

Chemical Composition and Antioxidant Activity of the Extract and Essential oil of *Rosa damascena* from Iran, Population of Guilan

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ABSTRACT

Background and the purpose of study: *Rosa damascena* Mill. (Rosaceae) has cooling, soothing, astringent, and anti-inflammatory effects, and has been used in the north of Iran as a cardiotoxic agent. The aim of this study was to identify components of *R. damascena* (cultivated in Guilan Province) extract and essential oil and to study their biological activities.

Methods: Essential oil of *R. damascena* was prepared by hydrodistillation and analyzed with GC/MS instrument. The antioxidant activity of hydro-alcoholic extract of petals and essential oil was measured using free radical scavenging activity with 2,2-diphenyl, 1-picrylhydrazyl (DPPH) and lipid peroxidation (ferric ammonium thiocyanate) methods.

Results: Hydro-alcoholic extract showed strong free radical scavenging capacity compared to lipid peroxidation inhibitory effects. IC₅₀ values of the extract were 2.24 µg/mL and 520 µg/mL in free radical scavenging and lipid peroxidation assays, respectively. The major components of essential oil were linalool (3.8%), nerol (3.05%), geraniol (15.05%), 1-nonadecene (18.56%), n-tricosane (16.68%), hexatriacontane (24.6%) and n-pentacosane (3.37%). The bioassay-guided fractionation of extract led to the isolation of three flavonol glycosides: quercetin-3-O-glucoside, kaempferol-3-O-rhamnoside and kaempferol-3-O-arabinoside. The IC₅₀ value of the radical scavenging activity of kaempferol-3-O-rhamnoside which was, 530 µg/mL was weaker than the extract.

Major conclusion: The petal of this cultivated rose has no bitter taste and because of its potential antioxidant activity and good taste, can be used as food flavor and a preventing agent for many diseases.

Keywords: *Rosa damascena*, Flavonoids, Antioxidant, Essential oil

INTRODUCTION

Rosa damascena Mill. (Rosaceae) is a small plant with aromatic, light pink flower which appears in spring (1). At present, *R. damascena* is the principle species cultivated for rose attar not only in Bulgaria but also in Iran and India (2). This plant is cultivated originally in Kashan Province (central part of Iran) for preparing rose water and attar. *R. damascena* and *R. gallica* var. *officinalis* Thory were widely grown in medieval times for medicinal purposes (3). *R. damascena* also has some other benefits such as cooling, soothing, astringent, and anti-inflammatory effects (4), especially it has been used in north of Iran as a cardiotoxic agent. Considerable interest has developed recently concerning the alternative utilization of aromatic plants rich in essential oils as antioxidant and antibacterial agents in the food industry (5). On the basis of an investigation on *R. damascena* from Kashan Province, by GC-MS β-citronellol (14.5-47.5%), nonadecane (10.5-40.5%),

geraniol (5.5-18%) and heneicosane (7-14%) were the major components of the oil (6). It has been reported that the main rose oil and its components (citronellol, geraniol and nerol) have strong antimicrobial activity against some bacteria (7-9). In a study by combined use of HPLC, paper chromatography and spectral analysis 285 mg/kg total anthocyanins were isolated from fresh petals. The main anthocyanin was characterized as cyanidin 3, 5-diglucoside and identified compounds were several kaempferol and quercetin glucosides, galactoside, arabinosides, and rhamnosides (10). In another investigation hot water infusions (teas) of air-dried petals of twelve rose cultivars were assayed for antioxidant activity, total phenols, and total anthocyanin contents. The range of total phenols content in rose teas was 50.7 to 119.5 mg gallic acid equivalents (GAE) per gram of dry matter, as compared to 62.1 mg GAE/g dry weight in the green tea. No clear relationship between anthocyanin level and radical-scavenging activity

was revealed (11). In a survey of flower flavonoids in 120 taxa from 10 sections of subgenus *Rosa*, 19 flavonols and six anthocyanins; six kaempferol glycosides, six quercetin glycosides, seven unidentified flavonols, two cyanidin, two peonidin glycosides and two unidentified anthocyanins were detected. (12). There are several reports antioxidant and antimicrobial properties of *Rosa* species (9, 11, 13, 14). The flower of *R. damascena* in Guilan has pleasant smell and taste but it is not used for preparation of Rose water and according to reports of folk uses, its petal has no laxative property (15). This differences between rose of Guilan and other origin sites especially Kashan, may be due to genetic and environmental factors, and there was not any report from essential oil and secondary metabolites of cultivated *Rosa* in Guilan. The aim of this study was to identify components of *R. damascena* extract and essential oil and to determine their biological activity.

