Comparison of radiosensitizing effect of Resveratrol on monolayer and spheroid culture of DU145 prostatic cell line

M.S. Nezamtaheri¹, S. Khoei², A.R. Nikoofar³, B. Goliaei¹*

¹Laboratory of Biophysics and Molecular Biology, Institute of Biochemistry and Biophysics, University of Tehran, Tehran, Iran
²Department of Medical Physics, School of medical basic science, Tehran University of Medical Science, Tehran, Iran
³Radiology Departments, Tehran University of Medical Sciences, Tehran, Iran

*Corresponding author: Dr. Bahram Goliaei, Laboratory of Biophysics and Molecular Biology, Institute of Biochemistry and Biophysics, University of Tehran, P.O. Box 13145-1384, Tehran, Iran. Fax: +98 21 66956985 E-mail: goliaei@ibb.ut.ac.ir

Background: Radiotherapy is an established therapeutic modality for prostate cancer. Resveratrol, a natural antioxidant, has been shown to inhibit carcinogenesis and to block the process of tumor initiation and progression. No data is available on the response of cellular spheroid to Resveratrol. In this study we have examined the effect of Resveratrol on the radiation response of human prostate cell line DU145 in monolayer and spheroid cultures.

Materials and Methods: Radiosensitivity was assessed using viability and colony formation assay. Apoptosis and necrosis were assessed using acridine orange/ethidium bromide double staining.

Results: The colony formation assay did not show any significant radio-sensitizing effect, but apoptosis assay showed significant radio-sensitizing effect of Resveratrol on DU145 cells grown as monolayer. In the spheroid cells the results of apoptosis test were not significant and corresponded closely to the result of survival curve.

Conclusion: While Resveratrol could sensitize DU145 cells in monolayer to ionizing radiation, it did not have any effect on sensitivity of cells cultured in spheroid cultures.

INTRODUCTION

Radiotherapy is broadly used for the therapy of cancer through induction of apoptosis in tumor cells (1). Biological radiosensitizers can enhance the sensitivity of tumors to ionizing radiation (2). Resveratrol, a potent anticancer and radiosensitizer compound, is a natural polyphenol derived from grapes, plums and peanuts (3). A large number of in vitro studies have dealt with the potent antiproliferative/pro-apoptotic effects of Resveratrol on different human prostatic cancer cell lines (4). Previous study by Scarlatti et al. has cleared the underling molecular mechanism of overcoming radioresistance in DU145 cells treated with Resveratrol. It has been shown that Resveratrol through increasing de novo production of ceramide in DU145 cell line results in apoptotic cell death (2). Furthermore, these cells can be self-assembled into morphologically large-size, spheroid-shaped and stable aggregates so-called multicellular tumor spheroids (MCTS) through intracellular communication networks. The microenvironment of MCTSs is closer to in vivo tumors than monolayer cultures (5). In 1972 Durand et al. demonstrated that exposing spheroids of Chinese Hamster V79-1716 cells to ionizing radiation made them more resistance than monolayer cultured cells (6). So far the effect of Resveratrol on cancer cells cultured in MCTS has not been evaluated. In this work we have compared the radiosensitizing effect of Resveratrol on MCTS and monolayer cultures of DU145 prostatic cell line.

MATERIALS AND METHODS

Reagent

Resveratrol from sigma-Aldrich was...
dissolved in ethanol (96% Merck) before use. Trypan blue, metal green, Penicillin and MTT (3-(4, 5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide, a yellow tetrazole), were from Sigma–Aldrich. Trypsin and ethidium bromide (EB) was from Merck. Cell culture media (RPMI-1640) and fetal bovine serum were from Gibco. Streptomycin was from Jaberebn-Hayan. Agar was from Difco, Detroit, MI, USA (Bacto agar). Acridine orange (AO) was from Hopkin & Williams Ltd.

