Cloning and Expression of N-terminal Region of IpaD from Shigella dysenteriae in E. coli

Mojtaba Saadati1,2,*, Mohammad Heiat2, Shahram Nazarian1, Babak Barati1, Hossain Honari2, Mohammad Doroudian1,2, Mahdi Hesaraki1, Fatemeh Malaei1, Marzieh Eghtedardoust1, Mohammad Rahbar3, Saeed Hossein zadeh4, Mehrdad Hashemi5

1Department of Biology, Imam Hosssein University, Tehran, Iran.
2Applied Biotechnology and Environmental Research Center, Baqiyatallah Medical Science University, Tehran, Iran.
3Microbiology group collaborating Center for Reference and Research on Rabies.
4Department of Food Microbiology, School of Veterinary Medicine, Shiraz University, Shiraz, Iran
5Department of genetics, Islamic Azad University, Tehran Medical Branch

*Corresponding Author: email address: Saadati.m@yahoo.com (Mojtaba Saadati)

ABSTRACT

Genus Shigella is one of the important members of the family enterobacteriacae. There are numerous antigens in Shigella carrying by a 220 kb plasmid. Among them, IpaD is the key virulence factor of S. flexneri. Apart from having effector functions that is essential for host cell invasion and intracellular survival, this protein also controls the secretion and translocation of other effector proteins into eukaryotic host cells. In the present study, we have cloned and expressed the ipaD in E. coli. The ipaD gene was amplified by PCR. Prokaryote expression vector pET-28a(+) ipaD was constructed, and used to transform E. coli BL21DE3 pIYSs. The expression of recombinant protein induced by IPTG was examined by SDS-PAGE. Western blot were used to determine immunoreactivity of IpaD-His by a rabbit monoclonal antibodies against his-tag. SDS-PAGE demonstrated that the constructed prokaryotic expression efficiently produced IpaD at the 1 mmol/L of IPTG. IpaD protein was able to react with the rabbit monoclonal antibody against His-tag. IpaD is essential for Shigella spp invasion. N-terminal region is most significant functional fragment of IpaD. Purification of IpaD from the wild type of Shigella is difficult furthermore profound study on a specific domain on the N-terminal of IpaD by using the wild type of purified IpaD is not feasible.

Keywords: Shigella spp; N-terminal region, ipaD; cloning; expression

INTRODUCTION

Enterobacteriaceae, are large groups of the enteric, non-spore forming bacteria which considered as normal inhabitants of intestine in both animals and human. E. coli, salmonella, klebsiella, proteus and enterobacter are some important genus in this family. Some strains such as E. coli are considering as normal inhabitants while some are normal flora of gastro-intestinal tract (GIT). A few genuses in the family, enterobacteriaceae such as Salmonella and Shigella are pathogen for human and animals [1]. Shigella dysenteriae is one of the major causes of dysentery during last century, with a different mortality rate [1, 2]. The prevention of such diseases is mainly depending on the level of sanitation. No effective vaccination program has been proposed, so far. Few antigens of shigella have expressed which were potentially immunogenic in human. A large number of these antigens are carrying by a 220 kDa plasmid [3, 4]. A number of operons are located on the plasmid in which the entry region is playing a major role in the antigenicity of the microorganism [5]. ipaD is one of the operon harboring Ipa,a, b, c, and d [6]. Each operon is playing a role in the invasion of shigella to the M-cells with the clone [7]. IpaD is a 37 kDa with a dumbbell shape structure which exactly localizes to the tip of the type III secretion system needle of Shigella spp [8]. IpaD is necessary to invade the microorganism to host cells, carrying two main regions; N- and C-terminals in which the latter is a hydrophobic domain and not available in the environment. IpaD is fused to the C-terminal domain of a protein called MxiH, the main subunit of T3SS, thus, the protein residues of MxiH is cleaving a ligand for the C-terminal portion of IpaD [9, 8]. IpaD is interacting to the surrounding environment by its globular N-terminal. The key factor in the invasion of shigella to host epithelial cells is depending on the function of IpaD [7, 8]. Bile salts are also necessary to induce the
invasive potential of the bacterium [10]. In fact, the reaction between bile salts especially deoxycholate with IpaD may recruitment and employment of other proteins to locate on cell membrane of the bacterium [8, 11]. The condition permits the bacterium to attach and invade the host cells [8, 12]. Laboratory techniques and statistical analysis of previous data are indicating that deoxycholate can directly attach to a specific region on the N-terminal portion of IpaD, between MixH and IpaD. Any interaction in the function of this region may limit the function of IpaD and thus block the attachment process and/or invasion of the bacterium to the target cells [8, 11]. Previous researches have shown that the antibody that detects the N-terminal of IpaD, can stop the ability of the microorganism in making pores in red blood cells (RBC) [8]. However, further researches have shown that anti-IpaD antibody may inhibit the entry of shigella into its host cells. Recent work on producing an effective vaccine against shigella is based on using IpaD and its functional derivatives was also successful [13]. Therefore, IpaD could be considered as potential candidate and thus, some more work on the antigenicity of IpaD are seems to be Recruitment. Cloning and expression of the ipaD and its derivatives are necessary to find more about the antigenicity of such compounds. In the present study, we have tried to clone and express the N-terminal portion of the IpaD.

