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Synthesis and Biological Investigation of some Novel Sulfonamide and Amide Derivatives Containing Coumarin Moieties

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Abstract

New sulfonamide and amide derivatives containing coumarin moieties; oxo-2H-chromen-sulfamoylphenylacetamides and oxo-2H-chromen-arylacetamides were synthesized starting from diverse 2-chloroacetamide derivatives and a wide range of coumarins. The structures of compounds were elucidated by IR and NMR spectra and also analytical elemental analysis. In the next step, the above mentioned compounds were screened for their antimicrobial and antioxidant activities.

Their antimicrobial activity was assigned using the conventional agar dilution method and the antioxidant activity was assessed using two methods, 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging method and ferric reducing antioxidant power (FRAP) assay. Although the compounds showed no remarkable antimicrobial activities, most of them exhibited good antioxidant activities. Compounds 5b showed the most potent DPPH activity, whereas 8c was the most efficient compound in FRAP assay.

Keywords: Sulfonamides; Amides; Coumarins; Antioxidant activity; Antimicrobial.

Introduction

Sulfonamide and amide derivatives comprise an important class of drugs with diverse biological applications. Sulfonamides are widely used as antimicrobial (1, 2), anticancer (3, 4), anti-inflammatory (5) and antiviral agents as well as HIV protease inhibitors (6). Also some derivatives are well recognized as an antimetabolite (7) and in another study new sulfonamides have shown good cytotoxic effects against breast cancer cells (8). Sulfonamides

were the first effective chemotherapeutic agents to be utilized efficiently to prevent and cure the bacterial infection in human beings (9-12). Recently some sulfonamide derivatives were screened for their antioxidant activity; the property which has not been investigated widely, and satisfactory results were obtained (13).

Amides are one of the main building blocks in Nature (14) and have important biological activities (15, 16). Also they have been reported as antibiotic (17) and antimicrobial agents (18). Recent studies shows that novel designed amides which were active as novel orally available fatty acid amide hydrolase (FAAH) inhibitors are therapeutically effective against

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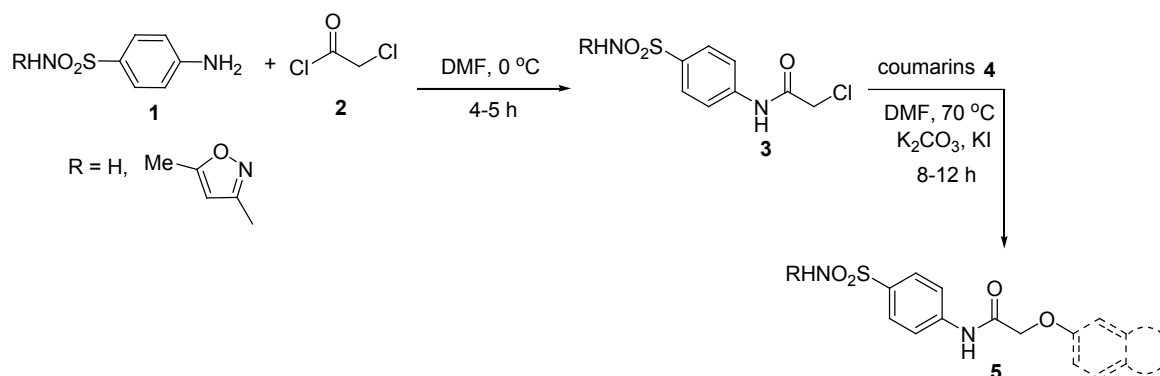


Figure 1. Synthesis of various sulfonamide derivatives 5.

pain (19). Antimalaria (20), antiproliferative (21) and cytotoxic activities (22) are other significant biological properties of amides. Valuable properties of sulfonamides and amides are true stimulation to present a large variety of these types of compounds and investigate their biological activity *in-vitro* and *in-vivo*.

In continuation of our research program on the synthesis of bioactive molecules, we have investigated sulfonamides (23), amides (24, 25), and also coumarin derivatives (26-28) as they possess various precious biological activities (29-31); in this report, we designed novel sulfonamide and amide derivatives containing coumarin moieties to profit both sulfonamide-coumarin/amide-coumarin properties (Figures 1 and 2).

Results and Discussion

Synthesis of sulfonamide derivatives 5 was started from sulfanilamide and its derivative, sulfamethoxazole 1 (Figure 1). They were reacted with chloroacetyl chloride 2, in DMF at 0 °C to obtain 2-chloroacetamide derivatives 3.

Then, we investigated reaction of 2-chloroacetamide derivatives 3 and various

coumarin derivatives 4 under different conditions. It should be noted that the best reaction medium to obtain the desired compounds 5 was dimethylformamide (DMF). Also we found that using potassium carbonate (K₂CO₃) along with potassium iodide (KI) led to the formation of 5 in higher yields.

It is worthwhile to mention that we prepared 3-chloroacetamide derivatives and examined their reaction with coumarin derivatives 4 under different conditions, no product or undesired products were obtained and further investigation was quitted.

In the next step, as similar to the procedures for the synthesis of sulfonamides 5, amide derivatives 8 were prepared starting from anilines 6, chloroacetyl chloride 2, and coumarin derivatives 4 (Figure 2).

The structure of coumarin derivatives which were utilized for the synthesis of compounds 5 and 8 have been depicted in Figure 3. They include 7-hydroxycoumarin 4a, 7-hydroxy-4-methylcoumarin 4b, 7-hydroxy-4-phenylcoumarin 4c, 4-hydroxycoumarin 4d, 4,7-dihydroxycoumarin 4e, 7,8-dihydroxy-4-methylcoumarin 4f, ethyl-7-hydroxycoumarin-

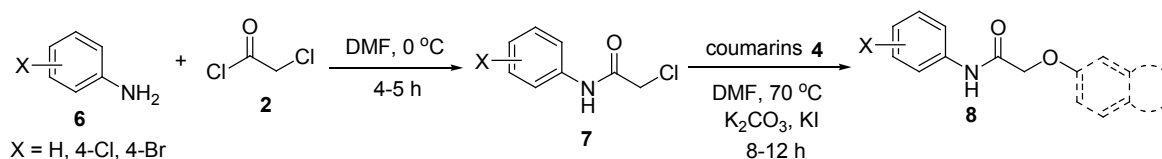
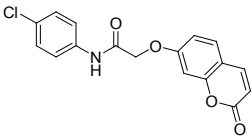
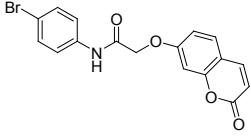
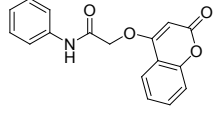
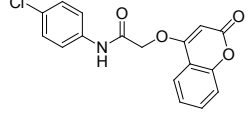
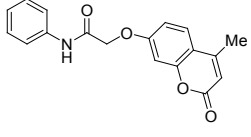
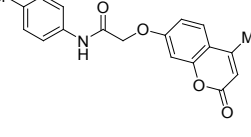
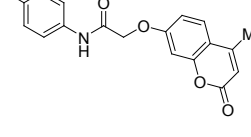
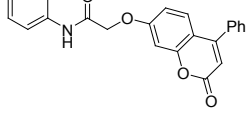
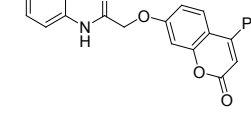


Figure 2. Synthesis of amide derivatives 8.

