Abstract
Botulinum neurotoxins constitute a family of bacterial toxins for botulism syndrome in human. The toxins bind with high affinity to nerve cells where they cause a complete inhibition and release of neurotransmitters and thereby produce flaccid paralysis. In this study the binding domain of type E neurotoxin was isolated by PCR and expressed in a proper expression vector. The results of this investigation can be used as a tool to study the mechanism of binding of holotoxins. This study is also implicated in antibody production against botulism syndrome.
Keywords: Botulinum neurotoxin type E, Binding domain, Expression

INTRODUCTION

The botulinum neurotoxins (BoNTs) are the causative agents of botulism and represent a family of seven structurally similar but antigenically distinct serotypes (A to G) (Éric and Marite, 2001; Humeau et al., 2000; Pellizzari et al., 1999 and Chaddock et al., 2002). These toxins exert their action by blocking the release of the neurotransmitter acetylcholine at the neuromuscular junction (Lalli et al., 2003 and Turton et al., 2002). BoNTs are usually expressed in Clostridium botulinum as a single polypeptide chain and then post-translationally nicked, forming a di-chain consisting of a 100 kDa heavy chain and a 50 kDa light chain held together by a single disulfide bond (Éric and Marite, 2001; Rossetto et al., 2001; Zhou and Singh, 2004; Jensen et al., 2003 and Agarwal et al., 2004). Topologically, these neurotoxins are composed of three domains, a binding domain, a translocation domain, and a catalytic domain, each of which is believed to play a role in intoxication (Lalli et al., 2003; Lalli et al., 1999; Zhou and Singh, 2004 and Jensen et al., 2003). The carboxyl-terminal portion of the heavy chain is responsible for binding to nerve cell receptor(s). After toxin binding, it is thought to be internalized into an endosome through receptor-mediated endocytosis. It is believed that the 50 kDa amino-terminal domain of the heavy chain possesses channel-forming capabilities in the acidic environment of the endosome, allowing internalization of the toxin. The final step in the mechanism involves zinc-dependent proteolysis by the catalytic domain of key cytosolic substrates necessary for neurotransmitter release (Humeau et al., 2000; Pellizzari et al., 1999 and Turton et al., 2002). The Clostridium botulinum neurotoxins specifically cleave a family of proteins, named SNAREs (soluble NSF attachment protein receptor) (Kotich et al., 2002 and Rickman et al., 2004). These proteins are involved in multiple steps leading to the docking and fusion of small synaptic vesicles with the presynaptic plasma membrane and in a variety of other intracellular trafficking events. BoNT/E instead, cleaves Arg180-Ile181 peptide bond of the SNAP-25 (synaptosomal-associated protein of 25 kDa) (Schiavo et al., 1998 and Pellizzari et al., 1999). The binding domain of BoNTs has been one of the most successful and frequently used tools in neurobiology and cell biology. Recombinant heavy chain C-terminal (HCc) fragment of BoNT/E antagonised the action of parental Clostridium botulinum neurotoxins, which is seen as a delay in the onset of paralysis (Lalli et al.,...
Mousavi et al. 184

MATERIALS AND METHODS

Enzyme and chemicals: T4 DNA ligase and Pfu DNA polymerase were from Fermentase. All other chemical reagents were obtained from Sigma (USA) and LaRoche (Germany). Miniprep kit for purification of DNA was from Qiagen (USA). Primers were synthesis by MWG Biotech Company (Germany). Specific antibody against Clostridium botulinum type E neurotoxin was the product of Medep Co. (Russia).

Bacterial strain and plasmids: Plasmid pUC18 was purchased from Pharmacia and PET32a was the product of Novagen (USA). The Plasmid pTZ57R obtained from Fermentase. Escherichia coli BL21 DE3 and DH5α were from Cinagene (Iran).

Microbial cultures: E. coli strain was grown in LB broth and Clostridium botulinum type E was grown in cooked meat (Difco, USA).

Bacterial growth and isolation of DNA: Clostridium botulinum type E cells were cultured anaerobically under 15% CO₂ at 35°C for 24-48 h in cooked meat medium Chromosomal DNA was prepared with alkaline method.

