

SID



سرویس های ویژه



سرویس ترجمه تخصصی



کارگاه های آموزشی



بلاگ مرکز اطلاعات علمی



عضویت در خبرنامه



فیلم های آموزشی

کارگاه های آموزشی مرکز اطلاعات علمی جهاد دانشگاهی



PROPOSAL

پروپوزال

مركز آموزش پروپوزال نویسی و پایان نامه نویسی

کارگاه آنلاین پروپوزال نویسی و پایان نامه نویسی



مركز آموزش روش تحقیق و مقاله نویسی علوم انسانی

کارگاه آنلاین روش تحقیق و مقاله نویسی علوم انسانی



ISI Scopus

مركز آموزش آشنایی با پایگاه های اطلاعات علمی بین المللی و ترکیه های جستجو

کارگاه آنلاین آشنایی با پایگاه های اطلاعات علمی بین المللی و ترکیه های جستجو

Protective Effect of *Cornus mas* Fruits Extract on Serum Biomarkers in CCl₄-Induced Hepatotoxicity in Male Rats

Seyed Moayed Alavian¹; Nafiseh Banihabib²; Masoud Es. Haghi^{3,4,*}; Farid Panahi⁵

¹Middle East Liver Disease Center (MELD), Tehran, IR Iran

²Liver and Gastrointestinal Diseases Research Center, School of Medicine, Tabriz University of Medical Sciences, Tabriz, IR Iran

³Baqiyatallah Research Center for Gastroenterology and Liver Disease (BRCGL), Baqiyatallah University of Medical Sciences, Tehran, IR Iran

⁴Drug Applied Research Center, School of Medicine, Tabriz University of Medical Sciences, Tabriz, IR Iran

⁵Coordination Center, School of Medicine, Tabriz University of Medical Sciences, Tabriz, IR Iran

*Corresponding Author: Masoud Es. Haghi, Baqiyatallah Research Center for Gastroenterology and Liver Disease, Baqiyatallah University of Medical Sciences, Tehran, IR Iran. Tel: +98-9148909189, Fax: +98-4113319716, E-mail: Masood.s.haghi@gmail.com

Received: January 17, 2013; Revised: October 25, 2013; Accepted: February 11, 2014

Background: Nowadays attention to use herbs such as cornelian cherry (*Cornus mas*) is increasing, which contains high levels of antioxidants and anthocyanins. *Cornus mas* fruits have been used for gastrointestinal and excretory disorders for many years in traditional medicine, also may improve liver and kidney functions, and have protective effects such as anti-allergic, antidiabetic, antibacterial, antimicrobial, antihistamine and antimalarial properties.

Objectives: The aim of this study was to investigate protective effects of *Cornus mas* fruits extract on serum biomarkers in CCl₄-induced hepatotoxicity in male rats.

Materials and Methods: Hepatotoxicity was induced by administration of carbon tetrachloride (1 mL/kg i.p.) in 1:1 dilution with olive oil. To evaluate the effect of *Cornus mas* fruits extract on disease progression, serum marker enzymes, serum total protein and albumin and liver lipid peroxidation were determined in CCl₄-induced hepatotoxicity.

Results: Oral administration of *Cornus mas* fruits extract to rats for 14 days provided a significant ($P < 0.05$) hepatoprotection by decreasing elevated serum level of enzymes, total serum protein, albumin and liver lipid peroxidation content.

Conclusions: *Cornus mas* fruit extract effect may be due to including some antioxidant components, which caused membrane stabilizing and normalization of fluctuated biochemical profiles induced by CCl₄ exposure. Our results validated the traditional use of *Cornus mas* in the treatment of liver disorders.