Table 1. Sulfonamide and amide derivatives and their antioxidant activities.

Entry	Structure	Product	DPPH radical scavenging activity (IC ₅₀ ^a , mM)	FRAP value (mM Fe ²⁺) ^b
1		5a	1.57 ± 0.01	4.7 ± 0.01
2		5b	0.66 ± 0.01	7.6 ± 0.02
3		5c	1.23 ± 0.02	4.2 ± 0.01
4		5d	0.95 ± 0.01	4.0 ± 0.01
5		5e	1.75 ± 0.02	4.5 ± 0.01
6		5f	1.15 ± 0.02	13.6 ± 0.03
7		5g	0.81 ± 0.01	13.5 ± 0.03
8		5h	1.08 ± 0.01	3.8 ± 0.01
9		8a	2.29 ± 0.03	2.7 ± 0.01

Table 1. (Continued).

10		8b	1.14 ± 0.01	2.9 ± 0.01
11		8c	0.94 ± 0.01	19.1 ± 0.04
12		8d	1.11 ± 0.03	17.5 ± 0.03
13		8e	1.23 ± 0.02	17.3 ± 0.03
14		8f	2.07 ± 0.02	15.4 ± 0.02
15		8g	2.08 ± 0.03	5.2 ± 0.01
16		8h	1.68 ± 0.01	8.0 ± 0.04
17		8i	1.86 ± 0.01	3.5 ± 0.01
18		8j	0.98 ± 0.01	5.1 ± 0.03
19	Quercetin		8.12 ± 0.13^a	87.1 ± 1.4

^aIn μM ^b100 μg of substances were used.

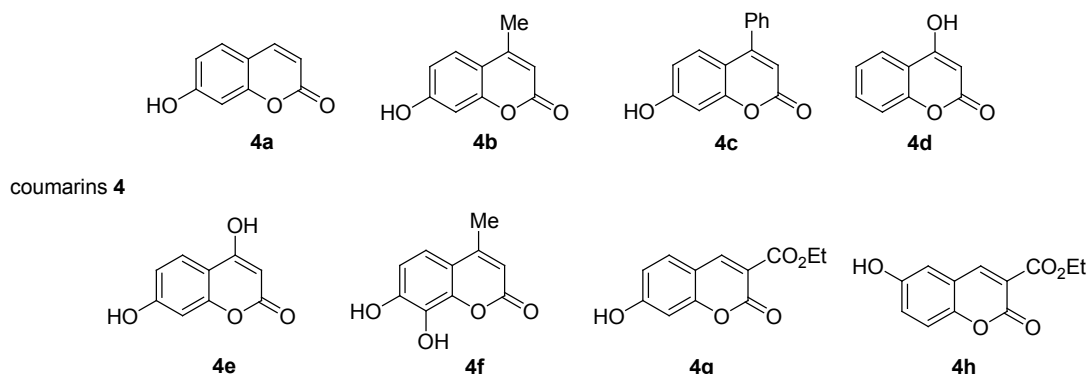


Figure 3. Coumarins used for the synthesis of sulfonamides **5** and amides **8**.

3-carboxylate **4g**, and ethyl-6-hydroxycoumarin-3-carboxylate **4h**. The corresponding products are shown in Table 1 (Entries 1-18).

Antimicrobial activity

Compounds **5a-h** and **8a-j** were evaluated for their antibacterial activity against Gram-positive (*Staphylococcus aureus* ATCC 6538p, *Staphylococcus epidermidis* ATCC 12228) and Gram-negative (*Escherichia coli* ATCC 8739, *Klebsiella pneumonia* ATCC 10031, and *Pseudomonas aeruginosa* ATCC 9027) bacteria using conventional agar-dilution method (32). The MIC (minimum inhibitory concentration) values were determined by comparison to ciprofloxacin as a reference drug. None of the compounds showed antimicrobial activities below 100 μM .

Antioxidant activity

In this study, antioxidant activity of compounds **5a-h** and **8a-j** were assessed using two different assay systems; DPPH (1,1-diphenyl-2-picrylhydrazyl) (33) and FRAP (ferric reducing antioxidant power) (34).

DPPH radical scavenging activity

The free radical scavenging activity of sulfonamide and amide derivatives **5a-h** and **8a-j** was evaluated by DPPH colorimetric method (33). Several dilutions of compounds were made and assayed to obtain concentration of the sample required to scavenge 50% (IC_{50}) of DPPH-free radical applying suitable regression analysis of the mean values. The results are given in Table 1.

All sulfonamide derivatives exhibited moderate radical scavenging activities (0.66 to 1.75 mM). The results indicate that various factors such as nature of substituents at position 3 or 4 and also position of hydroxyl group in the utilized coumarins **4** are very important and affect the related radical scavenging activity. Obviously presence of methyl group in the sulfonamide derivatives has significant effect since the best result is related to **5b** (0.66 mM) which possess methyl group at position 4. Also this effect is clear in compound **5g** (0.81 mM) which showed good activity, but introduction of two oxy-*N*-(4-sulfamoylphenyl)acetamide groups reduced the activity in comparison to **5b**. It should be noted that the presence of ethyl carboxylate group at position 3 diminished radical scavenging activity as well. Another noticeable factor is position of hydroxyl group in coumarins **4**. According to the results in Table 1, the product derived from 4-hydroxycoumarin **5c** (1.23 mM) is more active than the product derived from 7-hydroxycoumarin **5a** (1.57 mM). It should be noted that introduction of methoxazole ring in sulfonamide moiety, **5d** (0.95 mM) is effective and it has improved the activity of **5d** in comparison to **5a** (1.57 mM).

Similar results can be seen in the case of amides. Amides derived from 4-hydroxycoumarin **8d** (1.11 mM) and **8e** (1.23 mM) are more active than compounds derived from 7-hydroxycoumarin **8a** (2.29 mM) and **8b** (1.14 mM). Also compounds containing phenyl group at position 4, **8i** and **8j** show better activity

in comparison to those of having methyl group at the same position (8f-h). Another factor is related to the presence of halogen atom on the aromatic ring. We found that 4-bromophenyl enhanced the radical scavenging activity. In this manner, the best result is related to 8c (0.94 mM).