PCR amplification and cloning: A pair of primer 5’ ATAC GGAATT C AT AAT AGT ATT CCT TTT AAG C 3’ which adds a BamH1 site to the 5’ end and 5’ TCTA GTGCAC TTA TTT TTC TTG CCA TCC 3’ which adds a Sal1 restriction site to the 5’ end were designed from the published binding domain of BoNT/E (accession # x62683) to isolate this domain as residue 830-1251. PCR was performed in 50 µl total volume containing Pfu DNA polymerase (2 units) and its buffer, 0.2 mM of each dNTP, 20 pmol of both primers, and 20 µg DNA. Samples were preheated for 5 min at 95°C and then 30 cycles of PCR were performed: 1 min at 94°C, 30 s at 60°C, and 1.5 min at 74°C. After the last cycle, was continued for additional time of 7 min at 74°C. The size of amplified DNA fragment as monitored on 1.0% agarose gel was ~1.3 kb. The PCR product was isolated from a low melting agarose gel and purified with gel extraction kit and digested with BamH1 and Sal1. The digested PCR product was ligated into a pTZ57R previously digested with the same enzyme and transferred into DH5α host. One recombinant plasmid containing the expected size insert was sequenced by the dideoxy chain termination method. The recombinant plasmid was cleaved with SalI and BamHI, the SalI–BamHI fragment was isolated and recovered from the low melting agarose gel, and then ligated to the expression vector pET-32a. Clones were screened by digestion of a miniprep plasmid DNA with SalI and BamHI.

Expression and Purification: Five milliliters of LB Broth medium containing 50 mg/ml ampicillin was inoculated with 10 µl of the freezer stock of BL21 (DE3) cells with pET-BoNT/E binding domain. Following an overnight growth at 37°C, this culture was used to inoculate 100 ml of LB ampicillin medium in a 500 ml flask and the cells were grown at 37°C with shaking until the A₆₀₀ reached 0.6. At this point, 1mM IPTG was added to induce BoNT/E-HC expression. The cells were then incubated at 30°C for an additional 12h. The induced cells were harvested by centrifugation at 5000 rpm at 4°C for 10 min. The cell was resuspended in 4 ml lysis buffer (50 mM Na–phosphate, pH 8.0, 300 mM NaCl, 5 mM benzamidine and 0.5 mM PMSF) supplemented with protease inhibitor and lysozyme. The bacterial suspension was incubated at room temperature for 30 min to lyse cells. After the cells were completely lysed, 5 µg/ml deoxyribonuclease-I was added to digest any associated DNA. This step helps reducing the viscosity of the extract. After an additional 10 min, the lysate was centrifuged at 16000 rpm for 30 min to remove the insoluble cell debris. Each of the culture samples was tested for the expression of protein on sodium dodecyl sulfate–polyacrylamide gel (10%) electrophoresis (SDS–PAGE). The supernatant obtained from the above step was allowed to mix with 5 ml Ni–NTA.
agarose (nickel-nitrilotriacetic acid), pre-equilibrated with phosphate buffer (50 mM Na–phosphate, pH 8.0, 300 mM NaCl), at 4°C for an hour. The mixture of Ni–NTA agarose and supernatant was poured into the glass column and the flow-through of the soluble fraction was collected. The column was washed with 10 ml of phosphate buffer followed by 5 washes with buffer containing 50 mM imidazole. Tightly bound protein was eluted with 2 ml of 100 mM and 3 ml of 200 mM imidazole. Aliquots of all the above fractions were analyzed by electrophoresis on 12% SDS–PAGE gel followed by staining with Coomassie blue. A 71 kDa band corresponding to BoNT/E-HCc reproducibly eluted in 100–200 mM imidazole fractions was obtained. In order to separate the fusion protein from HCc chain the recombinant protein was treated with restriction protease enterokinase. The reaction was carried out as described in the manufacturer’s (Roche, Germany) manual. The protein was further purified employing Ni–NTA agarose column.

Western blotting analysis: The recombinant protein was detected by Western blotting using horse anti-clostridium botulinum toxin type E. The recombinant BoNT/E HCc was transferred to nitrocellulose membrane using Bio-Rad Mini Protean II System and transfer buffer (39 mM glycine, 48 mM Tris–base, 0.037% SDS, and 20% methanol). The membrane was incubated in the blocking buffer of 3% bovine serum albumin (BSA)/ phosphate-buffered saline (PBS, 137 mM NaCl, 2.7 mM KCl, and 4.3 mM Na2HPO4·7 H2O, pH 7.3), with gentle shaking for 1 h at room temperature. After decanting and discarding the blocking buffer, the membrane was incubated in a 1:3000 dilution of horse anti-clostridium botulinum toxin type E in the PBST (PBS containing 0.05% Tween), with gentle shaking for 1 h at RT. After washing the membrane with PBST for three times, each time for 5 min, blots incubated with a 1:1000 dilution of the polyclonal goat anti-horse

