لینک های مفید

- عضویت در خبرنامه
- کارگاه‌های آموزشی
- سرویس ترجمه تخصصی STRS
- فیلم‌های آموزشی
- بلاگ مرکز اطلاعات علمی
- سرویس‌های ویژه

40% تخفیف به مناسبت سالروز تاسیس مرکز اطلاعات علمی
Optimization of Hydrolysis Condition of Pumpkin Seeds with Alcalase Enzyme to Achieve Maximum Antioxidant and Nitric Oxide Inhibition Activity

Kosar Zakeri¹, Alireza Sadeghi Mahoonak²*, Mohammad Ghorbani¹, Ali Moayed³, Yahya Maghsodlou⁴

1- Master of Science (MSc), Department of Food Sciences and Technology, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, Iran
2- Associate Professor, Department of Food Science and Technology, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, Iran
* Corresponding author (sadeghiaz@gau.ac.ir)
3- Assistant Professor, Department of Food Science and Technology, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, Iran
4- Professor, Department of Food Science and Technology, Gorgan University of Agricultural Sciences and Natural Resources, Gorgan, Iran

Abstract
In this research, optimization of pumpkin (Cucurbita pepo) hydrolysis condition was investigated in order to achieve maximum 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging and nitric oxide inhibition activity using Design Expert software and response surface method. For this purpose, hydrolysis conditions including concentration of alcalase enzyme were selected within 0.7-3.30%, temperature 32-58 °C and hydrolysis time 30-290 min as independent variable. The results showed that optimum hydrolysis conditions to achieve the maximum DPPH radical scavenging and nitric oxide inhibition activity were temperature of 44 °C, hydrolysis time of 260 min, and enzyme to substrate concentration of 3% which under this condition the DPPH radical scavenging and nitric oxide inhibition activity of hydrolyzate was 72.03 and 89.34%, respectively that was largely similar to the results proposed by the software (75.33 and 84.71%). According to the results, pumpkin seed protein hydrolyzate showed high antioxidant and nitric oxide inhibitory properties and can be used as a suitable ingredient in food formulations.

Keywords
Alcalase
Antioxidant
Enzyme Hydrolysis
Nitric Oxide Inhibition
Pumpkin Seeds

Received: 12.03.2018
Accepted: 14.11.2018

Introduction
Proteins are an important source of nutrition for humans. These compounds provide nitrogen, amino acids and energy for the normal function of the body systems. Hydrolysis acts to break down proteins into free amino acids and peptides of a different size. The most common way of producing peptides is to hydrolyze proteins with the help of enzymes (Korhnen & Pihlanto, 2006). Nowadays, bioactive peptides derived from the hydrolysis of various dietary proteins are known to have various biological roles such as antihypertensive (Li et al., 2007), relaxing (Ohinata et al., 2007), anticholesterol (Cho et al., 2008), antioxidant (Kou et al., 2013) and anti-
cancer (Meisel & FitzGerald, 2003), which has led to the consideration of proteins, especially plant types, in the food and drug sector for the production and purification of bioactive peptides. One of the properties of these peptides is inhibition of nitric oxide, which acts as an intermediate compound in the production of nitrosamines. Nitrate and nitrite salts, which are used as an additive to stabilize color and inhibit the growth of some microorganisms in meat products, are very strong oxidants. As a result of the reaction of nitrite with the secondary and tertiary amines, the nitrosamine carcinogen compound is formed; if there is an antioxidant compound in processed meat, it reacts much more rapidly than amines with nitrite and prevents from the formation of nitrosamine (Hauser et al., 1980; Coss et al., 2004).

Pumpkin seeds (Cucurbita pepo) are known as a rich source of protein and contain significant amounts of minerals such as zinc, potassium, calcium, magnesium, iron, copper, phosphorus and β-carotene. The protein isolated from pumpkin is rich in antioxidants and is effective in reducing the malignant effects of malnutrition (Mohamed et al., 2009). Nourmohammadi et al. (2016) hydrolyzed the protein of pumpkin seed meal using pepsin enzyme using three variables of temperature, time and enzyme concentration. The results showed that produced peptides had more antioxidant power in hydrolysis conditions including 1% concentration of enzyme, temperature of 30 °C and hydrolysis time of 2 h, compared to other treatments obtained from the response surface method and central composite designs. Piri Ghashlaghi et al. (2015), Mehregan Niko et al. (2013) and Etemadi et al. (2014) respectively produced hydrolyzed proteins from whey protein concentrate, crucian carp and soybean meal by alcalase enzyme and the effect of temperature, time, and the ratio of enzyme to substrate on antioxidant activity was investigated.