MATERIALS AND METHODS

Enzymes, Vectors and Bacterial strains

Pfu DNA polymerase (2.5U/µl, Fermentas, Lithuania), Enzymes HindIII and EcoRI (Fermentas, Lithuania), IPTG (Vivantis, Malesia), Vector pET-28a(+) (Novagen USA). Vector pGEM-T (Promega, USA), S. dysenteria, was prepared from Milad hospital and confirmed by biochemical and serological tests. The genomic DNA of shigella dysenteria was Extracted and used as the temptple in PCR experiment. E. coli DH5α and E. coli BL21DE3 plysS were used for cloning and expression experiments respectively. Plasmid pGEM-T Vector and pET-28a(+) were used as cloning and expression vectors respectively.

Amplification of N-terminal section of ipaD gene

Genomic DNA was extracted by a routine CTAB NaCl method [14]. The DNA fragment coding for N-terminal region of ipaD gene, with accession number NC_007607.1, was amplified by using 2 primers. These primers which were designed by oligo software were IpDF (5'T CAT GAA TTC AGA ACA ACA AAT CAG 3') as a forward primer with an endonuclease site of EcoRI and IpaDR (5'T CTT AAG CTT TTA AGT ATA TGA ACT AAC G 3') as reverse primer with an endonuclease site of HindIII. Synthesis of those primers was performed by Cinaclon Biotech Company. Amplification was made in a total volume of 50 µl of reaction mixture containing 1µl of genomic DNA (0.5µg/µl), 5 µl of 10× Pfu buffer with MgSo₄, 5 µl dNTP Mix(2mM each), 2 µl of each primer (10 pmol) and 0.25 µl of Pfu DNA polymerase (2.5 unit/µl). By adding double-distilled water, the reaction mixture reached the Final volume 50 µl. A total of 32 cycles was performed with the first denaturation at 95 °C for 5 min, then 30 cycles at 95 °C for 30 second, 57 °C for 30 second and 72 °C for 30 second, and the final extension at 72 °C for 5 min. The amplified products were analyzed by electrophoresis with a 1% agarose gel followed by ethidium bromide staining and UV-transilluminator visualization.

Cloning, subcloning and sequencing

The PCR products of N-terminus region of ipaD were trailed with dATP, ligated with pGEM-T vector and transformed into E. coli DH5α. Ampicillin selection and some control tests ensured the presence of the recombinant plasmid. In order to subcloning, Fragments of EcoRI and HindIII-digested ipaD were inserted into the EcoRI/HindIII site of expression vector pET-28a(+), through enzyme digestion and ligation reactions. Then the recombinant plasmid pET-28a(+)-ipaD was confirmed by PCR and restriction enzyme digestion. Recombinant pET-28a(+)-ipaD was amplified in competent E. coli DH5α and then extracted by Alkaline lysis method [14]. The resulting plasmid pET-28a(+)-ipaD was Transformed into competent final host E. coli BL21DE3 plysS and kanamycin resistance was used for selection [14]. A recombinant plasmid was prepared and identified by restriction enzymes. In order to sequence of inserted fragment, 20 µl of purified recombinant plasmid was sequenced by Koser Biotech Company.

Expression and identification of the fusion protein
The N-terminal ipaD expression system pET-28a(+)ipaD BL21DE3 was cultured in LB medium at 37 °C. After the OD of bacteria reached to 0.6, it induced by isopropylthio-β-D-galactoside (IPTG) at final concentration of 1 mmol/L. The precipitate and incubated for 5h. The bacteria were collected by centrifugation and the cell pellet was broken by B Buffer (NaH₂PO₄:13/8gr, Tris.HCl:1/2gr, urea 480/5gr, add DDW to 1liter, adjust pH in 8). The molecular weight of IpaD fusion protein was identified by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE).