MATERIALS AND METHODS

Plant material

Rosa damascena was collected from Rasht (Guilan Province) in May 2004. This species was confirmed by Dr. V. Mozaffarian (Dept. of Botany, Research Institute of Forests and Rangelands, Tehran, Iran) and deposited in herbarium of Pharmacognosy (Voucher No. 6555 TEH), Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran.

Extraction

Fresh petals (400 g) were extracted with methanol and water (80:20). The solvent was evaporated under low temperature and low pressure to give gummy residue (crude extract).

Isolation of the essential oil

The fresh petals of *R. damascena* (500 g) were subjected to hydrodistillation using a Clevenger-type apparatus for 4 hrs and the distillate was dried over anhydrous sodium sulfate and stored at 4 °C in a sealed vial for analysis. Hydrodistillation of petals yielded 0.20% (v/ w) of essential oil. The yield was based on dry materials of petals.

GC/MS analysis

The oil was analyzed by GC/MS using a Hewlett Packard 5973 mass selective detector connected to a HP 6890 gas Chromatograph. The separation was achieved by capillary column, HP-5MS (5% phenyl methyl polysiloxane) (30 m × 0.25 mm, film thickness 0.25 µm). The column temperature was kept at 60°C for 5 min and programmed to 220°C at a rate of 6°C/min. The flow rate of helium (the carrier gas) was 1 mL/min and MS was taken at 70 eV. The relative percentage of the oil

constituents was expressed as percentages by peak area normalization. The identification of individual compounds was based on comparison of their relative retention indices with those of authentic samples on the HP-5MS capillary column and by matching of their mass spectra with those obtained from Wiley libraries and published data (16-18).

Inhibition of lipid peroxidation

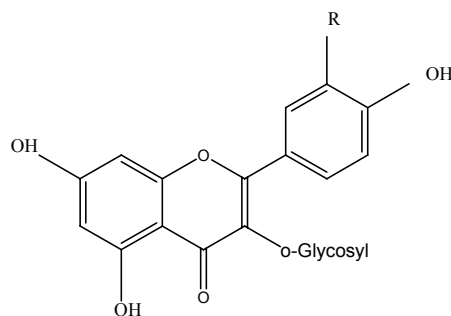
The ferric ammonium thiocyanate method was used for determination of antioxidant activity of petal extract (19, 20). One mL of different concentrations of crude extract in methanol (300, 225 and 150 µg/mL) was mixed with an emulsion of 2.51% linoleic acid in absolute ethanol (0.5 mL) and 0.05 M sodium hydrogen phosphate buffer, pH=7 (1 mL) in a screw capped tube and incubated in an oven at 40°C in the dark. The same mixture without sample extract was used as the control. During incubation (every 24 hrs), 0.1 ml of each tube was diluted with 4.7 mL of 75% ethanol and 0.05 mL of 30% ammonium thiocyanate. Precisely 3 min after addition of 0.05 mL of 0.02 M ferrous chloride in 3.5% hydrochloric acid to the reaction mixture, the absorbance was measured against blank (75% ethanol) at 500 nm every 24 hrs for 96 hrs. The degree of linoleic acid peroxidation was calculated during 96 hrs using the following formula:

$$AA\% = 100 - \left[\frac{\text{absorbance of sample } t}{\text{absorbance of control } t} \times 100 \right]$$

Vitamin E and BHT were included as natural and synthetic antioxidant standards respectively for comparison.