To sum up, considering the above mentioned factors, better radical scavenging activities are related to compounds 5b, 5g, 8c and 8j.

FRAP ability

The FRAP assay measures the ability of a compound to reduce the ferric 2,4,6-tripyridyl-s-triazine complex to the colored ferrous complex (34). FRAP values are obtained by comparing the absorbance change at 593 nm in test reaction mixtures with those containing ferrous ions in known concentration. The FRAP values for the tested compounds (Table 1) revealed that the synthesized compounds showed noticeable activities.

Among sulfonamides, 5f and 5g; compounds derived from 7-hydroxycoumarin and 7,8-dihydroxycoumarin, showed the best results (13.6 and 13.5 mM, respectively). Contrary to DPPH radical scavenging activities, compound derived from 7-hydroxycoumarin 5a (4.7 mM) exhibited better result in comparison to compound derived from 4-hydroxycoumarin 5c (4.2 mM) and also presence of ethyl carboxylate group at position 3 increased the activity; see related data for compounds 5e and 5f in Table 1. Similar to DPPH results, compounds containing methyl group at position 4, 5b and 5g showed satisfactory results (7.6 and 13.5 respectively).

As can be seen in Table 1, amides exhibited better FRAP values and similar to DPPH results amides containing methyl at position 4 and 4-bromophenyl in their structure showed very good results (8f and 8c; 15.4 and 19.1 mM, respectively). There are two significant data in Table 1, related to 8d and 8e with values of 17.5 and 17.3 mM, respectively. These compounds have been derived from 4-hydroxycoumarin and are more active in comparison to 8a and 8b which were derived from 7-hydroxycoumarin. It is worthwhile to remind that introduction of bromine in the structure of the synthesized amides showed remarkable effect and in brief, according to Table 1, the best result is related

to 8c which involves 4-bromophenyl in the structure and this is in good agreement with the results from DPPH radical scavenging activity test.

Experimental

General

Melting points were taken on a *Kofler* hot stage apparatus and are uncorrected. ¹H and ¹³C NMR spectra were recorded on *Bruker FT-500*, using TMS as an internal standard. The IR spectra were obtained on a *Nicolet Magna FTIR 550* spectrophotometer (in KBr).

Chemicals

All chemicals, reagents and solvents were obtained from Sigma-Aldrich and Merck chemical companies and were used without any purification. Some coumarins were prepared according to references 35 and 36. TLC analyses were performed on a 3-10 cm aluminum sheet pre-coated with silica gel 60-254 (Merck). Visualization of spots on TLC plate was accomplished with UV light.

General procedure for synthesis of sulfonamides 5 and amides 8

Synthesis of 2-chloroacetamide derivatives 3 and 7

Chloroacetylchloride 2 (40 mmol) was added to a solution of sulfanilamide 5 or aniline 6 derivatives (20 mmol) in DMF (20 mL) at 0 °C, and the reaction was continued at room temperature for 5 h. After completion of reaction, the mixture was poured in cold water and the precipitate was filtered off and the pure products were obtained by recrystallization from H₂O or H₂O/EtOH.

Synthesis of sulfonamide and amide derivatives 5 and 8

To a solution of 2-chloroacetamide derivatives 3 or 7 (2 mmol) in DMF (20 mL), coumarin derivatives 4 (2 mmol), potassium carbonate K₂CO₃ (2 mmol), and potassium iodide KI (2 mmol) were added and the reaction mixture was heated at 70 °C for 8-12 h. After completion of the reaction, the mixture was poured in cold water and the precipitate was filtered off and the

pure products were obtained by recrystallization from EtOH.

2-((2-Oxo-2H-chromen-7-yl)oxy)-N-(4-sulfamoylphenyl)acetamide (5a)

Yield: 75 %; mp 266-268 °C; IR ν_{\max} (KBr): 3390, 3341, 3250, 3094, 2980, 1723, 1622, 1530, 1404, 1329, 1155; ^1H NMR (500 MHz, DMSO- d_6): 10.50 (s, NH), 8.00 (d, $J = 8.5$ Hz, 1H, H_4), 7.80-7.78 (m, 4H, ArH), 7.58 (d, $J = 8.0$ Hz, 1H, H_5), 7.28 (s, 2H, NH_2), 7.07-7.04 (m, 2H, H_6, H_8), 6.32 (d, $J = 8.5$ Hz, 1H, H_3), 4.90 (s, 2H, OCH_2); ^{13}C NMR (125 MHz, DMSO- d_6): 166.5, 160.9, 160.2, 155.2, 144.2, 141.2, 138.9, 129.6, 127.7, 126.7, 119.2, 112.9, 112.7, 101.7, 67.2. Anal. Calcd for $\text{C}_{17}\text{H}_{14}\text{N}_2\text{O}_6\text{S}$: C, 54.54; H, 3.77; N, 7.48. Found: C, 54.27; H, 3.50; N, 7.75.

2-((4-Methyl-2-oxo-2H-chromen-7-yl)oxy)-N-(4-sulfamoylphenyl)acetamide (5b)

Yield: 70 %; mp 295-298 °C; IR ν_{\max} (KBr): 3308, 3262, 3190, 3121, 1698, 1612, 1540, 1392, 1332, 1159; ^1H NMR (500 MHz, DMSO- d_6): 10.51 (s, 1H, NH), 7.82-7.79 (m, 4H, ArH), 7.72 (d, $J = 8.6$ Hz, 1H, H_5), 7.30 (s, 2H, NH_2), 7.07-7.04 (m, 2H, H_6, H_8), 6.23 (s, 1H, H_3), 4.91 (s, 2H, OCH_2), 2.40 (s, 3H, CH_3); ^{13}C NMR (125 MHz, DMSO- d_6): 166.5, 160.8, 160.1, 154.5, 153.4, 141.2, 138.9, 126.7, 126.6, 119.2, 113.7, 112.4, 111.5, 101.7, 67.2, 18.1. Anal. Calcd for $\text{C}_{18}\text{H}_{16}\text{N}_2\text{O}_6\text{S}$: C, 55.66; H, 4.15; N, 7.21. Found: C, 55.38; H, 4.32; N, 7.50.

2-((2-Oxo-2H-chromen-4-yl)oxy)-N-(4-sulfamoylphenyl)acetamide (5c)

Yield: 77 %; mp 164-165 °C; IR ν_{\max} (KBr): 3470, 3316, 3120, 3094, 1692, 1618, 1541, 1496, 1335, 1161; ^1H NMR (500 MHz, DMSO- d_6): 10.61 (s, 1H, NH), 7.95 (d, $J = 8.5$ Hz, 1H, H_5), 7.80-7.78 (m, 4H, ArH), 7.70 (t, $J = 8.0$ Hz, 1H, H_7), 7.44-7.42 (m, 2H, H_6, H_8), 7.30 (s, 1H, NH_2), 5.93 (s, 1H, H_3), 5.08 (s, 2H, OCH_2); ^{13}C NMR (125 MHz, DMSO- d_6): 165.1, 164.5, 161.4, 152.7, 141.1, 138.9, 132.9, 126.7, 124.3, 123.2, 119.2, 116.4, 115.0, 91.3, 67.5. Anal. Calcd for $\text{C}_{17}\text{H}_{14}\text{N}_2\text{O}_6\text{S}$: C, 54.54; H, 3.77; N, 7.48. Found: C, 54.80; H, 3.90; N, 7.15.