Keywords: Carbon Tetrachloride; *Cornus mas*; Hepatotoxicity; Lipid Peroxidation; Serum Biomarkers

1. Background

Hepatic injuries lead to attenuation of metabolic functions regulated by liver, and has remained one of the serious health problems (1) threatening the human society. The pathogenesis of liver fibrosis is not clear, but reactive oxygen species (ROS) have important roles in liver pathological changes (2). Unsaturated fatty acids of biological cell membranes (a sensitive part of cells against free radicals) may be affected by peroxidation reaction, which leads to a decrease in the fluidity and disruption of membrane function and integrity, and finally serious pathological changes (3). Oxygen free radicals and oxygen free-radical generating agents such as hydrogen peroxide (H₂O₂), superoxide anion (O₂^{•-}) and hydroxyl radical ([•]OH) are known as ROS. During the metabolic processes, free radicals are usually generated especially oxygen-derived radicals with potency of oxidizing and damaging biomolecules (4). Biotransformed of trichloromethyl-free radical (CCl₃[•] or CCl₃OO[•]) from carbon

tetrachloride (CCl₄) by hepatic microsomal cytochrome P450, is a well-known model compound for producing chemical hepatic injury (5-9). Overproduction of trichloromethyl-free radicals is considered the initial step in a chain of events leading to membrane lipid peroxidation and eventually cell apoptosis or necrosis (10-13). ROS production and their leading damages would be confined by several endogenous protective mechanisms (3). When ROS formation is extreme, additional protective mechanisms of dietary antioxidants may be of great importance (14, 15).

Antioxidant nutrients and enzymes are the cooperative protective defensive systems against free radical damages (16). Antioxidant and radical scavengers have been studied on the mechanism of CCl₄ toxicity and protect liver cells from CCl₄ induced damage by lipid peroxidation (12). Catalase (CAT), superoxide dismutase (SOD), glutathione reductase (GR) and glutathione peroxidase

Implication for health policy/practice/research/medical education:

Decrease hospitalization, complication and health cost of patients of liver diseases/decrease hepatotoxicity in patients with systemic disease/ Effect of different nutrition materials on hepatotoxicity/ pharmacokinetic effect of nutrition materials on hepatic cells.

Copyright © 2014, Kowsar Corp.; Published by Kowsar Corp. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

(GPx) as antioxidative enzymes and glutathione (GSH), vitamins C and E as nonenzymatic antioxidants are biological antioxidants (17). Natural antioxidants such as fruits and vegetables, which provide protection against free radicals can decrease the incidence and mortality rates due to cancer and heart diseases, in addition to a number of other health benefits (18). Recently, an increasing trend of herbs usage has been seen, such as cornelian cherry (*Cornus mas*), which contains high level of antioxidants and anthocyanins. *Cornus mas* fruits have been used for gastrointestinal and excretory disorders in traditional medicine (19); also improve liver and kidney functions, and have protective effects such as antiallergic, antidiabetic, antibacterial, antimicrobial, antihistamine and antimalarial properties (20, 21).

Cornus mas (*C. mas*) is known as the European and Asiatic Cornelian Cherry. This plant has been used in traditional medicine for the treatment of diarrhea, intestinal inflammation, fever, malaria treatment, kidney stones and kidney and bladder infections. *C. mas* fruits contain iron, calcium, vitamins such as E, B2, B1, C, folic acid, anthocyanins, flavonoids, and plenty of oxalic acid (22, 23). It also contains antioxidant substances including butylated hydroxyanisole and butylated hydroxytoluene and has the potential in the treatment of some cancers (24, 25).

2. Objectives

This study was designed to investigate the protective effects of *Cornus mas* fruits extract on serum biomarkers in CCl₄-induced hepatotoxicity in male rats.

3. Materials and Methods

3.1. Chemicals

Ethylene diamine tetra acetic acid (EDTA) and trichloroacetic acid (TCA) were obtained from Sigma-Aldrich Chemical Co. Ltd. (USA). Carbon tetrachloride (CCl₄) and thiobarbituric acid (TBA) were obtained from Merck Co. (Germany). Assay kits for the estimation of biochemical factor such as aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP) were purchased from Pars Azma (Iran).