The results of various experiments have shown that the hydrolyzed proteins derived from the protein hydrolysis of fish (Khattaphant & Benjakul, 2008), the hydrolyzed protein of tracheal meat (Klompong et al., 2009), soybean protein (Chen et al., 1995), milk proteins such as α- lactalbumin and β-lactoglobulin (Hernandez-Ledesma et al., 2005), egg white protein (Davalos et al., 2004) and gluten protein (del Castillo et al., 2007) showed high antioxidant activity. Lee et al. (2012) hydrolyzed the proteins derived from oysters (Ruditapes philippinarum) by 8 protease enzymes and ultimately obtained peptides with the highest nitric oxide inhibition property by the alcalase enzyme.

The pumpkin seeds are commonly used in the production of oil. Meal obtained from pumpkin seed oil contains high percentage of protein, which is currently not processed and is a cheap source of protein for the consumption in animal feed. Considering the high potential of this material in the production of bioactive peptides, this study aimed to optimize the enzymatic hydrolysis of pumpkin meal by alcalase enzyme in order to produce protein hydrolyzate with the highest 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging and nitric oxide inhibition activity.

**Material and methods**

**Materials**

Pumpkin seed meal was purchased from Abkar Gorgan Company. The alcalase enzymes and DPPH radical obtained from Sigma and NaOH, hydrochloridric acid, sulfonilamide, phosphoric acid, sodium nitroprusside and ethanol from Merck, and naphthyl ethylenediamine dihydrochloride were obtained from Ridley Company, Germany. All materials...
used were of laboratory grade.

**Preparation of pumpkin seed meal**
After removing the foreign material, the meal was turned into flour by milling machine (Perten, 3100, Germany) and passed through a sieve with 30 meshes. The resulting flour was defatted using hexane with the ratio of 1:3 for 16 h, and after that it was kept at room temperature for 48 h in order to remove the solvent residues completely, and finally kept in the refrigerator (Kaur & Singh, 2007).

**Pumpkin meal flour**
The defatted meals were ground at this stage by a grinding machine (Perten, 3100, Germany) so that the flour particles could pass through the sieve with 70 meshes. Samples were stored in sealed polyethylene bags at temperature of 4 °C in the refrigerator (Glew et al., 2006).

**Preparation of pumpkin protein concentrate**
The defatted pumpkin seed meal was dispersed in water with the ratio of 1 to 10 and then the pH of the solution was adjusted to 10 by 1 N NaOH and mixed at room temperature for 1 h. The resulting mixture was centrifuged at 5000g using a refrigerated centrifuge (Combi 514R, South Korea) for 20 min at the temperature of 4 °C. In order to precipitate pumpkin seeds proteins, the pH resulting supernatant was adjusted to pH=5 by 1 N HCl, and centrifuged under similar condition. Then the precipitate obtained was washed with distilled water and dried using a freezing dryer (FD4 model, Operon Company, South Korea) and then kept at freezer at temperature -18 °C for further experiments (Zivanovic et al., 2011).

**Hydrolysis of pumpkin seed protein concentrate**
Protein concentrate with the proportion of 5% (w/v) was dispersed in a tris-hydrochloridic acid buffer with pH=8 (Optimal pH of alcalase enzyme) and the enzyme was added at concentration of 1-3%. Then, the hydrolysis was carried out in the temperature range of 35-55 °C and time of 60-260 min in a shaking incubator (VS-8480 model, South Korea) with speed of 200 rpm. Finally the enzymatic reaction was stopped at 85 °C for 10 min, and the resulting mixture was centrifuged at the 5000g for 30 min to remove the insoluble compounds (Villanueva et al., 1999). The resulting supernatant was kept at the temperature of -18 °C and freeze dried.

**Determination of chemical composition of pumpkin meal**
Moisture, ash and protein were determined using the AACC methods of 15-44, 01-08 and 12-46, respectively (AACC, 1999) and the fat content was measured by soxhlet method (Parvane, 2006).

**Measurement of DPPH radical scavenging activity**
1000 μL of hydrolyzed protein was mixed with 1000 μL of DPPH (0.1 mM) prepared in 96% ethanol and kept at room temperature in the dark for 60 min and finally the absorbance of solution was measured at a 517 nm using spectrophotometer (Bougatef et al., 2009). In the control sample, 1000 μL of distilled water was used instead of the hydrolyzed protein sample. The DPPH radical inhibition activity was calculated as Eq. (1):

\[
\text{DPPH radical scavenging activity} = \frac{\text{absorption of control-sample absorption}}{\text{absorption of control}} \times 100
\]