N-(4-(N-(5-Methylisoxazol-3-yl)sulfamoyl)phenyl)-2-((2-oxo-2H-chromen-7-yl)oxy)-N-

acetamide (5d)

Yield: 65 %; mp 274-277 °C; IR ν_{\max} (KBr): 3457, 3348, 3088, 2922, 1732, 1626, 1531, 1464, 1301, 1134; ^1H NMR (500 MHz, DMSO- d_6): 10.30 (s, 1H, NH), 8.00 (d, $J = 8.8$ Hz, 1H, H_4), 7.66 (d, $J = 8.0$ Hz, 1H, H_5), 7.61-7.59 (m, 4H, ArH), 7.05-7.03 (m, 2H, H_6, H_8), 6.31 (d, $J = 8.8$ Hz, 1H, H_3), 5.74 (s, 1H, methoxazole), 4.85 (s, 2H, OCH_2), 2.09 (s, 3H, CH_3); ^{13}C NMR (125 MHz, DMSO- d_6): 166.1, 165.9, 165.5, 160.9, 160.2, 155.1, 144.2, 142.8, 138.9, 129.5, 126.7, 124.9, 118.7, 112.9, 112.6, 101.7, 97.0, 67.2, 12.2. Anal. Calcd for $\text{C}_{21}\text{H}_{17}\text{N}_3\text{O}_7\text{S}$: C, 55.38; H, 3.76; N, 9.23. Found: C, 55.19; H, 3.92; N, 9.44.

Ethyl 2-oxo-7-(2-oxo-2-((4-sulfamoylphenyl)amino)ethoxy)-2H-chromen-3-carboxylate (5e)

Yield: 60 %; mp 295-298 °C; IR ν_{\max} (KBr): 3420, 3335, 3251, 3088, 2983, 1755, 1709, 1611, 1528, 1434, 1377, 1157; ^1H NMR (500 MHz, DMSO- d_6): 10.53 (s, 1H, NH), 8.74 (s, 1H, H_4), 7.89 (d, $J = 8.5$ Hz, H_5), 7.79-7.74 (m, 4H, ArH), 7.28 (s, 2H, NH_2), 7.11-7.08 (m, 2H, H_6, H_8), 4.96 (s, 2H, OCH_2), 4.30 (q, $J = 7.0$ Hz, 2H, OCH_2CH_3), 1.31 (t, $J = 7.0$ Hz, 3H, OCH_2CH_3); ^{13}C NMR (125 MHz, DMSO- d_6): 166.6, 163.6, 157.1, 156.6, 149.5, 141.6, 139.3, 132.1, 127.2, 119.6, 118.7, 114.2, 114.0, 113.0, 112.4, 67.7, 61.4, 14.5. Anal. Calcd for $\text{C}_{20}\text{H}_{18}\text{N}_2\text{O}_8\text{S}$: C, 53.81; H, 4.06; N, 6.27. Found: C, 53.98; H, 3.90; N, 6.41.

Ethyl 2-oxo-6-(2-oxo-2-((4-sulfamoylphenyl)amino)ethoxy)-2H-chromen-3-carboxylate (5f)

Yield: 65 %; mp 266-268 °C; IR ν_{\max} (KBr): 3354, 3251, 1750, 1692, 1572, 1535, 1491, 1320, 1170; ^1H NMR (500 MHz, DMSO- d_6): 10.48 (s, 1H, NH), 8.72 (s, 1H, H_4), 7.85-7.83 (m, 4H, ArH), 7.54 (d, $J = 8.0$ Hz, 1H, H_5), 7.56-7.52 (m, 2H, H_6, H_8), 7.28 (s, 2H, NH_2), 4.83 (s, 2H, OCH_2), 4.30 (q, $J = 6.9$ Hz, 2H, OCH_2CH_3), 1.31 (t, $J = 6.9$ Hz, 3H, OCH_2CH_3); ^{13}C NMR (125 MHz, DMSO- d_6): 166.7, 162.6, 156.1, 154.2, 149.3, 148.3, 141.2, 138.8, 126.7, 122.7, 119.2, 118.1, 118.0, 117.3, 113.1, 67.4, 61.2, 14.0. Anal. Calcd for $\text{C}_{20}\text{H}_{18}\text{N}_2\text{O}_8\text{S}$: C, 53.81; H, 4.06; N, 6.27. Found: C, 53.52; H, 4.22; N, 6.01.

2-[(4-Methyl-2-oxo-8-[(4-sulfamoylphenyl)carbamoyl]methoxy]-2H-chromen-7-yl)oxy]-N-

(4-sulfamoylphenyl)acetamide (5g)

Yield: 80 %; mp > 300 °C; IR ν_{\max} (KBr): 3345, 3255, 3120, 2920, 1695, 1615, 1530, 1405, 1350, 1148; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.50 (s, 1H, NH), 11.10 (s, 1H, NH), 7.90-7.69 (m, 8H, ArH), 7.53 (d, $J = 9.0$ Hz, 1H, H_5), 7.29 (s, 4H, 2NH₂), 7.17 (d, $J = 9.0$ Hz, 1H, H_6), 6.30 (s, 1H, H_3), 5.10 (s, 2H, OCH₂), 4.88 (s, 2H, OCH₂), 2.41 (s, 3H, CH₃); $^{13}\text{C NMR}$ (125 MHz, DMSO- d_6): 167.9, 166.0, 159.9, 154.1, 153.3, 147.1, 141.5, 139.1, 133.8, 132.6, 129.9, 127.4, 127.0, 121.2, 119.7, 119.4, 115.1, 112.2, 110.28, 72.2, 68.0, 18.7; Anal. Calcd for C₂₆H₂₄N₄O₁₀S₂: C, 50.64; H, 3.92; N, 9.09. Found: C, 50.83; H, 3.74; N, 9.32.

