کارگاه‌های آموزشی مرکز اطلاعات علمی

مقاله نویسی علوم انسانی

اصول تنظیم قراردادها

آموزش مهارت های کاربردی در تدوین و چاپ مقاله
Expression of acrA and acrB Genes in Escherichia coli Mutants with or without marR or acrR Mutations

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Abstract

Objective(s): The major antibiotic efflux pump of Escherichia coli is AcrAB-TolC. The first part of the pump, AcrAB, is encoded by acrAB operon. The expression of this operon can be kept elevated by overexpression of an activator, MarA following inactivation of MarR and AcrR repressors due to mutation in encoding genes, marR and acrR, respectively. The aims of this research were to use E. coli mutants with or without mutation in marR or acrR to search for the presence of possible mutation in acrR and to quantify the expression of acrAB.

Materials and Methods: The DNA binding region of acrR gene in these mutants were amplified by PCR and sequenced. The relative expression of acrA and acrB were determined by real time PCR.

Results: Results showed that W26 and C14 had the same mutation in acrR, but none of the mutants overexpressed acrA and acrB in comparison with wild type strain.

Conclusions: The effect of marR or acrR mutation on acrAB overexpression is dependent on levels of resistance to tetracycline and ciprofloxacin.

Introduction

Generation of multiple drug resistant phenotypes of pathogenic bacteria, like Escherichia coli is a worldwide clinical concern. These phenotypes are associated with increase in the activity of membrane transporters mainly AcrAB-TolC, which belongs to the resistance-nodulation-division (RND) family of transporters (1). This transporter or efflux pump consists of three ingredients, including AcrA, a periplasmic membrane-fusion protein; AcrB, the inner membrane protein; and TolC, an outer membrane channel. These ingredients are encoded by acrA, acrB and tolC. The first two genes are located in the same operon, while tolC is placed on different site of bacterial chromosome (2, 3).

AcrR, the repressor of acrAB operon is encoded by acrR gene (4). Its location is upstream of acrAB operon and transcribed divergently from the same promoter (Figure 1). Attachment of AcrR through its DNA-binding helix-turn-helix (HTH) motif to operator site of acrAB operon causes operon repression (6). On the other hand, this operon is under the positive regulation of MarA, a transcriptional activator (7). Its binding site is shown in Figure 1.

Expression of MarA happens following dissociation of MarR repressor from operator site of marRAB operon (8). Both AcrR and MarR repressors possess DNA binding and drug binding sites (6, 9). In separate studies, it was shown that mutation in their encoding genes, marR and acrR, can maintain the overactivity of the AcrAB-TolC pump (10, 11). It was found that two clones C14 (without a mutation in marR) and C16 (with a mutation in marR) slightly overexpressed marA (12, 13). This in turn may promote overexpression of acrAB operon. The aims of this research were first, to study the possible presence of mutations in acrR gene and then to quantify the expression of acrA and acrB in these clones.

Materials and Methods

Antimicrobial agent and media

The stock of 4 mg/ml tetracycline hydrochloride (Tc) (Sigma) was used in this research. LB broth (Merck, Germany) and LBA containing 1.5% agar (Merck, Germany) were used for cultivation of control strain and mutants.
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Bacterial strain and mutants
Bacterial strain and mutants are listed and described in Table 1. MG1655 was the wild type strain. W26 and W49 are mutants isolated from cultivation of wild type strain on LBA plus 40 ng/ml ciprofloxacin (14). Clones C14 and C16 were generated during the previous work (15). They were derived from mutants W26 and W49 following cultivation on LBA agar containing up to 20 µg/ml Tc (15). Based on the previous data, these clones and mutants show low to medium levels of resistance to ciprofloxacin and tetracycline (16, 17).