**Measurement of nitric oxide inhibition activity**
60 μL of hydrolyzed protein was mixed with 60 μL of sodium nitroprusside in phosphate buffer (0.025 M) and placed
in the plates and kept in the incubator for 150 min at room temperature (ascorbic acid was used as a control sample). Then, the equal amount (120 μL) of Grease reagent (including sulfanilamide, naphthyl ethylene dihydrochloride and phosphoric acid) was added to plates and the absorbance was measured at 546 nm (Tsai et al., 2007) and the inhibition activity was calculated as Eq. (2):

\[
\text{Nitric oxide inhibitory percent} = \left[ \frac{\text{absorption of control} - \text{sample absorption}}{\text{absorption of control}} \right] \times 100
\]

Optimization of enzymatic hydrolysis conditions
In order to optimize the process from the viewpoint of maximizing the antioxidant and nitric oxide inhibition properties, Design Expert software version 11 and the response surface method with central composite designs were used for 3 independent variables of time (X₁), temperature (X₂) and the ratio of enzyme to substrate (X₃) at 5 levels (-α, -1, 0, +1 and +α). Approximate amount of each independent variable was obtained according to the experimental tests results. The examined responses were DPPH radical scavenging and nitric oxide inhibition activity. Total 20 treatments were randomly selected by the software with six replications at the central point. The different levels of the independent variables and the relevant treatments are presented in Tables (1) and (2), respectively.

Table 1. Levels of independent variables used to optimize DPPH radical scavenging activity and nitric oxide inhibitory activity of pumpkin seeds protein

<table>
<thead>
<tr>
<th>Independent variables</th>
<th>The limits of change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hydrolysis time (min)</td>
<td>-α 60 160 260 290</td>
</tr>
<tr>
<td>Hydrolysis temperature (°C)</td>
<td>-α 32 45 55 58</td>
</tr>
<tr>
<td>The ratio of enzyme to substrate (%)</td>
<td>-α 0.70 2 3 3.30</td>
</tr>
</tbody>
</table>

The value of α is equal to 1.3.

Table 2. Random treatments and DPPH radical scavenging and nitric oxide inhibitory activity of the hydrolyzed pumpkin seed protein

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Time (min)</th>
<th>Hydrolysis temperature (°C)</th>
<th>The ratio of Enzyme to Substrate (%)</th>
<th>DPPH radical scavenging activity (%)</th>
<th>Nitric oxide inhibitory activity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>60</td>
<td>35</td>
<td>3</td>
<td>67.09</td>
<td>80.83</td>
</tr>
<tr>
<td>2</td>
<td>160</td>
<td>45</td>
<td>2</td>
<td>79.01</td>
<td>72.71</td>
</tr>
<tr>
<td>3</td>
<td>160</td>
<td>45</td>
<td>2</td>
<td>79.73</td>
<td>77.29</td>
</tr>
<tr>
<td>4</td>
<td>160</td>
<td>32</td>
<td>2</td>
<td>74.06</td>
<td>86.65</td>
</tr>
<tr>
<td>5</td>
<td>60</td>
<td>55</td>
<td>1</td>
<td>78.64</td>
<td>58.89</td>
</tr>
<tr>
<td>6</td>
<td>160</td>
<td>58</td>
<td>2</td>
<td>77.21</td>
<td>80.06</td>
</tr>
<tr>
<td>7</td>
<td>260</td>
<td>35</td>
<td>3</td>
<td>67.48</td>
<td>85.96</td>
</tr>
<tr>
<td>8</td>
<td>160</td>
<td>45</td>
<td>0.70</td>
<td>77.02</td>
<td>83.59</td>
</tr>
<tr>
<td>9</td>
<td>260</td>
<td>55</td>
<td>3</td>
<td>76.49</td>
<td>84.89</td>
</tr>
<tr>
<td>10</td>
<td>160</td>
<td>45</td>
<td>3.30</td>
<td>81.11</td>
<td>85.73</td>
</tr>
<tr>
<td>11</td>
<td>60</td>
<td>55</td>
<td>2</td>
<td>80</td>
<td>85.27</td>
</tr>
<tr>
<td>12</td>
<td>290</td>
<td>45</td>
<td>2</td>
<td>72.42</td>
<td>87.27</td>
</tr>
<tr>
<td>13</td>
<td>160</td>
<td>45</td>
<td>2</td>
<td>80</td>
<td>77.98</td>
</tr>
<tr>
<td>14</td>
<td>260</td>
<td>55</td>
<td>1</td>
<td>70.32</td>
<td>87.04</td>
</tr>
<tr>
<td>15</td>
<td>60</td>
<td>35</td>
<td>1</td>
<td>48.46</td>
<td>85.43</td>
</tr>
<tr>
<td>16</td>
<td>260</td>
<td>35</td>
<td>1</td>
<td>55.89</td>
<td>88.19</td>
</tr>
<tr>
<td>17</td>
<td>160</td>
<td>45</td>
<td>2</td>
<td>79</td>
<td>74.32</td>
</tr>
<tr>
<td>18</td>
<td>160</td>
<td>45</td>
<td>2</td>
<td>69.88</td>
<td>74.46</td>
</tr>
<tr>
<td>19</td>
<td>30</td>
<td>45</td>
<td>2</td>
<td>75.63</td>
<td>88.57</td>
</tr>
<tr>
<td>20</td>
<td>160</td>
<td>45</td>
<td>2</td>
<td>81.45</td>
<td>77.13</td>
</tr>
</tbody>
</table>
The regression models 1 and 2 were proposed to predict the first and second responses (DPPH radical scavenging and nitric oxide inhibition activity) by the following Eq. (3 and 4):