2-[(2-Oxo-4-[(4-sulfamoylphenyl)carbamoyl]methoxy]-2H-chromen-7-yl)oxy]-N-(4-sulfamoylphenyl)acetamide (5h)

Yield: 80 %; mp 205-207 °C; IR ν_{\max} (KBr): 3278, 3127, 1691, 1616, 1535, 1405, 1326, 1155; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.61 (s, 1H, NH), 10.53 (s, 1H, NH), 7.89 (d, $J = 8.6$ Hz, 1H, H_5), 7.83-7.76 (m, 8H, ArH), 7.30 (s, 4H, 2NH₂), 7.12-7.08 (m, 2H, H_6 , H_8), 5.79 (s, 1H, H_3), 5.06 (s, 2H, OCH₂), 4.93 (s, 2H, OCH₂); $^{13}\text{C NMR}$ (125 MHz, DMSO- d_6): 166.4, 165.2, 164.9, 161.8, 161.5, 154.4, 141.2, 141.1, 139.0, 138.9, 127.8, 126.8, 126.7, 124.5, 119.3, 112.6, 108.7, 101.5, 89.0, 67.5, 67.2. Anal. Calcd for C₂₅H₂₂N₄O₁₀S₂: C, 49.83; H, 3.68; N, 9.30. Found: C, 49.59; H, 3.49; N, 9.51.

2-(2-Oxo-2H-chromen-7-yloxy)-N-phenylacetamide (8a)

Yield: 75 %; mp 195-196 °C; IR ν_{\max} (KBr): 3375, 3057, 1725, 1675, 1594, 1530; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.17 (s, 1H, NH), 7.99 (d, $J = 9.5$ Hz, 1H, H_4), 7.66 (d, $J = 8.4$ Hz, 1H, H_5), 7.63 (d, $J = 7.7$ Hz, 2H, H_2 , H_6), 7.33 (t, $J = 7.7$ Hz, 2H, H_3 , H_5), 7.10 (t, $J = 7.7$ Hz, 1H, H_4), 7.07-7.03 (m, 2H, H_6 , H_8), 6.30 (d, $J = 9.5$ Hz, 1H, H_3), 4.85 (s, 2H, OCH₂); $^{13}\text{C NMR}$ (125 MHz, DMSO- d_6): 165.9, 161.0, 160.3, 155.2, 144.3, 138.3, 129.6, 128.8, 123.9, 119.7, 113.0, 112.9, 112.8, 101.7, 67.3. Anal. Calcd for C₁₇H₁₃NO₄: C, 69.15; H, 4.44; N, 4.74. Found: C, 68.88; H, 4.28; N, 4.96.

2-(2-Oxo-2H-chromen-7-yloxy)-N-(4-chlorophenyl)acetamide (8b)

Yield: 70 %; mp 226-228 °C; IR ν_{\max} (KBr): 3357, 1701, 1624, 1544, 1493; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.31 (s, 1H, NH), 7.99 (d, $J = 9.5$ Hz, 1H, H_4), 7.67-7.65 (m, 3H, H_5 , H_2 , H_6), 7.38 (t, $J = 8.8$ Hz, 2H, H_3 , H_5), 7.04-7.02 (m, 2H, H_6 , H_8), 6.30 (d, $J = 9.5$ Hz, 1H, H_3), 4.84 (s, 2H, OCH₂); $^{13}\text{C NMR}$ (125 MHz, DMSO- d_6): 166.1, 160.9, 160.3, 155.2, 144.3, 137.3, 129.6, 128.8, 127.4, 121.3, 112.9, 112.8, 101.7, 67.3. Anal. Calcd for C₁₇H₁₂ClNO₄: C, 61.92; H, 3.67; N, 4.25. Found: C, 62.15; H, 3.83; N, 4.02.

2-(2-Oxo-2H-chromen-7-yloxy)-N-(4-bromophenyl)acetamide (8c)

Yield: 70 %; mp 205-208 °C; IR ν_{\max} (KBr): 3356, 3093, 1732, 1683, 1627, 1590, 1540, 1492; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.31 (s, 1H, NH), 8.00 (d, $J = 9.4$ Hz, 1H, H_4), 7.67 (d, $J = 9.1$ Hz, 1H, H_5), 7.61 (d, $J = 8.8$ Hz, 2H, H_2 , H_6), 7.52 (t, $J = 8.8$ Hz, 2H, H_3 , H_5), 7.05-7.03 (m, 2H, H_6 , H_8), 6.31 (d, $J = 9.4$ Hz, 1H, H_3), 4.85 (s, 2H, OCH₂); $^{13}\text{C NMR}$ (125 MHz, DMSO- d_6): 166.1, 160.9, 160.2, 155.2, 144.3, 137.7, 131.6, 129.6, 121.7, 115.5, 112.9, 112.8, 101.7, 67.3. Anal. Calcd for C₁₇H₁₂BrNO₄: C, 54.57; H, 3.23; N, 3.74. Found: C, 54.23; H, 2.98; N, 3.28.

2-(2-Oxo-2H-chromen-4-yloxy)-N-phenylacetamide (8d)

Yield: 70 %; mp 251-253 °C; IR ν_{\max} (KBr): 3360, 3283, 3139, 1700, 1680, 1618, 1567, 1495; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.29 (s, 1H, NH), 7.95 (d, $J = 7.8$ Hz, 1H, H_5), 7.70 (t, $J = 7.8$ Hz, H_7), 7.62 (d, $J = 7.5$ Hz, H_2 , H_6), 7.45-7.40 (m, 2H, H_6 , H_8), 7.35 (t, $J = 7.5$ Hz, H_3 , H_5), 7.11 (t, $J = 7.5$ Hz, H_4), 5.90 (s, 1H, H_3), 5.04 (s, 2H, OCH₂); $^{13}\text{C NMR}$ (125 MHz, DMSO- d_6): 164.6, 164.5, 161.5, 152.8, 138.2, 133.0, 128.9, 124.3, 123.9, 123.3, 119.6, 116.5, 115.0, 91.2, 67.7; Anal. Calcd for C₁₇H₁₃NO₄: C, 69.15; H, 4.44; N, 4.74. Found: C, 69.30; H, 4.73; N, 4.90.

2-(2-Oxo-2H-chromen-4-yloxy)-N-(4-chlorophenyl)acetamide (8e)

Yield: 70 %; mp 260-262 °C; IR ν_{\max} (KBr): 3421, 3288, 3139, 1700, 1685, 1618, 1490; $^1\text{H NMR}$ (500 MHz, DMSO- d_6): 10.41 (s, 1H, NH), 7.95 (d, $J = 8.9$ Hz, 1H, H_5), 7.45-7.40 (m, 4H, H_2 , H_3 , H_5 , H_6), 7.70 (m, 3H, H_6 , H_7 , H_8),

5.91 (s, 1H, H₃), 5.03 (s, 2H, OCH₂); ¹³C NMR (125 MHz, DMSO-*d*₆): 165.1, 165.0, 161.9, 153.2, 137.6, 133.4, 129.2, 127.9, 124.7, 123.7, 121.7, 116.9, 115.4, 91.7, 68.0; Anal. Calcd for C₁₇H₁₂ClNO₄: C, 61.92; H, 3.67; N, 4.25. Found: C, 62.09; H, 3.91; N, 4.05.