3.2. Plant Material

Cornus mas fruits were obtained from suburbs of kaleibar (East Azarbayjan, Iran) at the end of summer 2012. The fruits were air-dried and powdered. In all steps, components were protected from direct sunlight. The powder was kept at 8 °C.

3.3. Extraction

Air-dried fruits of *Cornus mas* were powdered coarsely. Then 500 g of the powder was mixed with methanol: wa-

ter (7:3) at 25 ± 2 °C. The solvent was completely removed by rotary vacuum evaporator at 50 °C. Then *Cornus mas* fruits extract (CMFE) was frozen at -20 °C until use.

3.4. Animals and Treatment

Thirty animals (Wistar strain male albino rats [250-300 g]) were kept in polypropylene cages in a room at 22 ± 2 °C with humidity of 44-55%, the light and dark cycles (12/12 h for each) for one week before and during the experiments. Animals were fed with a standard rodent pellet diet and clean drinking water ad libitum. The protocol of this study was approved by the Animal Ethic Committee of Baqiyatallah University of Medical Sciences.

Animals were divided into five groups (n = 6) as follows:

Group I: normal control, received drinking water orally for 14 days, on the 14th day olive oil (1 mL/kg i.p.).

Group II: toxic control, received drinking water for 14 days orally, on the 14th day CCl₄ (1 mL/kg i.p.) in 1:1 dilution with olive oil.

Groups III and IV: treatment groups, received CMFE at the doses of 200 and 500 mg/kg orally for 14 days, respectively, on the 14th day received CCl₄ (1 mL/kg i.p.) at the same dose, two hours after the last dose of extract administration. Group V: standard drug Silymarin, received silymarin (100 mg/kg, orally) for 14 days and on the 14th day received CCl₄ (1 mL/kg i.p.) in 1:1 dilution with olive oil, two hours after administration of the last dose of silymarin.

3.5. Preparation of Serum

Animals were killed 24 hours after administration of CCl₄. Blood samples were collected from the heart left ventricular and allowed to clot for 30 minutes. Samples were centrifuged at 2500 rpm for 15 minutes and used for biochemical analysis. The collected serum samples were frozen at -20 °C. Aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) enzymes activity, total protein (TP) and albumin (Alb) in all samples were measured by standard diagnostic test kits (Pars Azma, Iran).

3.6. Provision of Liver Homogenate

Hepatic tissues were homogenized in KCl [10 mM] phosphate buffer (1.15%) with ethylenediamine tetra acetic acid (EDTA: pH = 7.4) and centrifuged at 3000 rpm for 30 min. The supernatant was collected to use for the measurement of malondialdehyde (MDA). Total protein content was determined based on the Lowry's method (26).

3.7. Designation of Lipid Peroxidation

Liver homogenate lipid peroxidation was measured based on the formation of thiobarbituric acid reactive substance (TBARS). Two milliliters of thiobarbituric acid reagent (15% w/v TCA, 0.375% w/v TBA and 0.25 M HCl) was added to 2 mL of supernatant. The dilution was heated

for 15 minutes in boiling water. After cooling, centrifuged at 1000g for ten minutes and precipitate was removed. Malondialdehyde (MDA) forms mixed with TBA, which was measured by spectrophotometer at 532 nm. The concentration of MDA was computed based on the absorbance coefficient of the TBA-MDA complex ($\epsilon = 1.56 \times 10^5/\text{M}/\text{cm}$), and it was presented as nmol/mg protein (27).

3.8. Statistical Analysis

Data were presented as mean \pm S.E. Tukey post hoc test was used to compare different parameters between the groups. The significant level was considered at P value < 0.05.

4. Results

4.1. CMFE Effect on Serum Enzymes Activity

As shown in Table 1, activities of serum ALT, AST and ALP were markedly elevated in toxic group compared to control group, indicating liver damage. Treatment with *Cornus mas* fruits extract (CMFE) in the two different dosages at 200 and 500 mg/kg for 14 days remarkably prevented CCl_4 induced elevation of serum enzymes activity.