\[ R_1=b_0+b_1x_1+b_2x_2+b_3x_3+b_1b_2x_1x_2+b_1b_3x_1x_3+b_2b_3x_2x_3+b_1b_2b_3x_1x_2x_3 \]

\[ R_2=b_0+b_1x_1+b_2x_2+b_3x_3+b_1b_2x_1x_2+b_1b_3x_1x_3+b_2b_3x_2x_3+b_1b_2b_3x_1x_2x_3 \]

In above Eq. (3 and 4), \( R_1 \) and \( R_2 \) are the responses or dependent variables (DPPH radical scavenging and nitric oxide inhibition activity, respectively), \( b_0 \) is a constant, and \( b_1, b_2 \) and \( b_3 \) are linear effects, \( b_{11}, b_{22} \) and \( b_{33} \) are quadratic effects and \( b_{12}, b_{13} \) and \( b_{23} \) are interaction effects.

Analysis of variance, regression coefficients, plotting and optimization were performed by Design Expert software and the significance of the tests were analyzed by Duncan multiple range test and SPSS V16 software at a significant probability level of \( P<0.05 \).

**Result and discussion**

According to the results of Table (3), the pumpkin seed protein concentrate contain high protein content. The extraction process with the help of alkali led to the separation of non-protein parts from the meal and the accumulation of protein in the remainder section (Khartaphant et al., 2011a). The amount of fat in the concentrate was lower than the fat in the meal. The reason of reducing the fat content of the concentrate compared to the meal was the defatting process done on the pumpkin seed meal before preparation of the concentrate and the process of extracting the protein in alkaline solution and precipitation at the isoelectric point (Khartaphant et al., 2011b). The amount of ash in the meal and protein concentrate was 7.63 and 0.815, respectively, which is equivalent to the values reported by Nourmohammadi et al. (2016) and Mazloomi et al. (2017). The moisture content of the meal and protein concentrate was 7.03 and 12.56, respectively. The moisture content of protein concentrate was higher than that of the above mentioned sources, and the reason of this matter can be its long-term preservation that causes some moisture absorption.

**Table 3. Chemical composition of meal and protein concentrate prepared from pumpkin seed**

<table>
<thead>
<tr>
<th>Matter</th>
<th>Protein</th>
<th>Fat</th>
<th>Moisture</th>
<th>Ash</th>
</tr>
</thead>
<tbody>
<tr>
<td>Defatted Meal</td>
<td>51.8±1.19</td>
<td>9.65±0.15</td>
<td>7.03±0.09</td>
<td>7.63±0.02</td>
</tr>
<tr>
<td>Protein Concentrate</td>
<td>76±1</td>
<td>1.29±0.46</td>
<td>12.56±0.01</td>
<td>0.815±0.015</td>
</tr>
</tbody>
</table>

* Results are the average of three replicates.

Linear Eq. (5) and (6), taking into account the regression coefficients for the DPPH radical scavenging activity and nitric oxide inhibition property, respectively, which were suggested by the following factors according to the significance of coefficients (table 4).

DPPH radical inhibition activity = \( +79.33 \times (0.72 \times \text{time}) + (6.21 \times \text{temperature} + (3.78 \times \text{the ratio of enzyme to substrate}) + (2.46 \times \text{time} \times \text{temperature}) - (0.28 \times \text{the ratio of enzyme to substrate}) + (2.84 \times \text{the ratio of enzyme to substrate}) \)

(5)

Nitric oxide inhibition activity = \( +76.92 + (0.61 \times \text{time}) - (0.52 \times \text{temperature}) - (0.60 \times \text{the ratio of enzyme to substrate}) - (0.89 \times \text{time} \times \text{temperature}) + (0.10 \times \text{the ratio of enzyme to substrate}) + (0.20 \times \text{the ratio of enzyme to substrate}) + (4.79 \times \text{time} \times \text{temperature}) + (2.09 \times \text{temperature} \times \text{time}) + (2.86 \times \text{the ratio of enzyme to substrate}) \)