2-(4-Methyl-2-oxo-2H-chromen-7-yloxy)-N-phenylacetamide (8f)

Yield: 72 %; mp 232-234 °C; IR ν_{max} (KBr): 3365, 1700, 1680, 1629, 1541, 1501, 1393; ¹H NMR (500 MHz, DMSO-*d*₆): 10.18 (s, 1H, NH), 7.72 (d, *J* = 8.7 Hz, 1H, H₅), 7.64 (d, *J* = 7.8 Hz, 2H, H₂, H₆), 7.33 (t, *J* = 7.8 Hz, 1H, H₃, H₅), 7.11-7.03 (m, 3H, H₆, H₈, H₄), 6.21 (s, 1H, H₃), 4.85 (s, 2H, OCH₂), 2.40 (s, 3H, CH₃); ¹³C NMR (125 MHz, DMSO-*d*₆): 165.8, 160.8, 160.1, 154.5, 153.4, 138.3, 128.8, 126.6, 123.8, 119.7, 113.6, 112.4, 111.4, 101.7, 67.3, 18.1; Anal. Calcd for C₁₈H₁₅NO₄: C, 69.89; H, 4.89; N, 4.53. Found: C, 70.05; H, 4.56; N, 4.38.

2-(4-Methyl-2-oxo-2H-chromen-7-yloxy)-N-(4-chlorophenyl)acetamide (8g)

Yield: 75 %; mp 238-240 °C; IR ν_{max} (KBr): 3359, 2914, 1705, 1680, 1593, 1536, 1490; ¹H NMR (500 MHz, DMSO-*d*₆): 10.31 (s, 1H, NH), 7.73 (d, *J* = 8.7 Hz, 1H, H₅), 7.67 (d, *J* = 8.7 Hz, 1H, H₂, H₆), 7.38 (t, *J* = 8.7 Hz, 1H, H₃, H₅), 7.07-7.03 (m, 2H, H₆, H₈), 6.24 (s, 1H, H₃), 4.86 (s, 2H, OCH₂), 2.41 (s, 3H, CH₃); ¹³C NMR (125 MHz, DMSO-*d*₆): 166.5, 161.2, 160.5, 154.9, 153.8, 137.7, 129.1, 127.8, 127.0, 121.6, 114.1, 112.8, 111.9, 102.1, 67.6, 18.6; Anal. Calcd for C₁₈H₁₄ClNO₄: C, 62.89; H, 4.10; N, 4.07. Found: C, 62.63; H, 3.88; N, 4.25.

2-(4-Methyl-2-oxo-2H-chromen-7-yloxy)-N-(4-bromophenyl)acetamide (8h)

Yield: 68 %; mp 259-262 °C; IR ν_{max} (KBr): 3360, 2913, 1705, 1680, 1623, 1532, 1490; ¹H NMR (500 MHz, DMSO-*d*₆): 10.29 (s, 1H, NH), 7.72 (d, *J* = 8.6 Hz, 1H, H₅), 7.62 (d, *J* = 8.3 Hz, 2H, H₂, H₆), 7.52 (t, *J* = 8.3 Hz, 2H, H₃, H₅), 7.06-7.03 (m, 2H, H₆, H₈), 6.23 (s, 1H, H₃), 4.86 (s, 2H, OCH₂), 2.40 (s, 3H, CH₃); ¹³C NMR (125 MHz, DMSO-*d*₆): 166.0, 160.7, 160.0, 154.5, 153.3, 137.6, 131.6, 126.5, 121.6, 115.4, 113.6, 112.3, 111.4, 101.7, 67.2, 18.1; Anal. Calcd for C₁₈H₁₄BrNO₄: C, 55.69; H, 3.63; N, 3.61. Found:

C, 55.92; H, 3.45; N, 3.85.

2-(2-Oxo-4-phenyl-2H-chromen-7-yloxy)-N-phenylacetamide (8i)

Yield: 73 %; mp 230-232 °C; IR ν_{max} (KBr): 3351, 3133, 1716, 1680, 1621, 1540, 1401; ¹H NMR (500 MHz, DMSO-*d*₆): 10.19 (s, 1H, NH), 7.64 (d, *J* = 7.8 Hz, 2H, H₂, H₆), 7.56-7.53 (m, 5H, Ph), 7.39 (d, *J* = 8.9 Hz, 1H, H₅), 7.33 (t, *J* = 7.8 Hz, 2H, H₃, H₅), 7.14 (d, *J* = 2.0 Hz, 1H, H₈), 7.09 (t, *J* = 7.8 Hz, 1H, H₄), 7.03 (dd, *J* = 8.9, 2.0 Hz, 1H, H₆), 6.26 (s, 1H, H₃), 4.87 (s, 2H, OCH₂); ¹³C NMR (125 MHz, DMSO-*d*₆): 166.2, 161.4, 160.3, 155.7, 155.5, 1387, 135.4, 130.1, 129.3, 129.2, 128.8, 128.3, 124.2, 120.1, 113.2, 112.7, 112.1, 102.6, 67.7. Anal. Calcd for C₂₃H₁₇NO₄: C, 74.38; H, 4.61; N, 3.77. Found: C, 74.08; H, 4.38; N, 3.92.

2-(2-Oxo-4-phenyl-2H-chromen-7-yloxy)-N-(4-chlorophenyl)acetamide (8j)

Yield: 75 %; mp 266-268 °C; IR ν_{max} (KBr): 3416, 1716, 1690, 1613, 1531, 1430; ¹H NMR (500 MHz, DMSO-*d*₆): 10.33 (s, 1H, NH), 7.67 (d, *J* = 8.8 Hz, 1H, H₅), 7.57-7.53 (m, 5H, Ph), 7.40-7.38 (m, 4H, H₂, H₃, H₅, H₆), 7.14 (d, *J* = 2.4 Hz, 1H, H₈), 7.03 (dd, *J* = 8.8, 2.4 Hz, H₆), 6.27 (s, 1H, H₃), 4.88 (s, 2H, OCH₂); ¹³C NMR (125 MHz, DMSO-*d*₆): 166.4, 161.4, 160.3, 160.3, 155.7, 155.5, 137.7, 135.4, 130.1, 129.3, 129.1, 128.9, 127.8, 121.6, 113.2, 112.8, 112.1, 102.6, 67.7; Anal. Calcd for C₂₃H₁₆ClNO₄: C, 68.07; H, 3.97; N, 3.45. Found: C, 68.29; H, 4.16; N, 3.33.