Free radical scavenging

The DPPH method (21) was used with some



Compound	R	-o-Glycosyl
C1	OH	glucose
C2	H	rhamnose
C3	H	arabinose

Figure 1. Structures of C1: quercetine-3-o-glucoside, C2: kaempferol-3-o-rhamnoside and C3: kaempferol-3-o-arabinoside

Table 1. ¹H and ¹³C-NMR (DMSO d₆ + CDCl₃) data of compounds **C1-3** (400MHz for H and 100 MHz for C; δ ppm)^a

positions	C1 (J)	¹³ C	C2, C3 (J)	¹³ C
2	-	149.00	-	146.90
3	-	136.19	-	135.74
4	-	176.19	-	175.84
5	-	160.93	-	160.72
6	6.25, <i>d</i> , (2 Hz)	098.05	6.29, <i>d</i> , (2 Hz)	098.18
7	-	163.67	-	163.89
8	6.40, <i>d</i> , (2 Hz)	093.37	6.39, <i>d</i> , (2 Hz)	093.35
9	-	156.75	-	156.13
10	-	102.56	-	103.01
1'	-	131.72	-	121.95
2'	7.78, <i>d</i> , (2 Hz)	129.52	8.06, <i>d</i> , (8.4 Hz)	115.06
3'	-	115.45	7.06, <i>d</i> , (8.4 Hz)	145.05
4'	-	160.44	-	147.70
5'	6.94, <i>d</i> , (8.4 Hz)	115.45	7.06, <i>d</i> , (8.4 Hz)	115.60
6'	7.64, <i>dd</i> , (8.4, 2 Hz)	129.52	8.06, <i>d</i> , (8.4 Hz)	119.97

^aC₅-OH appeared at 12.35, 12.30 ppm for **C1** and **C2, C3** respectively. Other OH signals appeared between 8.5 -10.5 ppm.

modification in our laboratory. Different samples from hydro-alcoholic extract (90, 60, 30, 3 μg/mL) and essential oil (10, 5, 1 μl/mL) in methanol were added to 2 ml of DPPH solution (4×10⁻⁵ g/mL MeOH). Control consisted of sample which was added to methanol up to 3 mL; Blank consisted of 1 mL methanol which was added to 2 mL of DPPH solution. The absorbance of samples was measured at 517 nm after 30 min. The percentage of inhibition activity was calculated as follows:

Inhibition% = 100 - [(Sample absorption-control absorption)/Blank absorption] × 100.

Statistical analyses

Analyses of at least three samples were carried out in triplicates. Student's *t*-test was carried out to compare the data and results were considered statistically significant at P<0.05. IC₅₀ was calculated with Curve Expert 1.3.

Isolation procedure

Extract (3 g) was submitted to paper chromatography. The chromatograms were developed descending in the long direction of Whatman No. 3 chromatographic papers in the chromatocab using 15% acetic acid as eluent for 5 hrs (22) to obtain four fractions (A, B, C and D).

For further purification, fraction **C** was subjected to paper chromatography with TBA (t-butyl alcohol: acetic acid and water; 3:1:1) as eluent (22) to separate compounds **C1, C2, C3** (16 mg, Rf= 0.57; 54 mg, Rf= 0.71; 12 mg, Rf= 0.48 respectively). Sephadex LH₂₀ was used for the last purification of **C1, C2** and **C3**.

Separation of aglycon and identification of sugars

About 10 mg of each compound was hydrolyzed with 2N HCl and solution was extracted with diethyl ether (aglycon Portion). Aqueous layer was chromatographed on Whatman No. 3 paper chromatography, alongside with some of the more common sugars such as glucose, rhamnose, xylose, arabinose and galactose. The sugars were identified according by literature methods (23).

Identification of aglycons

Compounds **C1** (*Quercetin aglycon*), **C2** and **C3** (*Kaempferol aglycon*) were identified using spectroscopic methods and comparison with authentic samples. ¹H and ¹³C-NMR data are in Table 1.

Free radical scavenging activity of kaempferol-3-O-rhamnoside (C2)

Among isolated flavonoids, since only the amount of kaempferol 3-O-rhamnoside was enough, its antioxidant activity was determined by DPPH method.