(6)
The value of $R^2$ was calculated 0.7811 for the first equation (Eq. 5) and 0.7365 for the second equation (Eq. 6), which indicates the relatively suitable distribution of the data. The lack of fitness, which is a criterion for the suitability of the presented model, was 0.1625 and 0.0521 in the first and second equations (Eq. 5 and 6), respectively. The high level of lack of fitness compared to the significant level of probability (95%) or, in other words, the non-significant of this factor indicates the appropriateness of the proposed model and fitness of the model based on the considered responses.

| Table 4. Analyze of variance related to DPPH radical scavenging and nitric oxide inhibition test |
|-----------------------------------------------|-----------------------------------------------|
| Factor                                      | Degree of freedom | Sum of squares | Average of squares | F-Value | P-Value |
| DPPH radical scavenging activity             |                  |                |                    |         |         |
| Model                                        | 9                | 1101.52        | 122.39             | 3.97    | 0.0214  |
| Time                                         | 1                | 5.88           | 5.88               | 0.19    | 0.6717  |
| Temperature                                  | 1                | 438.30         | 438.30             | 14.20   | 0.0037  |
| Enzyme concentration                         | 1                | 162.98         | 162.98             | 5.28    | 0.0444  |
| Time×Temperature                             | 1                | 48.27          | 48.27              | 1.56    | 0.2396  |
| Time×Enzyme                                 | 1                | 0.62           | 0.62               | 0.020   | 0.8900  |
| Temperature×Enzyme                           | 1                | 64.35          | 64.35              | 2.08    | 0.1793  |
| Time×Time                                    | 1                | 148.23         | 148.23             | 4.80    | 0.0532  |
| Temperature×Time                             | 1                | 94.25          | 94.25              | 3.05    | 0.1111  |
| Enzyme×Temperature                           | 1                | 19.82          | 19.82              | 0.64    | 0.4415  |
| Residue                                      | 10               | 308.66         | 30.87              |         |         |
| Non fitness                                  | 5                | 222.000        | 44.40              | 2.56    | 0.1625  |
| Net error                                    | 5                | 86.66          | 17.33              |         |         |
| Total                                        | 19               | 1410.19        |                    |         |         |
| Nitric oxide inhibition                      |                  |                |                    |         |         |
| Model                                        | 9                | 367.82         | 40.87              | 3.11    | 0.0460  |
| Time                                         | 1                | 4.27           | 4.27               | 0.32    | 0.5815  |
| Temperature                                  | 1                | 3.05           | 3.05               | 0.23    | 0.6408  |
| Enzyme concentration                         | 1                | 4.08           | 4.08               | 0.31    | 0.5896  |
| Time×Temperature                             | 1                | 6.34           | 6.34               | 0.48    | 0.5035  |
| Time×Enzyme                                 | 1                | 0.088          | 0.088              | 6.704E-003 | 0.9364 |
| Temperature×Enzyme                           | 1                | 2.06           | 2.06               | 0.16    | 0.7006  |
| Time×Time                                    | 1                | 154.05         | 154.05             | 11.71   | 0.0065  |
| Temperature×Time                             | 1                | 29.34          | 29.34              | 2.23    | 0.1662  |
| Enzyme×Temperature                           | 1                | 55.00          | 55.00              | 4.18    | 0.0681  |
| Residue                                      | 10               | 131.57         | 13.16              |         |         |
| Non fitness                                  | 5                | 109.43         | 21.89              | 4.94    | 0.0521  |
| Net error                                    | 5                | 22.14          | 4.43               |         |         |
| Total                                        | 19               | 499.39         |                    |         |         |

The effect of independent variables on DPPH radical scavenging activity
In this section, the effect of temperature, hydrolysis time and enzyme to substrate ratio on the DPPH free radicals scavenging activity has investigated. Therefore, using the 3D-graphs obtained by Design Expert software, the effect of mentioned parameters on the antioxidant property was investigated and interpreted and the results were compared and evaluated with the results of previous studies. It should be noted that in each figure (1, 2 and 3) the effects of the two parameters are investigated when the third parameter was at the optimal point.

The effect of enzyme concentration and hydrolysis temperature on DPPH radical scavenging activity is shown in Figure (1). As it can be seen, the increase in enzyme concentration and
also increase in hydrolysis temperature led to increase in DPPH radical scavenging activity, but the increase in enzyme concentration was less effective than temperature increase in DPPH radical scavenging activity. Therefore, it can be said that the increase of these two parameters has a direct effect on the increase of the amount of free radical inhibition; thus the highest free radical inhibitory power was achieved at temperature of 55 °C and enzyme concentration of 3%.