Antimicrobial activity

Compounds 5a-h and 8a-j were evaluated for their antibacterial activity against Gram-positive (*Staphylococcus aureus* ATCC 6538p, *Staphylococcus epidermidis* ATCC 12228) and Gram-negative (*Escherichia coli* ATCC 8739, *Klebsiella pneumonia* ATCC 10031, and *Pseudomonas aeruginosa* ATCC 9027) bacteria using conventional agar-dilution method (32). The MIC (minimum inhibitory concentration) values were determined by comparison to ciprofloxacin as a reference drug. Two-fold dilution of the test compounds and the standard antibacterial agents were prepared in dimethyl sulfoxide (DMSO; 1 mL). Each dilute was added

to molten Mueller-Hinton (MH) agar (19 mL) at 50 °C to give a final concentration of 100, 50, 25, 12.5, 6.25, 3.125, 1.56, 0.78, 0.39, 0.195, 0.098, 0.049, 0.025, 0.012, 0.006 and 0.003 µg mL⁻¹. The bacterial inocula were prepared by suspending overnight colonies from MH agar media in 0.85% saline. The inocula were adjusted photometrically at 600 nm to a cell density equivalent to approximately 0.5 McFarland standards (1.5 × 10⁸ CFU/mL). The suspensions were then diluted in 0.85% saline to make 10⁷ CFU/mL. The plates were spot-inoculated with 1 µL of bacterial suspensions (10⁴ CFU/spot); including a control plate containing 1 mL DMSO without any antibacterial agent. The plates were incubated at 35-37 °C and examined after 18 h. The MIC was determined as the lowest concentration of the agent that completely inhibits visible growth of the microorganisms.

Antioxidant activity

DPPH radical scavenging assay

Several concentrations of test compounds 5a-h and 8a-j in DMSO were prepared. The compound solution (1.0 mL) was added to the methanolic DPPH solution (2.0 mL, 0.1 mM), and the mixture was kept in the dark for 15 min. The absorbance at 517 nm was then measured by an UV / visible spectrophotometer. The percent scavenging activity was calculated using the following formula: inhibition (%) = 100. (Abs_{control} - Abs_{compound}) / Abs_{control}. The DPPH radical scavenging activity of compounds was expressed in terms of IC₅₀ (mM), which is obtained from linear regression plot between concentrations of test compound and percent inhibitions (33).

Ferric reducing antioxidant power (FRAP) assay

The FRAP assay reagent was prepared by adding 10 vol of 300 mM acetate buffer, pH 3.6 (3.1 g sodium acetate and 16 mL glacial acetic acid), 1 vol of 10 mM 2,4,6-tripyridyl-s-triazine prepared in 40 mM HCl, and 1 vol of 20 mM FeCl₃. The mixture was diluted to 1/3 with methanol and prewarmed at 37 °C. This reagent (3 mL) was mixed with 0.1 mL diluted test compounds 5a-h and 8a-j. The mixture was shaken and incubated at 37 °C for 8 min, and the absorbance was read at 593 nm. A blank with

only 0.1 mL methanol was used for calibration. The difference in absorbance between the tested sample and the blank reading was calculated, and the data were expressed as mM of ferric reduced to ferrous form (34).

Conclusion

In summary, a series of new sulfonamide and amide derivatives containing coumarin derivatives were synthesized with a simple procedure in two steps starting from sulfanilamide and aniline derivatives using chloroacetyl chloride and various coumarin derivatives. The biological activity of target compounds was screened. Although they did not exhibit good antimicrobial activities but they showed moderate to noticeable antioxidant activities.

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References

- (1) Genç Y, Özkanca R and Bekdemir Y. Antimicrobial activity of some sulfonamide derivatives on clinical isolates of *Staphylococcus aureus*. *Ann. Clin. Microbiol. Antimicrob* (2008) 7: 17-22.
- (2) Özbek N, Katircioğlu H, Karacan N and Baykal T. Synthesis, characterization and antimicrobial activity of new aliphatic sulfonamide. *Bioorg. Med. Chem.* (2007) 15: 5105-5109.
- (3) Mun J, Jabbar AA, Devi NS, Yin S, Wang Y, Tan C, Culver D, Snyder JP, Van Meir EG and Goodman MM. Design and *in-vitro* activities of N-alkyl-N-[(8-R-2,2-dimethyl-2H-chromen-6-yl)methyl] heteroarylsulfonamides, novel, small-molecule hypoxia inducible factor-1 pathway inhibitors and anticancer agents. *J. Med. Chem.* (2012) 55: 6738-6750.
- (4) El-Sayed NS, El-Bendary ER, El-Ashry SM and El-Kerdawy MM. Synthesis and antitumor activity of new sulfonamide derivatives of thiadiazolo[3,2-a]pyrimidines. *Eur. J. Med. Chem.* (2011) 46: 3714-3720.
- (5) Borne RF, Peden RL, Waters IW, Weiner M, Jordan R and Coats EA. Anti inflammatory activity of para substituted N-benzenesulfonyl derivatives of anthranilic acid. *J. Pharm. Sci.* (1974) 63: 615-617.
- (6) De Clercq E. New developments in anti-HIV chemotherapy. *Curr. Med. Chem.* (2001) 8: 1543-1572.

