the previously implanted patch. Similar findings were recorded by Kader et al. (2005) despite the use of two folds of implanted patch.

In conclusion, the promising results obtained from the clinical use of commercial polyester/cotton fabric (Damour) in terms of body tolerance, availability and its economic effectiveness has encouraged us to potentiate its use for hernioplasty of large sized hernias in calves, sheep and goat.

References


Coda, A; Bendavid, R and Botto, F (2003). Structural alteration of prosthetic meshes in humans. Hernia. 7: 29-34.


Parulker, BG; Mathur, SK; Sup, AN and Vora, IM (1986). Use of indigenous knitted nylon mesh to repair large abdominal defects—an experimental study. JPGM. 32: 214-218.


مآذن سنتی جدید برای هریبولاوئیس در نشخوارکنگدان

محمدرضا اسماعیلی مربی و خالد صلاح احمد ایوبنصر
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هدف از این کار مطالعه اکتشافاتی استفاده از یک پارچه بیلی استرکتان تجاری برای ترمیم فتق‌های پنجره نافی و شکمی در گوساله‌ها و نشخوارکنگدان کوچک بود. 24 حیوان (16 گوساله، 8 نر و 8 گوساله) یک فتق نافی یا شکمی وارد این مطالعه شدند. قطر حلقه فتق از 12 تا 25 سانتی‌متر متغیر بود. از تکنیک زیر لایه‌ای برای نگهداری حیوان مورد نظر مصرفی استفاده شد که در پرینتویم احتمالی بعد از استنپیکس قرار داده و به بافت گیرنده با الگوی غیر منتظم روي هم افتاده با استفاده از زخم‌های نافی به رجوع به دسته‌بندی پلی‌پروپیلن محکم کردید. بعد از ایک سال، 30 حیوان نمونه در 2 مورف (80٪) مراحل کریز کردند. در نتیجه، نتایج مثبت پیشنهاد نموده که بهتر با استریکتان تجاری (دامور) پک جایگزین مقرن به صرفه‌سازی جدید برای مشاوره متداول مورد استفاده را چهت هریبولاوئیس فتق‌های پنجره نافی و شکمی در گوساله‌ها و نشخوارکنگدان کوچک ارائه می‌دهد.

وژه‌های کلیدی: هریبولاوئیس، بیلی استرکتان، گوساله، نشخوارکنگدان کوچک

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Torque teno midi virus/Small anellovirus (TTMDV/SAV)

مقاله کوتاه: جستجوی در مرگان اهلی روستایی و انتقال عمودی آن از مرغ به تخم مرغ

محمدرضا اسماعیلی و شریف سلمانی‌زاده

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دریافت مقاله: 20 آذر 1392، پذیرش نهایی: 18 شهریور 1393

گرچه عفونت حیوانات و پرندگان‌های غیر انسان با پیش‌بینی دیگر خانواده‌ای گوشی‌وریز دیگری است، ولی گزارش‌هایی در مورد عفونت با

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