**Irradiation procedure**

**Irradiation of monolayer cell culture**

Cells were cultured at 25×10^4 cells per flask in T-25 culture. After 4 days, the cells were exposed to 2, 4, 6, 8 and 10 Gy X-rays with dose rate of 200 cGy/min using linear accelerator (primus, Siemens). Control cells were not exposed to X-ray. For treatment of Resveratrol, after 24 h, cells were treated with 25 µM Resveratrol or 0.05% of ethanol (as vehicle) for 72 h. Then, cells were irradiated and cell viability and apoptotic and necrotic cells were evaluated.

**Irradiation of spheroid culture**

Cells were cultured at 5×10^5 cells per Petri dish in 100 mm dishes. After 11 days, the spheroids were divided into T25 flasks. The cells were then exposed to 2, 4, 6 and 8 Gy of X-rays with dose rate of 200 cGy/min. After 8 day, spheroids were treated with 50 µM Resveratrol or 0.05% of ethanol (as vehicle) for 72 h. Then, cells were irradiated and cell viability and apoptosis and necrosis were evaluated.

**MTT assay**

Exponentially growing cells plated into 96 well plates and incubated 24 h at 37 °C in 5% CO2. For spheroids, after 11 days, spheroids were dispersed, plated into 96 well plates, and incubated 24 h at 37 °C in CO2. Cells were subsequently exposed to incremental concentration of Resveratrol (10 -100µM for monolayer and 50-200 µM for spheroid) in 200 µL RPMI and incubated for 72 h. Then, 20 µL MTT (5 mg/ml) was add to each well protected from light and incubated at 37 °C for 4 h. Formosan crystals were dissolved by adding 100 µL of DMSO (99% HPLC- Merck ) for 30 min. An ELISA plate reader (Lab systems multiskan MS) was used to read the absorbance with a wavelength of 570nm.

**Cell survival clonogenic assay**

Briefly, 2×10^5 cells were incubated into T-25 Flask. After 24 h, the cells were incubated for 72 h at 37 °C with or without Resveratrol (25µM). For spheroids, the cells were treated with Resveratrol (50 µM) at 8 days after spheroid formation and were incubated with or without Resveratrol for 72h. Next, the cells were irradiated with X-rays (2Gy). Cells were seeded in 60-mm dishes at various cell densities. After 9 days, the resulting colonies were stained with crystal violet dissolved in PBS. Colonies containing more than 50 cells were scored as survivors. Cell survival curve was analyzed with multi-target single-hit model (MTSH).

**Assay for apoptosis and necrosis**

Briefly, 2×10^5 cells were incubated into T-25 flasks. After 24 h, the cells were incubated for 72h at 37 °C with or without Resveratrol (for monolayer 25 µM and for spheroid 50 µM). Next, the cells were irradiated with X-rays (2 Gy). Then, the cells were stained with AO (100 mg/ml) and EB (100 mg/ml). Viable, apoptotic and necrotic cells were counted under a fluorescence microscope.

**Statistical analysis**

Differences between groups were analyzed by using repeated measures analysis of variance (ANOVA) and P-value < 0.05 was considered to be significant.

**RESULTS**

**MTT Assay**

The effect of Resveratrol on the viability of DU145 cells determined by MTT assay is
shown in figure 1a. The half maximal inhibitory concentration (IC50) for Resveratrol was calculated as 43 μM. Cytotoxicity of Resveratrol on spheroids was measured in the same way as monolayer cultures and IC50 was calculated 163 μM (figure 1b). There was significant difference in IC50 for monolayer and spheroid cells.

significantly, increased the radiosensitivity of Du145 cells cultured in monolayer condition.