- (7) Mengelers MJB, Hougee PE, Janssen LHM and Van Miert ASJPAM. Structure-activity relationships between antibacterial activities and physicochemical properties of sulfonamides. *J. Vet. Pharmacol. Ther.* (1997) 20: 276-283.
- (8) Mirian M, Zarghi A, Sadeghi S, Tabaraki P, Tavallae M, Dadrass O and Sadeghi-Aliabadi H. Synthesis and cytotoxic evaluation of some novel sulfonamide derivatives against a few human cancer cells. *Iran. J. Pharm. Res.* (2011) 10: 741-748.
- (9) Bhusari KP, Khedekar PB, Umathe SN, Bahekar RH and Rao ARR. Synthesis of 8-bromo-9-substituted-1,3-benzothiazolo-[5,1-b]-1,3,4-triazoles and their anthelmintic activity. *Indian J. Heterocyclic Chem.* (2000) 9: 275-278.
- (10) Bhusare SR, Pawar RP and Vibhute YB. Synthesis and antibacterial activity of some new 2-(substituted phenyl sulfonamido)-6-substituted benzothiazoles. *Indian J. Heterocyclic Chem.* (2001) 11: 79-80.
- (11) Ahmed B, Khan SA and Alam T. Synthesis and antihepatotoxic activity of some heterocyclic compounds containing the 1,4-dioxane ring system. *Pharmazie* (2003) 58: 173-176.
- (12) Shekar BC, Roy K and De AU. Synthesis of some new p-toluene sulfonamido glutaramides. *Indian J. Heterocyclic Chem.* (2001) 10: 237-238.
- (13) Doungsoongnuen S, Worachartcheewan A, Pingaew R, Suksrichavalit T, Prachayasittikul S, Ruchirawat S and Prachayasittikul V. Investigation on biological activities of anthranilic acid sulfonamide analogs. *EXCLI J.* (2011) 10: 155-161.
- (14) Humphrey JM and Chamberlin AR. Chemical synthesis of natural product peptides: coupling methods for the incorporation of noncoded amino acids into peptides. *Chem. Rev.* (1997) 97: 2243-2266.
- (15) Koz'minykh VO. Synthesis and biological activity of substituted amides and hydrazides of 1,4-dicarboxylic acids. *Pharm. Chem. J.* (2006) 40: 8-17.
- (16) Tehrani M, Zarghi A and Jabarian L. Design and synthesis of new imidazole derivatives of captopril. *Iran. J. Pharm. Res.* (2005) 1: 37-41.
- (17) Huczyński A, Janczak J, Stefańska J, Antoszczak M and Brzezinski B. Synthesis and antimicrobial activity of amide derivatives of polyether antibiotic-salinomycin. *Bioorg. Med. Chem. Lett.* (2012) 22: 4697-4702.
- (18) Narasimhan B, Belsare D, Pharande D, Mourya V and Dhake A. Esters, amides and substituted derivatives of cinnamic acid: Synthesis, antimicrobial activity and QSAR investigations. *Eur. J. Med. Chem.* (2004) 39: 827-834.
- (19) Kono M, Matsumoto T, Kawamura T, Nishimura A, Kiyota Y, Oki H, Miyazaki J, Igaki S, Behnke CA, Shimojo M and Kori M. Synthesis, SAR study, and biological evaluation of a series of piperazine ureas as fatty acid amide hydrolase (FAAH) inhibitors. *Bioorg. Med. Chem.* (2013) 21: 28-41.
- (20) Peña S, Scarone L, Manta E, Stewart L, Yardley V, Croft S and Serra G. Synthesis of a microcystis aeruginosa predicted metabolite with antimalarial activity. *Bioorg. Med. Chem. Lett.* (2012) 22: 4994-4997.
- (21) Hranjec M, Sović I, Ratkaj I, Pavlović G, Ilić N, Valjalo L, Pavelić K, Pavelić SK and Karminski-Zamola, G. Antiproliferative potency of novel benzofuran-2-carboxamides on tumour cell lines: cell death mechanisms and determination of crystal structure. *Eur. J. Med. Chem.* (2013) 59: 111-119.
- (22) Boonya-Udtayan S, Eno M, Ruchirawat S, Mahidol C and Thasana N. Palladium-catalyzed intramolecular C-H amidation: synthesis and biological activities of indolobenzazocin-8-ones. *Tetrahedron* (2012) 68: 10293-10301.
- (23) Salimi M, Ghahremani MH, Naderi N, Amini M, Salimi E, Amanlou M, Abdi K, Salehi R and Shafiee A. Design, synthesis and pharmacological evaluation of 4-[2-alkylthio-5(4)-(4-substitutedphenyl)imidazole-4(5)yl]benzenesulfonamides as selective COX-2 inhibitors. *Acta. Pharmacol. Sin.* (2007) 28: 1254-1260.
- (24) Amini M, Navidpour L and Shafiee A. Synthesis and antitubercular activity of new N,N-diaryl-4-(4,5-dichloroimidazole-2-yl)-1,4-dihydro-2,6-dimethyl-3,5-pyridinedicarboxamides. *Daru* (2008) 16: 9-12.
- (25) Aliabadi A, Shamsa F, Ostad SN, Emami S, Shafiee A, Davoodi J and Foroumadi A. Synthesis and biological evaluation of 2-phenylthiazole-4-carboxamide derivatives as anticancer agents. *Eur. J. Med. Chem.* (2010) 45: 5384-5389.
- (26) Khoobi M, Foroumadi A, Emami S, Safavi M, Dehghan G, Alizadeh BH, Ramazani A, Ardestani SK and Shafiee A. Coumarin-Based bioactive compounds: facile synthesis and biological evaluation of coumarin-fused 1,4-thiazepines. *Chem. Biol. Drug Des.* (2011) 78: 580-586.
- (27) Alipour M, Khoobi M, Foroumadi A, Nadri H, Moradi A, Sakhteman A, Ghandi M and Shafiee A. Novel coumarin derivatives bearing N-benzyl pyridinium moiety: potent and dual binding site acetylcholinesterase inhibitors. *Bioorg. Med. Chem.* (2012) 20: 7214-7222.
- (28) Khoobi M, Emami S, Dehghan G, Foroumadi A, Ramazani A and Shafiee A. Synthesis and free radical scavenging activity of coumarin derivatives containing a 2-methylbenzothiazoline motif. *Arch. Pharm. Chem. Life Sci.* (2011) 344: 588-594.
- (29) Liu J, Wu F, Chen L, Zhao L, Zhao Z, Wang M and Lei S. Biological evaluation of coumarin derivatives as mushroom tyrosinase inhibitors. *Food Chem.* (2012) 135: 2872-2878.
- (30) Catto M, Pisani L, Leonetti F, Nicolotti O, Pesce P, Stefanachi A, Cellamare S and Carotti A. Design, synthesis and biological evaluation of coumarin alkylamines as potent and selective dual binding site inhibitors of acetylcholinesterase. *Bioorg. Med. Chem.* (2013) 21: 146-152.
- (31) Li B, Pai R, Di M, Aiello D, Barnes MH, Butler MM, Tashjian TF, Peet NP, Bowlin TL and Moir DT. Coumarin-based inhibitors of bacillus anthracis and staphylococcus aureus replicative DNA helicase:

- chemical optimization, biological evaluation, and antibacterial activities. *J. Med. Chem.* (2012) 55: 10896-10908.
- (32) Baron EJ, Bailey WR and Finegold SM. *Bailey and Scott's Diagnostic Microbiology*. 8th ed. Mosby, St. Louis (1990).
- (33) Brand-Williams W, Cuvelier ME and Berset C. Use of a free radical method to evaluate antioxidant activity. *LWT-Food Sci. Technol.* (1995) 28: 25-30.
- (34) Benzie IFF and Strain JJ. Ferric reducing/antioxidant power assay: direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methods Enzymol.* (1999) 299: 15-27.
- (35) Shen Q, Shao J, Peng Q, Zhang W, Ma L, Chan ASC and Gu L. Hydroxycoumarin derivatives: novel and potent α -Glucosidase inhibitors. *J. Med. Chem.* (2010) 53: 8252-8259.
- (36) Xie L, Takeuchi Y, Cosentino LM and Lee KH. Anti-AIDS agents. 37. Synthesis and structure-activity relationships of (3R,4R)-(+)-cis-khellactone derivatives as novel potent anti-HIV agents. *J. Med. Chem.* (1999) 42: 2662-2672.
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