4.2. Effect of CMFE on Serum Level of Total Protein and Albumin

As given in Table 1, the TP and Alb concentrations in toxic group were lower compared with control group and it attained near the normal value in groups treated with CMFE.

4.3. Effect of CMFE on Lipid Peroxidation

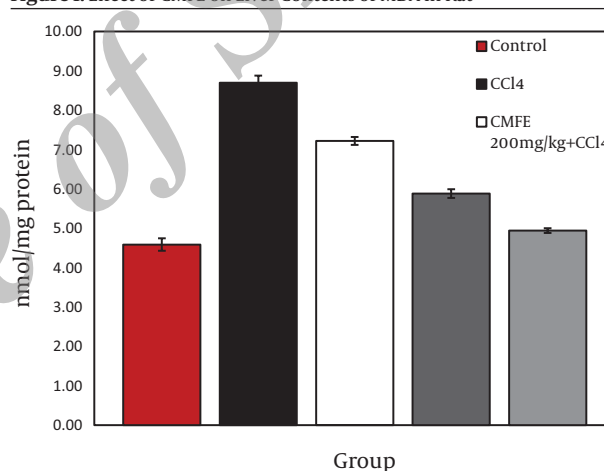
Evaluated level of Malondialdehyde content in homogenate of rat liver is shown in Figure 1. MDA content in liver homogenate was significantly ($P < 0.05$) increased in CCl_4 group compared to normal control. MDA level of CMFE treatment at 200 and 500 mg/kg were significantly ($P < 0.05$) decreased compared to toxic (Group 2).

5. Discussion

Our study showed the protective effects of *Cornus mas* fruits extract (CMFE) on serum biomarkers against hepatotoxicity induced by CCl_4 in rats. Increased generation of reactive oxygen species (ROS) has a major effect on the pathogenesis and toxicity of a wide range of compounds (28). Nowadays hepatoprotective drugs are generally used against liver damages induced by CCl_4 (is a hepatotoxic agent). Histological signs and features of liver damage by CCl_4 are similar to acute viral hepatitis. CCl_4 has been commonly used in rat experimental models to investigate the oxidative stress induced in various organs. To the best of our knowledge, this is the first study to evaluate these effects of CMFE in an attempt to prevent liver damage from CCl_4 .

In liver damages, the serum level of enzymes located normally in cytosol increased due to releasing into the blood. The type and range of hepatocellular damage can be estimated by the serum level of these enzymes (29).

Figure 1. Effect of CMFE on Liver Contents of MDA in Rat



* Indicates significance at $P < 0.05$ probability from control group; # Indicates significance at $P < 0.05$ probability from CCl_4 group.

Table 1. Effect of CMFE on Serum Profile in Rat ($n = 6$)^{a, b}

Treatment	AST, U/L	ALT, U/L	ALP, U/L	T-Protein, g/dL	Albumin, g/dL
Control + olive oil	58.12 \pm 5.52	43.86 \pm 6.35	214.29 \pm 4.83	7.85 \pm 0.15	3.65 \pm 0.06
1 mL/kg CCl_4	155.74 \pm 4.21 ^c	197.4 \pm 5.05 ^c	322.63 \pm 10.98 ^c	5.83 \pm 0.23 ^c	1.31 \pm 0.08 ^c
200 mg/kg CMFE + CCl_4	101.82 \pm 4.69 ^d	133.60 \pm 5.59 ^d	248.97 \pm 6.11 ^d	7.10 \pm 0.11 ^d	2.20 \pm 0.09 ^d
500 mg/kg CMFE + CCl_4	115.03 \pm 2.96 ^d	116.87 \pm 5.65 ^d	235.23 \pm 5.24 ^d	7.25 \pm 0.14 ^d	2.85 \pm 0.12 ^d
100 mg/kg Silymarin + CCl_4	85.22 \pm 3.52 ^d	72.92 \pm 3.25 ^d	223.64 \pm 5.23 ^d	7.91 \pm 0.16 ^d	3.20 \pm 0.09 ^d

^a Abbreviations: ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; CCl₄, carbon tetrachloride; CMFE, *Cornus mas* fruits extract.