RESULTS AND DISCUSSION

Fresh petals of *R. damascena* were collected from north of Iran (Guilan Province). Its essential oil was prepared by hydrodistillation and was analyzed with GC/MS. Seventeen out of twenty one compounds were identified (97.42% v/w). The oil of *R. damascena* was rich of nonterpenoid hydrocarbons comprising 73.38% of the oil. There were not sesquiterpene compounds in the oil. Linalool (3.68%), nerol (3.05%), geraniol (15.5%),

Table 2. Essential oil composition of *R. damascena* from Iran Population of Guilan.

Compounds Name	Percent	^a RRI	^b KI	Method of identification
β-myrcene	0.13	990	991	MS-RRI
linalool	3.68	1092	1097	MS-RRI
phenylethyl alcohol	0.86	1106	1110	MS-RRI
rose oxide	0.1	1125	1127	MS-RRI
nerol	3.05	1234	1230	MS-RRI
geraniol	15.5	1254	1253	MS-RRI
linalyl acetate	0.3	1260	1257	MS-RRI
eugenol	0.18	1365	1359	MS-RRI
geranyl acetate	0.24	1386	1381	MS-RRI
n-heptadecane	2.84	1732	1700	MS-RRI
1-nonadecene	18.56	1445	-	MS
n-octadecane	2.8	1822	1800	MS-RRI
n-eicosane	0.32	1980	2000	MS-RRI
n-heptacosane	2.47	2110	2100	MS-RRI
n-tricosane	16.68	2326	2300	MS-RRI
n-pentacosane	5.11	2545	2500	MS-RRI
n-hexatriacontane	24.6	3552	3600	MS-RRI
Monoterpen hydrocarbons	0.13			
Monoterpen oxygenated	22.87			
Sesquiterpen hydrocarbons	0.00			
Sesquiterpen oxygenated	0.00			
Phenylpropanoids	0.18			
Nonterpenoids	73.38			
Unknown	3.44			
Total identified	97.42			

a: Relative Retention Indices as determined on a HP-5MS column using the homologous of n-alkanes. b: Kovats Indices

1-nonadecene (18.56%), n-tricosane (16.68%), n-pentacosane (5.11%), n-hexatriacontane (24.6%) were the major components of essential oil (Table 2). The oil contained Phenylethyl alcohol (0.86%), rose oxide (0.1%), linalyl acetate (0.3%), eugenol (0.18%) and geranyl acetate (0.24%) that could be observed in essential oils of *R. damascena* from other regions (24–26). Result of investigation on *R. damascena* essential oil from Kashan showed some similarity with oil of the *R. damascena* from Guilan, but there were some variations in amount of their components (6).

The components of petal extracts of the other *Rosa* species (e.g., *R. persica*, *R. damascena*, *R. chinensis* and *R. rugosa*) are closely related and also demonstrated good antioxidant activity (27). There is a report describing that forty landraces of Damask rose were collected from 28 provinces in Iran (13 origin sites Os1- Os13) and evaluated to determine the diversity among them. The cluster analysis confirmed the distinctiveness of the Os12 which included Guilan, Mazandaran, Golestan

provinces with a specific climate condition of humid temperate from other origin sites as a whole. This distinctiveness was already confirmed by the bi-plot of the principal components (28). The results of mentioned report may confirm different properties of cultivated rose in Guilan (North Iran) and other regions of Iran.

In this investigation the rose petals were extracted with 80% methanol and evaporated to dryness under vacuum (45g). Since most investigations show that *Rosa* species have good antioxidant activity, the extract was subjected to evaluation for antioxidant activity using 2,2-diphenyl-1-picrylhydrazyl (DPPH) for measurement of free radical scavenging activity and with ferric ammonium thiocyanate method for evaluation of lipid peroxidation properties. The IC_{50} of the extract with DPPH method and FTC method are reported in Table 3. Based on the statistical analyses, there were significant differences ($P < 0.02$) between results of extract, vitamin E and BHT were; the results showed that the extract has powerful free radical scavenging activity compared

Table 3. IC₅₀ of Samples from *R.damascena* with DPPH and FTC methods.