PCR amplification and DNA sequencing of acrR
PCR was used to amplify the 5′ end of acrR gene in wild type and mutants (14). A single colony from each mutant and clone on LB agar was suspended in 100 µl of sterile water and after boiling at 95°C for 3 min; it was cooled on ice and used as a PCR template for acrR amplification. Forward and reverse primers for amplification were 5′-CACGAACATATGGCACG-3′ and 5′-GCCTGATACTCAAGCTC-3′, respectively. The amplified PCR products were 240 bp. The sequence of these products was compared with that of MG1655 obtained from NCBI (NC_000913.2) following DNA sequencing.

acrA and acrB expression analysis by real time PCR
A fresh culture of bacteria was prepared in LB broth plus 3 µg/ml Tc (except for the wild type) and incubated at 37°C with shaking at 150 rpm and grown to mid-logarithmic phase. Each culture was used for extraction of RNA using an RNeasy Mini Kit (Qiagen, Germany) following stabilization in RNA protect bacterial reagent (Qiagen, Germany). RNase-free DNase I was used to eliminate contaminating genomic DNA according to the manufacturer’s instruction (Fermentas, Life science research) and the absence of DNA was confirmed by amplification of RNA samples plus a DNA sample as a positive control. The concentration of total RNA was estimated at OD 260 using spectrophotometer (Ultrospec 1100, Amersham Pharmacia BioTech).

Purified total RNA (2 µg) was used as a template in RT-PCR using a RevertAid Reverse Transcriptase kit (Fermentas, Life science research). The cDNAs obtained from reverse transcription were used to quantify the level of acrA, acrB and gapA, as an endogenous reference gene by real time PCR in a Rotor Gene 6000 thermocycler (Corbett Research, Australia) using a SYBR Green kit (Takara, Japan). The specific primers used for real time PCR are listed in Table 2. Thermal cycling conditions were described previously (3). Relative gene expression was calculated using the efficiency method Pfaffl (ratio of target gene expression, acrA and acrB, to gapA expression) (18). All data on gene expression are the mean of triplicate analyses. The data were presented as mean±SD. Statistical analysis of relative expression was done by SPSS version 16. T-test was used for comparison of relative gene expression data.

Results
Mutants were used to be evaluated for the presence of possible mutation in 5′ end of acrR gene corresponding to HTH motif of encoded protein and

Table 1. Bacterial strain and mutants

<table>
<thead>
<tr>
<th>Strain/Mutant /Clone</th>
<th>Relevant properties</th>
<th>MIC Ciprofloxacin (ng/ml)</th>
<th>MIC Tetracycline (µg/ml)</th>
<th>Source/Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>MG1655</td>
<td>Wild type</td>
<td>35</td>
<td>3</td>
<td>A gift from Prof. R. G. Lloyd</td>
</tr>
<tr>
<td>W26</td>
<td>Wild type; gyrA (Ser&lt;sub&gt;83&lt;/sub&gt;→Leu)</td>
<td>75</td>
<td>4</td>
<td>Pourahmad &amp; Mohiti, 2010</td>
</tr>
<tr>
<td>W49</td>
<td>Wild type; gyrA and marOR (20 bp duplication in operator)</td>
<td>625</td>
<td>4</td>
<td>Pourahmad &amp; Mohiti, 2010</td>
</tr>
<tr>
<td>C14</td>
<td>W26; gyrA (Ser&lt;sub&gt;83&lt;/sub&gt;→Leu)</td>
<td>1000</td>
<td>30</td>
<td>Pourahmad &amp; Ebadi, 2013</td>
</tr>
<tr>
<td>C16</td>
<td>W49; gyrA and marOR (20 bp duplication in operator)</td>
<td>1000</td>
<td>30</td>
<td>Pourahmad &amp; Ebadi, 2013</td>
</tr>
</tbody>
</table>

Figure 1. acrAB operon, acrR and the regulatory region between them. Modified and adapted from Dzwokai et al (12)
Expression of acrA and acrB in E. coli Mutants
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Table 2. List of real time PCR primers

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequence (5′-3′)</th>
<th>Length of amplicon (bp)</th>
<th>Reference</th>
</tr>
</thead>
</table>
| acrA | F: TTGAAATTACGCTTCAGGAT  
      | R: CTTAGCCCTAAGGATGTG | 189        | Viveiros et al, 2007 |
| acrB | F: CGTACACAGAAAGTGCCTAA  
      | R: GGCCTCAACTTTGTTTTTCTT | 183       | Viveiros et al, 2007 |
| gapA | F: ACTTACGAGCAGATCAAAGC  
      | R: AGTTTCACGAAGTTGTCGTG | 170       | Viveiros et al, 2007 |

Figure 2. PCR products of acrR gene in wild type (wt) and mutants. First lane shows the 1 Kb ladder and other lanes show PCR products.