^b Data are presented in Mean ± SE.

^c Indicates significance at P < 0.05 probability from control group.

^d Indicates significance at P < 0.05 probability from CCl₄ group.

AST, ALT and ALP are authentic markers for liver function. Increased serum level of enzymes such as aspartate aminotransferase and alanine amino transferase have been seen in rats received CCl₄ leading to increased cell damage, permeability, and hepatocytes necrosis. Alkaline phosphatase as a membrane bound enzyme, is excreted in association with bile when liver is affected, defective excretion increased the serum level of this enzyme (30). Treatment by oral administration of CMFE at doses of 200 and 500 mg/kg in rats decreased the mentioned enzymes activity. This event may be followed by plasma membrane stabilization and improve in CCl₄ induced hepatic tissue damage, thus the serum levels of transaminases become normal with hepatic parenchyma healing and regeneration of hepatocytes (31). Total protein is a common laboratory test to evaluate the effect of various toxic chemicals (32). Decline in TP content can be deemed as a useful index of severity of hepatocellular damage. The lowered levels of TP and Alb recorded in the serum as well as in the liver of CCl₄-treated rats revealed the severity of hepatopathy (33). In the present study, TP and Alb concentration were very low in rats treated with CCl₄. In groups treated with CMFE, these markers significantly (P < 0.05) increased compared to the toxic group and the values were closer to those of the control group.

Oxidative stress initiates lipid peroxidation of cell membranes polyunsaturated fatty acids (34). Lipid peroxidation represents one of the most frequent reactions resulting from free radicals invasion on biological structures and by cumulating oxidized lipids in the cell membrane (35). Our result showed the reduction effect of CMFE on TBARS production. In this study, CCl₄ administration significantly (P < 0.05) increased the hepatic MDA content, which may indicate an increase in lipid peroxidation. Significant decrease in the hepatic malondialdehyde content, as a marker of lipid peroxidation, confirmed that treatment with CMFE could have a great protective effect against the CCl₄-induced hepatic lipid peroxidation. This protection mechanism preserves the liver from toxin induced damages through the hepatic regeneration stimulating and liver lipid peroxidation inhibiting (36).

Cornus mas fruit extract effect may be due to some antioxidant components, leading to membrane stabilizing and normalization of fluctuated biochemical profiles induced by CCl₄ exposure. Therefore, plant extract compounds effect on the liver is to keep its normal function and decrease derangements of cell membrane. Purification of the active component(s) of *Cornus mas* to determine the exact protective effects on hepatocytes is recommended for further studies.

Acknowledgements

Authors would like to thank all staff of Baqiyatallah Research Center for Gastroenterology and Liver Disease for their helps and supports.

Authors' Contribution

Seyed Moayed, Alavian: study design and data interpretation; Banihabib, Nafiseh: data collection, statistical analysis and data interpretation; Es. Haghi, Masoud: data collection, literature review, manuscript preparation and statistical analysis; Panahi, Farid: manuscript preparation and literatures review.

Financial Disclosure

The authors declared no potential conflicts of interest.