Samples	FTC/72 h µg/mL ^a	DPPH/30 min µg/mL ^a
Extract	520 ± 9.68	2.24 ± 0.98
essential oil	ND	3.54 µL ± 1.22
kaempferol 3-O-rhamnoside	ND	531 ± 11.02
BHT	153.9 ± 7.24	110.98 ± 8.25
Vitamine E	20.98 ± 2.98	22.72 ± 1.91

^aValues are mean ± SD (n=3). ND: not determined

to standards. Therefore the extract was fractionated by paper chromatography and isolated substances were purified by Sephadex LH20 to obtain three compounds (**C1**, **C2**, **C3**). Isolated compounds were identified by UV, MS and NMR instruments. All these components were glycosides and the results obtained from acidic hydrolysis showed the presence of glucose, rhamnose, and arabinose as the sugar part of each compound respectively. The spectral data of compounds showed good agreement with those of literature data (29, 30) and

confirm the structure of compound **C1** as: 3, 5, 7, 3', 4'-tetrahydroxy flavone glycosylated at position 3 with glucose and the structure of compounds **C2** and **C3** as: 3, 5, 7, 4'-trihydroxy flavone glycosylated at position 3 with rhamnose and arabinose respectively. The three compounds have been reported previously from *R. damascena* and some species of other origin (10, 12, 31). Additionally the antioxidant activity of the essential oil and kaempferol 3-O-rhamnoside were determined by DPPH procedure. The results showed radical scavenging effect of essential oil was stronger than BHA and vitamin E ($P < 0.03$) with IC₅₀ = 3.54 µL ± 1.22. IC₅₀ of kaempferol 3-O-rhamnoside in comparison to IC₅₀ of extract showed weak antioxidant activity. The potential antioxidant activity of extract could be due to existence of quercetin 3-O-glucoside and other flavonoids in extract with more hydroxyl groups in B ring (32).

In conclusion the petal of *Rosa damascena* from Gilan Province has gentle smell without bitter taste, and powerful antioxidant activity. Therefore it can be used as a food flavor, natural antioxidant and a preventing agent for many diseases which are caused by free radicals.

REFERENCES

- Mozaffarian V. A Dictionary of Iranian Plant Names, Farhang Moaser Publication, Tehran, 1995; p. 462
- Groom N. The Perfume Handbook, Blackie Academic Professional, UK, 1997; p. 292.
- Bown D. Encyclopedia of Herbs and their uses, Dorling Kindersley, London, 1995; p. 342.
- Ody Mnimh P. The Herbs Society's Complete Medicinal Herbal, Dorling Kindersley, London, 1995; pp. 162-168.
- Soliman KM, Badeaa RI. Effect of oil extracted from some medicinal plants on different mycotoxigenic fungi. Food Chem. Toxicol.. 2002; 40: 1669-1675.
- Loghmani-Khouzani H, Sabzi Fini O, Safari J. Essential Oil Composition of *Rosa damascena* Mill Cultivated in Central Iran. Scientia Iranica, 2007; 14(4): 316-319.
- Aridogam BC, Baydar H, Kaya S, Demirci M, Ozbasar D, Mumcu E. Antibacterial activity and chemical composition of some essential oils. Arch. Pharm. Res. 2002; 25: 860-864.
- Kivanc M, Akgül A. Antibacterial activities of essential oils from Turkish spices and *Citrus*. Flavour Fragr J. 1986; 1: 175-179.
- Ozkan G, Sagdiç O, Baydar NG, Baydar H. Antioxidant and Antibacterial Activities of *Rosa Damascena* Flower Extracts. Food Sci. Technol. Int. 2004; 10(4): 277-281.
- Velioglu Y.S, Mazza G. Characterization of flavonoids in petals of *Rosa damascena* by HPLC and Spectral Analysis. J. Agric. Food. Chem. 1991; 39: 463-467.
- Vinokur Y, Rodov V, Reznick N, Goldman G, Horev B, Umiei N, Friedman H. Rose Petal Tea as an Antioxidant rich Beverage: Cultivar Effects. Food Sci. 2006; 71(1): 842-847.
- Mikanagi Y, Yokoi M, Ueda Y, Saito N. Flower Flavonol and Anthocyanin Distribution in Subgenus *Rosa*. Biochem. Sys. Ecol. 1995; 23(2): 183-200.
- Ng TB, Gao W, Li L, Niu SM, Zhao L, Liu J, Shi LS, Fu M, Liu F. Rose (*Rosa rugosa*)-flower extract increases the activities of antioxidant enzymes and their gene expression and reduces lipid peroxidation. Biochem. Cell Biol. 2005; 83: 78-85.
- Jassbi AR, Zamanizadehnajarib S, Tahara S. Polyphenolic antioxidant constituents of *Rosa persica*. J. Chem. Soc. Pakistan. 2003; 25: 323-327.
- Zargari A. Medicinal plants. Vol 2. 5th ed. Tehran University Press, Tehran 1992; 281-284.
- Massada Y. Analysis of Essential oil by GC and spectrometry. Wiley, New York. 1976.
- Adams RP. Identification of essential oils component by Gas Chromatography/ Mass Spectroscopy. Allured Publishing Corporation, Illinois. 2001; P. 469.
- Lawrence BM, Mookherjee BD and Willis BJ. Flavours and Fragrances. A World Prospective, Development in Food Science 18, Elsevier Science Publisher, Amsterdam. 1988.