Figure 3. Sequence output from acrR PCR product of C14 mutant (first part) and wild type (second part) using forward and reverse primers. Underlined nucleotides show the differences between nucleotide sequences of two parts.

Discussion

The increased level of resistance to fluoroquinolones, such as ciprofloxacin and other structurally unrelated antibiotics, like tetracycline, which causes multiple resistance phenotypes is attributed to over activation of multidrug efflux pumps, mainly AcrAB-TolC pump in E. coli (1-3). Overactivity of this pump was seen even in other bacteria following the induction with increasing amounts of tetracycline or acquiring high levels of resistance (19).

Generally, fluoroquinolone resistance has been attributed to point mutations in the quinolone resistance-determining regions of the target genes, such as gyrA (20). However, higher levels of resistance can be achieved following overactivation of...
AcrAB-ToIC pump (21). This happens following the overexpression of marA and thereby overactivation of acrAB and toIC genes. The expression of these genes has been determined in clinical isolates by real time PCR (22-24). Therefore, it was decided to quantify the expression of acrA and acrB genes in mutants and clones with or without a mutation in marR after evaluation of the possibility of acquiring mutation in acrR.

It was found that none of the mutants overexpresses acrA and acrB following the addition of 3 μg/ml Tc. This may be due to the levels of resistance to antibiotics in mutants and clones. This is also possible that the level of marA overexpression in clones was not enough to overexpress acrAB. This possibility arises following the suggestion that the level of marA overexpression is important for activation of acrAB operon (25).

It was shown that Arg at position 45 of AcrR is highly conserved and its alteration to cysteine enhances the expression of acrB in mutants with high levels of resistance to ciprofloxacin (10). In the present work, it was found that W26 and its derived clone C14 had alteration at position 45. However, this alteration did not promote overexpression of acrAB. This may imply that mutation at this location is not the only cause of acrAB overexpression. This is consistent with the previous findings indicating that in stress conditions, expression of acrAB enhances independent of AcrR activity. However, after overexpression of acrAB, the presence of active AcrR is important to regulate the levels of acrAB expression (26).

Moreover, the importance of the repressor binding site on DNA, and repressor DNA binding motif for MarR and AcrR repressor were mentioned previously as mutations in these locations along with overactivity of MarA promote overexpression of acrAB operon (10, 11). The finding that mutants and clones harboring mutations in either of these locations in marR or acrR could not promote overexpression of acrA and acrB, again reveals that the level of resistance is important for overexpression of acrAB operon.

In addition, it was shown that tolerance of organic solvents, such as cyclohexane happens following overexpression of acrAB-toIC (27). It was previously found that none of these mutants and clones could tolerate cyclohexane (13, 15). Thus, the findings of this study reconfirm the relation between organicsolvent tolerance and the level of acrAB expression.

## Table 3. Relative expression of acrA and acrB in wild type (MG1655) and mutants as determined by real time PCR

<table>
<thead>
<tr>
<th>Strain/mutant</th>
<th>Relative expression of acrA</th>
<th>Relative expression of acrB</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild type (MG1655)</td>
<td>1±0</td>
<td>1±0</td>
</tr>
<tr>
<td>W26</td>
<td>1.16±0.021</td>
<td>1.13±0.012</td>
</tr>
<tr>
<td>W49</td>
<td>1.62±0.01</td>
<td>1.45±0.015</td>
</tr>
<tr>
<td>C14</td>
<td>1.28±0.013</td>
<td>1.17±0.011</td>
</tr>
<tr>
<td>C16</td>
<td>1.4±0.013</td>
<td>1.31±0.02</td>
</tr>
</tbody>
</table>

## Conclusion

Upregulation of acrAB operon occurs after acquisition of high levels of resistance to Tc and ciprofloxacin. Thus, the effect of marR or acrR mutation on acrAB overexpression is dependent on the levels of resistance to these antibiotics. At high levels of resistance, evaluation of the synthesis of AcrAB-ToIC pump ingredients along with mRNA quantification by real time PCR would reconfirm AcrAB-ToIC overactivity.

## Acknowledgment

This work was financially supported by the University of Shahrekord, Iran. We thank Prof R G Lloyd for his kind gift of MG1655.

## References

10. Webber MA, Talukder A, Piddock LJ V. Contribution of mutation at amino acid 45 of AcrR to acrB expression and ciprofloxacin resistance in clinical and veterinary
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