Funding/Support

Baqiyatallah Research Center for Gastroenterology and Liver Disease, Baqiyatallah University of Medical Sciences, Tehran, Iran

References

1. Wolf PL. Biochemical diagnosis of liver disease. *Indian J Clin Biochem.* 1999;**14**(1):59-90.
2. Poli G, Parola M. Oxidative damage and fibrogenesis. *Free Radic Biol Med.* 1997;**22**(1-2):287-305.
3. Sies H. Strategies of antioxidant defense. *Eur J Biochem.* 1993;**215**(2):213-9.
4. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol.* 2007;**39**(1):44-84.
5. Brattin WJ, Glende EA, Jr, Recknagel RO. Pathological mechanisms in carbon tetrachloride hepatotoxicity. *J Free Radic Biol Med.* 1985;**1**(1):27-38.
6. Recknagel RO, Glende EA, Jr. Carbon tetrachloride hepatotoxicity: an example of lethal cleavage. *CRC Crit Rev Toxicol.* 1973;**2**(3):263-97.
7. Rikans LE, Hornbrook KR, Cai Y. Carbon tetrachloride hepatotoxicity as a function of age in female Fischer 344 rats. *Mech Ageing Dev.* 1994;**76**(2-3):89-99.
8. Shenoy KA, Somayaji SN, Bairy KL. Hepatoprotective effects of Ginkgo biloba against carbon tetrachloride induced hepatic injury in rats. *Indian J Pharmacol.* 2001;**33**(4):260-6.
9. Eshaghi M, Zare S, Banihabib N, Nejati V, Farokhi F, Mikaili P. Cardioprotective Effect of Cornus Mas Fruit Extract against Carbon Tetrachloride Induced-Cardiotoxicity in Albino Rats. *Journal of Basic and Applied Scientific Research.* 2012;**2**(11):1106-14.
10. Basu S. Carbon tetrachloride-induced lipid peroxidation: eicosanoid formation and their regulation by antioxidant nutrients. *Toxicology.* 2003;**189**(1-2):113-27.
11. Brautbar N, Williams J, 2nd. Industrial solvents and liver toxicity: risk assessment, risk factors and mechanisms. *Int J Hyg Environ Health.* 2002;**205**(6):479-91.
12. Weber LW, Boll M, Stampfl A. Hepatotoxicity and mechanism of