The DPPH radical scavenging activity is used to check the hydrogenation capability of hydrolyzed proteins. Free radical removal is a mechanism by which antioxidant compounds are capable to prevent oxidative reactions. DPPH radical is one of the few stable radicals at room temperature. When this compound is exposed to a hydrogen donator compound like an antioxidant, it accepts a hydrogen, converts to a stable compound and after radical inhibition, a notable color change from purple to yellow and absorption reduction at 517 nm can be observed (Taheri et al., 2011). The type of initial material, the specificity of the enzyme, the hydrolysis conditions, and the size, amount and structure of amino acids and peptides produced are the factors affecting the antioxidant activity (Mehregan Niko et al., 2013). On the one hand, the antioxidant power of hydrolyzed materials depends highly on the structural arrangement and specific amino acid sequences of the peptide chain. Some researchers believe that histidine, hydrophobic amino acids, and peptides with sequences of proline-histidine-histidine have anti-oxidant potency (Phelan et al., 2009).

The effect of enzyme concentration and hydrolysis time on the DPPH radical scavenging activity is shown in Figure (2). According to Figure (2), the amount of DPPH free radical scavenging activity increased with increasing enzyme concentration, while with increasing hydrolysis time up to 160 min, the scavenging activity reached its highest level and then, by increasing the time, the amount of free radical scavenging activity decreased. Reduction of DPPH radical scavenging activity by hydrolyzed protein with increasing the hydrolysis time can be due to the progression of the hydrolysis and the greater effect of the enzyme on the protein substance, which causes breaking the chain of some antioxidant peptides formed in the early stages of hydrolysis (Taheri et al., 2011). After optimization of production of hydrolyzed protein from tuna fish waste using the response surface method, Ovissipour et al. (2009) reported that with increasing hydrolysis time, hydrolysis intensity reduced and after that remained constant. Guerard et al. (2002) stated that by increasing the concentration of enzyme, the production of antioxidant peptides increases. They also stated that by increasing hydrolysis time, the degree of hydrolysis increases, and after that the intensity and speed of hydrolysis reduced due to the reduction of the available peptide bands to the enzyme, reduction in the enzyme activity and the formation of inhibitors compounds.
According to the results, it can be said that the hydrolysis temperature and the concentration of the alcalase enzyme have a direct effect on the DPPH radical scavenging activity of hydrolysed product, thereby by increasing these two parameters the antioxidant activity can be increased. However, the increase of hydrolysis time up to a certain limit (160 min) can cause an increase in antioxidant activity of pumpkin seed protein; but with the excessive increase in hydrolysis time, the intensity of hydrolysis increases and antioxidant peptides can be degraded, therefore the antioxidant activity decreases (Taheri et al., 2011).

The effect of independent variables on nitric oxide inhibitory activity
In this section, as in the previous section, the effect of temperature, hydrolysis time and alcalase enzyme concentration on the nitric oxide inhibition capability was investigated. Therefore, in the same way as previously mentioned, using the three-dimensional graphs obtained by Design Expert software, the effect of hydrolysis parameters on the nitric oxide inhibitory power were investigated and interpreted. It should be noted that in each Figure (4, 5 and 6) the effects of the two parameters are investigated when the third parameter is at its optimal point. The effect of enzyme concentration and hydrolysis temperature on the nitric oxide inhibition property is presented in Figure (4). According to the Figure (4), a decrease in nitric oxide inhibition activity has been observed at lower temperatures (35 to 45 °C) and increase in the enzyme concentration up to 2%, and then, with an increase in temperature and enzyme concentration, the amount of inhibition activity increased with a moderate slope. Thus, the lowest nitric oxide inhibition activity was observed at temperature of 45 °C and the enzyme concentration of 2%. Thus, according to the graph, the
The highest nitric oxide inhibition activity (87%) is observed at temperature close to 35 °C and the enzyme concentration of 1%. The nitric oxide inhibitory property depends on the antioxidant property of that compound. The antioxidant property of peptides is related to the selectivity of the protease used, the degree of hydrolysis, the nature of released peptides (such as molecular weight, amino acid sequence and peptide structure), and other characteristics, such as the ability to bond to free radicals, the activity of metal ions, chelation and electron donating power, the structural arrangement and sequence of amino acids and the presence of specific amino acids in their peptide chains. The nitric oxide inhibition property also depends on these factors. The highest nitric oxide inhibition activity observed at the lowest temperature and concentration of pepsin enzyme (35 °C and 1%), but the increase in enzyme concentration (at a certain limit 2%) caused higher enzymatic activity and consequently further breaking of antioxidant and nitric oxide inhibitor peptides occur; so the nitric oxide inhibition activity decreases.