Immunoblot analysis

Immunoreactivity of IpaD fusion protein was determined by Western blot. For Western blotting, proteins were transferred to nitrocellulose membrane and incubated with the rabbit antiserum against Histidin and HRP-labeling sheep anti-rabbit IgG as the first and second antibodies, respectively.

RESULTS

Construction of recombinant pET-28a(+)ipaD

The PCR product amplified from genomic DNA of shigella is shown in figure 1. The 344-bp expected fragment amplified by PCR contained a gene ipaD. The 344-bp PCR product was cloned into pGEM-T vector at first step. The recombinant pGEM-T-ipaD vector digested with EcoRI and HindIII enzymes and ipaD fragments ligated into the corresponding sites of pET-28a(+). The recombinant plasmids pET-28a(+)ipaD were digested by EcoRI and HindIII and analyzed on agarose gel electrophoresis is shown in figure 2.

Expression of recombinant fusion protein

The recombinant pET-28a(+)ipaD was transformed into E. coli BL21DE3 strains and the fusion protein was expressed. The 1 mmol/L of IPTG was able to efficiently induce expression of IpaD fusion protein with a predicted molecular mass of 16.7 kDa is shown in figure 3. A culture of recombinant E. coli BL21DE3 plysS that wasn’t induced by IPTG used as a negative control.

Western blot analysis using a specific

Figure 1. The target fragment of N-terminal of ipaD gene amplified from Shigella dysenteriae.
Lane 1: The target amplification of ipaD gene (S. dysenteriae)
Lane 2: 100 bp DNA size marker. (#SM0623 purchased from fermentas co.)

Figure 2. Agarose gel electrophoresis analysis of recombinant pET-28a(+)ipaD
Lane1: Double digests of recombinant pET-28a(+)ipaD with EcoRI and HindIII
Lane2: Recombinant pET-28a(+)ipaD without digestion
Lane3: 100bp DNA size marker (#SM0623 purchased from fermentas co.)
antibody against Histidine tag.
Expression of the N-terminal region of IpaD was investigated in E. coli. For this reason, an antibody against the Histidine tag was employed. The expression of the recombinant protein was then confirmed by appearing a proper band control is shown in figure 4.

DISCUSSION
The main agent of shigellosis is S. dysenteriae, which is a gram negative pathogenic bacterium characterized by its invasion to the epithelial cells of large intestine. Attachment of the microorganism to the host cells may induce an explosive secretion of proteins via the type three secretion system (TTSS) of the bacterium. Shigella is transmitted by the fecal-oral route, typically via contaminated water, and is spread efficiently due to an unusually low 50% infective dose [7]. After ingestion, acid-tolerant S. dysenteriae passes through the stomach to the colonic mucosa, by transcytosis through M cells and encounters resident macrophages [15]. There, the shigellae enter macrophages and efficiently induce apoptosis, leading to the eventual release of the pathogen on the basal side of the colonic epithelium [16]. The Shigella then invade these epithelial cells by inducing major cytoskeletal rearrangements [17]. The Shigella invasive genes have located on a 31-kb region of its large virulence plasmid which includes the genes for the type III secretion system (TTSS) [18]. The first group of proteins in which are excreted are: IpaD, IpaB and IpaC. The IpaD is essential component to invade the host cells. Antibodies are able to recognize the surface
epitopes of IpaD. In the present study, the antigenic properties of IpaD was investigated by calculative and laboratory techniques. The overall surface available epitopes of the IpaD are surrounded within the first 180 amino acids composition of the protein, whereas, the carboxyl region in the surface of Shigella is not available. Even though, hydrophilic areas are present in the carboxyl side, recent studies on the serum of animals have shown that the production of antibodies against the C-terminal is considerably lower than the N-terminal region [19]. Purification of IpaD from the wild type of Shigella has previously explained. However, the technique is difficult, firstly because the microorganism produces the IpaD protein in low level. The second difficulty in the purification of IpaD is referring to find it among the plenty of proteins which need expensive and sophisticated biochemical techniques [20]. However, a profound study on a specific domain on the N-terminal of IpaD by using the wild type of purified IpaD is not feasible; furthermore, the risks of working with the wild type of Shigella and also high costs experiments are making them to be impractical.

The microorganism produces the IpaD protein in low level and purification of IpaD among the plenty of proteins is very difficult, one of the suitable method for producing of high level of protein is utilizing of recombinant method. Using of this method also endow the ability of studying on a specific region of antigen like IpaD. Although purification of IpaD from wild type bacterium is difficult but it can proposed as a vaccine candidate for shigellosis.

REFERENCES

www.SID.ir