- action of haloalkanes: carbon tetrachloride as a toxicological model. *Crit Rev Toxicol*. 2003;**33**(2):105–36.
13. Williams AT, Burk RF. Carbon tetrachloride hepatotoxicity: an example of free radical-mediated injury. *Semin Liver Dis*. 1990;**10**(4):279–84.
 14. Lieber CS. Role of oxidative stress and antioxidant therapy in alcoholic and nonalcoholic liver diseases. *Adv Pharmacol*. 1997;**38**:601–28.
 15. Cervinkova Z, Drahotova Z. Enteral administration of lipid emulsions protects liver cytochrome c oxidase from hepatotoxic action of thioacetamide. *Physiol Res*. 1998;**47**(2):151–4.
 16. Sreelatha S, Padma PR, Umadevi M. Protective effects of Coriandrum sativum extracts on carbon tetrachloride-induced hepatotoxicity in rats. *Food Chem Toxicol*. 2009;**47**(4):702–8.
 17. Fridovich I. Fundamental aspects of reactive oxygen species, or what's the matter with oxygen? *Ann NY Acad Sci*. 1999;**893**:13–8.
 18. Shui G, Leong IP. Residue from star fruit as valuable source for functional food ingredients and antioxidant nutraceuticals. *Food Chem*. 2006;**97**(2):277–84.
 19. Celik S, Bakırcı I, Sat IG. Physicochemical and organoleptic properties of yogurt with cornelian cherry paste. *Int J Food Prop*. 2006;**9**(3):401–8.
 20. Tural S, Koca I. Physico-chemical and antioxidant properties of cornelian cherry fruits (*Cornus mas* L.) grown in Turkey. *Sci Hort*. 2008;**116**(4):362–6.
 21. Vared SK, Reddy MK, Schutzki RE, Nair MG. Anthocyanins in *Cornus alternifolia*, *Cornus controversa*, *Cornus kousa* and *Cornus florida* fruits with health benefits. *Life Sci*. 2006;**78**(7):777–84.
 22. Tengvall Linder M, Johansson C, Zargari A, Bengtsson A, van der Ploeg I, Jones I, et al. Detection of *Pityrosporum orbiculare* reactive T cells from skin and blood in atopic dermatitis and characterization of their cytokine profiles. *Clin Exp Allergy*. 1996;**26**(11):1286–97.
 23. Serteser A, Kargıoğlu M, Gök V, Bağcı Y, Özcan MM, Arslan D. Antioxidant properties of some plants growing wild in Turkey. *Grasas Y Aceites*. 2009;**60**(2):147–57.
 24. Seeram NP, Schutzki R, Chandra A, Nair MG. Characterization, quantification, and bioactivities of anthocyanins in *Cornus* species. *J Agric Food Chem*. 2002;**50**(9):2519–23.
 25. Kocyigit M, Ozhatay N. Wild plants used as medicinal purpose in Yalova (Northwest Turkey). *Turkish J Pharm Sci*. 2006;**3**(2):91–103.
 26. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951;**193**(1):265–75.
 27. Buege JA, Aust SD. Microsomal lipid peroxidation. *Methods Enzymol*. 1978;**52**:302–10.
 28. Halliwell B, Gutteridge JMC. Oxygen radicals and the nervous system. *Trends Neurosci*. 1985;**8**:22–6.
 29. Jadon A, Bhadauria M, Shukla S. Protective effect of Terminalia bellerica Roxb. and gallic acid against carbon tetrachloride induced damage in albino rats. *J Ethnopharmacol*. 2007;**109**(2):214–8.
 30. Nemesanszky E. Enzyme test in hepatobiliary disease. In: Moss DW, Rosalki SB, editors. *Enzyme Tests in Diagnosis*. London: Arnold.; 1996. pp. 25–59.
 31. Thabrew MI, Joice PD, Rajatissa W. A comparative study of the efficacy of *Pavetta indica* and *Osbeckia octandra* in the treatment of liver dysfunction. *Planta Med*. 1987;**53**(3):239–41.
 32. Nagata H, Takekoshi S, Takagi T, Honma T, Watanabe K. Antioxidative action of flavonoids, quercetin and catechin, mediated by the activation of glutathione peroxidase. *Tokai J Exp Clin Med*. 1999;**24**(1):1–11.
 33. Taniyama Y, Griendling KK. Reactive oxygen species in the vasculature: molecular and cellular mechanisms. *Hypertension*. 2003;**42**(6):1075–81.
 34. Janero DR. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. *Free Radic Biol Med*. 1990;**9**(6):515–40.
 35. Marathe GK, Harrison KA, Murphy RC, Prescott SM, Zimmerman GA, McIntyre TM. Bioactive phospholipid oxidation products. *Free Radic Biol Med*. 2000;**28**(12):1762–70.
 36. Sadasivan S, Latha PG, Sasikumar JM, Rajashekar S, Shyamal S, Shine VJ. Hepatoprotective studies on *Hedyotis corymbosa* (L.) Lam. *J Ethnopharmacol*. 2006;**106**(2):245–9.

SID



سرویس های ویژه



سرویس ترجمه تخصصی



کارگاه های آموزشی



بلاگ مرکز اطلاعات علمی



عضویت در خبرنامه



فیلم های آموزشی

کارگاه های آموزشی مرکز اطلاعات علمی جهاد دانشگاهی



PROPOSAL
پروپوزال

پروپوزال نویسی و پایان نامه نویسی

دوره آموزشی

کارگاه آنلاین
پروپوزال نویسی و پایان نامه نویسی



روش تحقیق و مقاله نویسی علوم انسانی

دوره آموزشی

کارگاه آنلاین
روش تحقیق و مقاله نویسی علوم انسانی



ISI
Scopus



آشنایی با پایگاه های اطلاعات علمی بین المللی و ترند های جستجو

دوره آموزشی

کارگاه آنلاین آشنایی با پایگاه های اطلاعات علمی بین المللی و ترند های جستجو