Figure 4. The effect of enzyme concentration and hydrolysis temperature on the nitric oxide inhibition activity at optimum hydrolysis time

The effect of enzyme concentration and hydrolysis time on the nitric oxide inhibition property (figure 5) shows that in the initial hydrolysis times, with an increase in enzyme concentration up to 2%, the nitric oxide inhibition activity shows a descending trend and by increasing hydrolysis time (after 160 min) the nitric oxide inhibition activity increases. The graph shows that the lowest nitric oxide inhibitory activity (81%) achieve at the hydrolysis time of 160 min and enzyme concentration of 2%, and the highest nitric oxide inhibitory activity observed at the hydrolysis time of 260 min and enzyme concentration of 1%. Lee et al. (2012) hydrolyzed the protein derived from oyster by 8 protease enzymes and ultimately peptides with the highest nitric oxide inhibition property obtained by the alcalase enzyme. The hydrolyzed product was then purified using high performance liquid chromatography (HPLC) and finally it was recognized that nitric oxide inhibitory peptide show a sequence of glutamine-cysteine-glutamine-glutamine-alanine-valine-glutamine-serine-alanine-valine at their N terminal section. Therefore, considering the mentioned findings in this section and the research carried out by Lee et al. (2012), it can be concluded that by increasing the hydrolysis time, enzyme activity and degree of hydrolysis increases and eventually the peptides with specific sequences obtain that results an increase in the nitric oxide inhibition activity of hydrolyzed products.

Figure 5. The effect of enzyme concentration and hydrolysis time on the nitric oxide inhibition activity at optimum temperature

Figure (6) shows the effect of hydrolysis time and temperature on the
nitric oxide inhibition activity. In the initial times (before 160 min) and by increasing the temperature up to 45 °C, the nitric oxide inhibition activity shows a descending trend, and by increasing the hydrolysis time to 160 min and temperature to 45 °C, the nitric oxide inhibition show an ascending trend. Therefore, the maximum nitric oxide inhibition activity can be achieved at hydrolysis time of 260 min and temperature of 35 °C. Low nitric oxide inhibition activity of hydrolyzed samples in the early stages of hydrolysis (before time of 160 min) may be due to low hydrolysis time that is insufficient to affect the substrate and to produce peptides with nitric oxide inhibitory activity. With the increase in the hydrolysis time and decrease in the size of produced peptides, the ability to inhibit free radicals by peptides enhances (Nourmohammadi et al. 2016).

Phelan et al. (2014) through enzymatic hydrolysis of milk proteins achieved a peptide with amino acid sequence of alanine-valine-proline-tyrosine-proline-glutamine-arginine and tyrosine-tyrosine-alanine-lysine-proline-alanine-alanine-valine-arginine with proper nitric oxide inhibition property.

Also, Kim et al. (2013) hydrolyzed the protein of shellfish with 8 different protease enzymes and investigated the nitric oxide inhibition activity of final products. Then, by purification using chromatography system they reported that the peptide which showed high nitric oxide inhibitory property contained 10 amino acids with sequence of glycine-valine-serine-leucine-leucine-glutamine-glutamine-phenylalanine-phenylalanine-leucine. According to the findings for achieving proper nitric oxide radical inhibitory activity of final product, the optimum hydrolysis condition include hydrolysis time of 260 min, temperature 35 °C and enzyme concentration of 1%.

Optimization and validation of the model
Optimal conditions were obtained by Design Expert software. Hydrolysis conditions for preparation of hydrolyzed protein with the highest DPPH radical scavenging activity and nitric oxide inhibition activity were obtained in accordance with temperature of 44 °C, hydrolysis time of 260 min and enzyme to substrate concentration of 3%, which corresponded to the nitric oxide inhibition activity and DPPH free radical scavenging activity of 4.71 and 75.33%, respectively. In order to confirm the suitability of conditions proposed by the software, additional experiments were carried out in the predicted situations obtained by the model. The DPPH free radical scavenging activity and nitric oxide inhibition activity of hydrolysed product were obtained 72.03 and 89.34%, respectively. The obtained results showed almost similar values with the predicted values proposed by the model, which indicated using predictive models it is possible to predict optimal conditions for the production of hydrolyzed protein with high antioxidant and nitric oxide inhibition activity from pumpkin seeds protein. Nourmohammadi et al. (2016) stated that the measured antioxidant activity of peptides produced using enzymatic hydrolysis of pumpkin seeds by pepsin at optimal conditions (82.07%) was largely similar to that
proposed by software (80.31%).