RESULTS

A DNA fragment encoding the binding domain of BoNT/E was amplified by PCR using sense and anti-sense primers. Specific restriction sites, BamHI and SalI, for unidirectional cloning and a stop codon were introduced into the amplified gene. Pfu DNA polymerase with proofreading was used for amplified gene (Fig. 1). The restriction map of the binding domain gene with EcoRI and BclI derived by software analysis showed three fragments of 811, 385 and 187 and two fragments of 978 and 324 base pairs which were produced on digestion with EcoRI and BclI respectively. The amplified fragment was thereby confirmed to be the correct one. The isolated DNA from low melting agarose was ligated into BamHI, SalI-cleaved pTZ57R vector. Restriction digestion and subsequent agarose gel electrophoresis of the clone indicated the presence of a 1302 bp fragment (Fig. 2). In an attempt
to get a higher yield, the DNA fragment cloned into pTZ57R, was excised with SalI and BamHI, and then ligated into pET-32a which is a bacteriophage T7-RNA polymerase promoter vector system. The T7-RNA polymerase is capable of transcribing almost any DNA linked to a T7 promoter (Studier et al., 1990). The pET-BoNT/E-HCc recombinant plasmid was digested with NdeI and BamH I or SalI, or BamH I only, to confirm the ligation. The recombinant plasmids were also confirmed by PCR. DNA sequence analysis confirmed its correct orientation when compared to the sequence of binding domain. The pET-BoNT/E-HCc was transformed into E. coli BL21. His6-tagged recombinant BoNT/E-HCc was found to be overproduced using IPTG as an inducer. A band corresponding to a 71 kDa His 6-tagged BoNT/E-HCc was observed in SDS–PAGE of total lysate of E. coli BL21 (pET-BoNT/E-HCc) culture after induction with IPTG (Fig. 3). The soluble fraction of the protein was allowed to selectively bind to Ni–NTA agarose through the His-tag. The protein was then eluted by ascending concentrations of imidazole. The protein was eluted largely at 200 mM imidazole concentrations (Fig. 4). The 71 kDa purified recombinant protein was digested with enterokinase as a result of which a 50 kDa HCc and 20 kDa fusion protein could be identified (Fig. 5). The protein identity was verified by Western blot analysis (Fig. 6). The assay revealed that the recombinant BoNT/E-HCc was strongly and specifically recognized by anti-clostridium botulinum toxin type E., Whereas, no reactivity was observed in controls.

**DISCUSSION**

The *Clostridium botulinum* toxins (type A-G) consist of two protein chains joined by a disulphide bridge. The light chain (50 kDa) is responsible for the intracellular catalytic (toxic) activity and heavy chain (100 kDa) for the binding and internalization of the toxin into the neuron. In particular it is the 50 kDa C-terminal domain of the heavy chain which is responsible for the binding of the toxin to neuronal membrane prior to toxin internalization. Recombinant protein of this construct has potent antagonists of the corresponding clostridium neurotoxins (Lalli et al., 1999), causing a significant delay in the onset of paralysis more than 150%, while percent for binding domain of BoNT/A and TeNT is lower than 75%. Therefore the data obtained indicates that the binding domain can be cloned and expressed in E. coli and recombinant protein can be considered for production of antibody. Recent works (Smith, 1998 and Byrne and Smith, 2000) have also shown that binding domain gene derived from botulinum toxins A and B can be exploited as vaccines against their respective toxin subtype.
On the basis of the structural similarity between these toxins and botulinum toxin E we investigated the possibility of exploiting carboxy-terminal fragment from the heavy chain of botulinum E toxin as vaccine. Although toxoid vaccines are available, there are numerous shortcomings with the current use and ease of production. Development of a new generation, recombinant vaccine could alleviate many of the problems associated with the toxoid. A recombinant vaccine would eliminate the need for a dedicated manufacturing facility. Presently, many current Good Manufacturing Practices (cGMP) facilities are in existence and available that could manufacture a recombinant product. There would be no need to culture large quantities of a hazardous toxin producing bacterium. Production yields from a genetically engineered product are expected to be high. Presumably, there would be no need to treat the vaccine with formalin if the recombinant vaccine candidate represented a fragment of the toxin. A fragment would not possess all three functional domains (i.e., binding, internalization, catalytic) which are all required for its mechanism of action. Recombinant products would be purer, less reactogenic, and more fully characterized. Human immunization with toxoid resulted in production of antibodies directed largely against the toxin light chain, with fewer antibodies binding heavy chain. Since antibody neutralization activity results largely from blockade of cellular receptor binding by heavy chain, so the heavy chain vaccine will be more protective than a toxin based vaccine, as more heavy chain antibodies are generated (Amersdorfer et al., 2002). Thus, the cost of a recombinant product would be expected to be much lower than a toxoid because there would be no expenditures required to support a dedicated facility, and the higher production yields would reduce the cost of the vaccine product.

References