There was significant difference in IC50 for monolayer and spheroid cells.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Dq</th>
<th>D0</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>X-ray and Resveratrol</td>
<td>1.16±0.087</td>
<td>0.75±0.004</td>
<td>4.63±0.49</td>
</tr>
<tr>
<td>X-ray</td>
<td>2.04±0.283</td>
<td>0.79±0.025</td>
<td>13.16±6.44</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Groups</th>
<th>Dq</th>
<th>D0</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>X-ray and Resveratrol</td>
<td>1.94±0.075</td>
<td>0.92±0.05</td>
<td>8.29±1.67</td>
</tr>
<tr>
<td>X-ray</td>
<td>2.11±0.755</td>
<td>1.16±0.26</td>
<td>6.18±9.35</td>
</tr>
</tbody>
</table>

**Effect of Resveratrol on radiosensitivity of monolayers**

Figure 2A shows the cell survival curves of actively growing DU145 cells in monolayer cultures exposed to X-rays with and without Resveratrol treatment. Fitting parameters for MTSH model is shown in table 1a. In survival curve of DU145 cells the value of n, decreased in the combined treatment of Resveratrol and radiation as compared to radiation alone. Based on Dn values, Resveratrol, slightly but not
Effect of Resveratrol on radiosensitivity of spheroids

The results of clonogenic survival assay are shown in figure 2B. Fitting parameters for MTSH model is shown in table 1b. Resveratrol, slightly but not significantly, increased the radiosensitivity of Du145 cells cultured in spheroid condition.

In the survival curve, Dq, which is a measure of the width of the shoulder of survival curve, showed a significant decrease in the combination of the two treatments (Resveratrol and radiation) in monolayer compared to spheroid in the same condition. D0 value represents the measure of sensitivity of the target. In the present study Resveratrol decreased D0 in both monolayer and spheroid cultures irradiated with 2 Gy of X-Ray.

Effect of Resveratrol on radiation-induced apoptosis and necrosis in DU145 cells

Figure 4 A shows the percentage of apoptotic and necrotic cells after X-irradiation with and without Resveratrol treatment. For cells irradiated without Resveratrol treatment, the percentages of apoptosis and necrosis were approximately 18.7 and 0.7 respectively. But in these cells, early apoptotic cells percentage was more than late apoptotic cells percentage. For cells irradiated with Resveratrol treatment, the percentage of apoptotic cells markedly increased, to approximately 59.5. These data were significantly more than the percentage of apoptotic cells induced by both Resveratrol and radiation treatment alone.

Effect of Resveratrol on radiation-induced apoptosis and necrosis on DU145 Cells grown as spheroid

Figure 3 shows images of apoptotic, necrotic, as well as control DU145 cells after AO/EB staining. Late apoptotic cells have an orange nucleus showing condensation of chromatin and necrotic cells displayed an orange nucleus with intact structure. Figure 4B shows the percentage of apoptotic and necrotic cells after X-irradiation with and without Resveratrol treatment in spheroid cells. For cells irradiated without Resveratrol treatment, the percentages of apoptosis and necrosis were approximately 24.8 and 6 respectively. For cells irradiated with Resveratrol treatment, the percentage of apoptotic cells increased but these data were not significant and corresponded closely to the result of survival curve.

DISCUSSION

Resveratrol is well-known as a potent radiosensitizer compound in a variety of cancer cell lines. Scarlatti et al. showed that resveratrol enhanced tumor cell killing and inhibited the clonogenic survival in resistant irradiated Du145 cells. In the survival curve (Resveratrol and X-ray) the shoulder region declined rapidly. Regarding the shoulder region of the curve as a measure of the repair capacity of the cell for
radiation damage after exposure to low doses, especially ~ 2 Gy \(^7\), it may be suggested that Resveratrol sensitizes DU145 in 2Gy dose of radiation through inhibition of repair enzymes. This effect was not clear in the survival curves of spheroids. The clonogenic radiosensetization was more pronounced for spheroid cultures compared to monolayers. According to results presented in the previous section the most pronounced effect of Resveratrol was on the induction of apoptosis. As expected, the apoptotic shock was more severe in monolayer cultures as compared to spheroids. Our results indicate that Resveratrol can sensitize DU145 prostatic cells in monolayer and spheroid cultures to radiation but, under similar conditions, Resveratrol and 2Gy of radiation are more effective on monolayers then on spheroid cultures. The low efficacy of Resveratrol on the cell death in spheroid cells is likely due to the fact that the drug cannot reach all the cells, especially those located in the core of the aggregate.

REFERENCES


http://www.SID.ir