Conclusion

Millions of tons of valuable by-products are produced every year during processing of food products. One of these valuable by-products is pumpkin seed meal, which has so far been used as supplement in animal feed. In order to increase the value of this by product which is economically important and due to its high protein content with the appropriate combination of amino acid, this study attempted to use the meal as a suitable protein source for preparation of protein hydrolyzate with high bioactive potential. In this study, optimization of production of hydrolyzed protein with high antioxidant and nitric oxide inhibition activity using alcalase enzyme was performed by response surface method. According to the results, preparation of pumpkin seeds protein hydrolyzate with high antioxidant and nitric oxide inhibition activity is influenced by reaction conditions including temperature, time and the enzyme concentration. Results showed that optimum conditions to achieve maximum DPPH radical scavenging activity and nitric oxide inhibition activity is temperature of 44 °C, hydrolysis time 260 min, and enzyme to substrate concentration of 3%. Under these conditions the antioxidant and nitric oxide inhibition property of protein hydrolyzate were 72.03 and 89.34%, respectively which were largely similar to the results proposed by the software (75.33 and 84.71%). Hence, the pumpkin seed protein hydrolyzate with high antioxidant and nitric oxide inhibitory activity can be used as an alternative to synthetic preservatives in different food formulations including meat products. Considering the consumer's tendency toward functional foods and present concerns about application of synthetic additives, the results of this study lead to the production of a functional ingredient that can be used in various types of food formulation. This product is particularly interesting regarding its nitric oxide inhibitory property which eliminates concerns about the nitrate and nitrite residues and prevention of nitrosamines formation in meat products such as sausage. Also due to its protein structural nature its application increases the nutritional value and creates a product with health promoting advantages.

References


بهینه‌سازی شرایط هیدرولیز پروتئین دانه کدو با آنزیم آکالاز به منظور دستیابی به حداکثر فعالیت ضدکاسپیانی و مهار کندگی اکسید نیتریک

کوترازکری؛ علیرضا صادقی ماهونک، محمد قربانی، علی مودی، بیژی مقیودلو

- ۱- دانشجوی کارشناسی ارشد، گروه علوم و صنایع غذایی، دانشگاه علوم کشاورزی و منابع طبیعی گرگان، گرگان، ایران
- ۲- دانشیار، گروه علوم و صنایع غذایی، دانشگاه علوم کشاورزی و منابع طبیعی گرگان، گرگان، ایران
- ۳- استاد، گروه علوم و صنایع غذایی، دانشگاه علوم کشاورزی و منابع طبیعی گرگان، گرگان، ایران
- ۴- استادیاری، گروه علوم و صنایع غذایی، دانشگاه علوم کشاورزی و منابع طبیعی گرگان، گرگان، ایران

نویسندگان مسئول

* sadeghiaz@gau.ac.ir

چکیده

در این پژوهش، بهینه‌سازی شرایط هیدرولیز پروتئین دانه کدو (Cucurbita pepo) با آنزیم آکالاز به منظور دستیابی به حداکثر خصوصیات مهار کندگی اکسید نیتریک (DPPH) و مهار کندگی اکسید نیتریک با استفاده از نرم‌افزار Design Expert و روش سطح یافته فردی (Vary) فراوری گرفت. با این منظور، غلظت آنزیم آکالاز ۰/۷۰۰–۰/۳۰۰ درصد، دمای ۵۸–۳۲ درجه سانتی‌گراد، زمان ۲۹–۳۰ دقیقه به عنوان سطوح متغیرهای مستقل انتخاب شدند. نتایج نشان داد که شرایط بهینه برای دستیابی به حداکثر خصوصیات مهار کندگی اکسید نیتریک (DPPH) و مهار کندگی اکسید نیتریک، دمای ۴۴ درجه سانتی‌گراد، زمان ۲۹ دقیقه و غلظت آنزیم به سوی سوی ناپایداری ۲ درصد و با قابلیت ضدکاسپیانی و مهار کندگی اکسید نیتریک برابر با ۰/۳۰۷/۲۴۸۹ دقیقه و ۳/۷۱۷۷/۳۴ درصد بود که با حدود زیادی مشابه با نتایج پیشنهادی توسط Nafar (۰/۷۷/۳۷/۳۷ و ۰/۷۸/۱/۳۷/۵۷) درصد بود. طبق نتایج به دست‌آمده، پروتئین هیدرولیزه‌شدگی دانه کدو از قابلیت ضدکاسپیانی و مهار کندگی اکسید نیتریک مناسب برخوردار می‌باشد.

واژه‌های کلیدی: آکالاز، آنزیم ضدکاسپیانی، دانه کدو، مهار کندگی اکسید نیتریک، هیدرولیز آنزیمی
لینک های مفید

- عضویت در خبرنامه کارگاه های آموزشی
- سرویس ترجمه تخصصی STRS
- فیلم های آموزشی
- بلاگ مربوط به مراکز اطلاعات علمی
- سرویس های ویژه

40% تخفیف به مناسبت سالروز تاسیس مرکز اطلاعات علمی