19. Shrififar F, Yassa N, Shafiee A. Antioxidant activity of *Otostegia persica* (Labiatae) and its constituents . Iran J. Pharm. Res. 2003; 235-239.
20. Yassa N, Sharififar F, Shafiee A. *Otostegia persica* as a Source of Natural Antioxidants Pharmaceutical Biology. 2005; 43: 33–38.
21. Sanchez-Moreno C, Larrauri J. A, Souara-Calixto F. Free radical scavenging capacity and inhibition of lipid peroxidation of wines, grape juices and related polyphenolic constituents. Food Res. Int. 1999; 32: 407-412.
22. Mabry TJ, Markham KR., Thomas MB. The Systematic Identification of Flavonoids. Springer -Verlag, Berlin. 1970.
23. Wagner H and Bladt S. Plant Drug Analysis, 2nd ed. Springer, Berlin. 1996.
24. Bayrak A, Akgul A. Volatile oil composition of Turkish rose (*Rosa damascena*). J. Food Sci. Agri. 1994; 64: 441-448.
25. Jirovets L, Buchbauer G, Stoyanova A, Balinova A, Guangjun Z, Xihan MA. Solid phase microextraction (gas chromatograph and olfactory analysis of the Scent and fixative properties of essential oil of *Rosa damascena* L. from China. Flavour Fragr J. 2004; 20: 7-12.
26. Babu KGD, Singh B, Joshi VP, Singh V. Essential oil composition of Damask rose (*Rosa damascene* Mill.) distilled under different pressures and temperatures. Flavour Fragr. J. 2002; 17: 136-140.
27. Cai YZ, Xing J, Sun M, Zhan ZQ, Corke H. Phenolic Antioxidants Hydrolyzable Tannins, Flavonols, and Anthocyanins) Identified by LC-ESI-MS and MALDI-QIT-TOF MS from *Rosa chinensis* Flowers. J. Agricult. Food Chem. 2005; 53(26): 9940-9948.
28. Tabaei-Aghdai SR, Babaei A, Khosh-Khui M, Jaimand K, Rezaee MB, Assareh MH, Naghavi MR. Morphological and oil content variations amongst Damask rose (*Rosa damascena* Mill.) landraces from different regions of Iran. Scientia Hort. 2007; 113(1): 44-48.
29. Moco S, de Vos R, Beek TV, Bino R, Vervoort J. (2007): Metabolic profiling and identification in Tomato by LC/UV/MS/NMR. Available at <http://www.dpw.wau.nl/pf/metabolomics/moco-mw05.pdf>.
30. Yang O, Fu Hong Z, Lei H, Hin WH and Ma GE. Chemical constituent from the leaves of *Koelreuteria paniculata* Laxm. Pharm. J. 1999; 34 (6): (from internet).
31. Chang YC, Chang FR, Yung-Chang WU. The Constituent of *Lindera glauca*. J. Chinese Chem. Soc. 2000; 47: 373- 380.
32. Pier-Giorgio P. Flavonoids as Antioxidants. J. Nat. Prod. 2000; 63: 1035-1042.

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