Evaluation of Anti-Platelet Aggregation Effect of Some *Allium* Species

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Abstract

Epidemiologic studies show that the cardiovascular diseases are associated with multiple factors such as raised serum total cholesterol, increased LDL, increased platelet aggregation, hypertension and smoking. *In-vitro* studies have confirmed the ability of some plants of *Allium* species to reduce these parameters. Therefore, we evaluated anti-platelet aggregation effect of some *Allium* species (*Allium ampeloprasum*, *A. hirtifolium*, *A. haemanthoides*, *A. vavillovi*, *A. atrovioleaceum*, *A. jesdianum*, *A. shelkovnikovii*) using arachidonic acid (AA) and adenosine diphosphate (ADP) as platelet aggregation inducers. The screening results for methanolic extract of *Allium* species showed that the maximum effect of anti-platelet aggregation was related to *A. atrovioleaceum*. This extract inhibited the *in-vitro* platelet aggregation induced by AA and ADP with IC\textsubscript{50} values of 0.4881 (0.4826-0.4937) mg/ml and 0.4945 (0.4137-0.5911) mg/ml respectively. These results support the hypothesis that the dietary intake of *Allium* could be beneficial for prevention of cardiovascular diseases.

Keywords: *Allium* species; Alliaceae; Cardiovascular disease; Anti-platelet; Platelet aggregation.

Introduction

Cardiovascular diseases are the most common reason of morbidity and mortality worldwide (1). In the western countries they account for almost 17 million deaths; in developing countries their occurrence is becoming important along with industrialization, urbanization and changing life styles (2). Cardiovascular disease is associated with multiple factors including the increased ability of platelets to aggregate (3). Increased platelet aggregation has an important role in the etiology of cardiovascular diseases, which might be the result of multiple different mechanisms. In addition, in the atherosclerotic states, platelets adhere to the damaged vessel surface and aggregate themselves in response to agonists such as adenosine-5-diphosphate (ADP), thrombin, collagen and thromboxane A\textsubscript{2} (4).

Interfering with the process of platelet aggregation is one of the therapeutic strategies for the treatment of platelet-related cardiovascular diseases.

In recent years, increasing attention has been drawn to health benefits of food, beverages, spices and seasonings beyond their nutritional, tasting and flavoring significance. Especially, plant-originating foodstuff such as green tea, soy bean, grape and their products have been recognized to exhibit the preventive and/or therapeutic effects on various diseases (5). Plant materials have long been used as traditional medicines for the treatment of a number of ailments and diseases.

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Scientific search for new therapeautic agents is strongly supported by epidemiological studies on medicinal plants. One would assert that it is feasible to make detailed studies so as to identify active ingredients of plant aiming to develop new drugs in the case the beneficial aspects are realized (6).

Since ancient time, a deep knowledge has supported the use of *Allium* species as food and/or medicine. *In vitro* studies have confirmed the ability of some plants of *Allium* species to reduce the parameters which are taken as risk factors in cardiovascular diseases such as raised serum total cholesterol, increased LDL, increased platelet aggregation, hypertension and smoking (7). *Allium* is the largest and most important representative genus of alliaceae family. Alliaceae is a family of flowering plants, includes about 30 genera and more than 670 species, and is widely distributed in the northern hemisphere (8).

It is believed that *Allium* is a very variable and taxonomically difficult genus. The most recent taxonomy proposal for this genus, based on morphological characters and as also molecular data, accepted about 780 species belonging to fifteen subgenera and 56 sections. Recently, more taxa were newly described raising this number to more than 800 species. Currently, about 115 species (including more than 40 endemic ones) are known in the territory of Iran. These numbers indicate that Iran belongs to main center of *Allium* diversity in southwest and central Asia (9).

*A. atrovioleacum* has been used as foods, spices, and herbal remedies (10). The initial report of scientific research on *Allium* began in second half of the 19th century, coined by the work of Louis Pasteur who highlighted the antibacterial properties of garlic in 1858. Garlic has also been used for several epidemic diseases. In recent years, much attention has focused on various aspects of the pharmacological effects of extracts and isolated compounds from these plants. Several biological activities have been reported for *Allium* species such as antibacterial, antiviral, antiparasitic, antifungal, anti-protozoa and anti-diabetic, effects on the circulatory and cardiovascular system, antioxidant, anticarcinogenic effects and treatment of common cold (8).

Many of the biological effects of *Allium* species are related to thiosulfinate, volatile sulfur compounds, typical of these plants, which are also responsible for their pungent aroma and taste properties. However, these compounds are unstable and give rise to transformed products. For this reason, recent attention has been focused on polar compounds which are more stable for cooking and storage. Among these compounds, sapogenins, saponins, and flavonoids are the main classes found (8, 11).

The present research was aimed at investigating the effect of total extracts of selected group of *Allium* species on platelet aggregation induced by AA and ADP. No previous anti-platelet aggregation study has been conducted on the plants which are selected for the present research.

**Experimental**

**Chemicals and Instruments**

All the chemicals were purchased from Sigma-Aldrich Chemie GmbH (Germany) or Merck (Germany) companies. The chemicals were of analytical grade. ADP and AA were purchased from Bio/Data, Corp (Germany). Anti-platelet aggregation activity of *Allium* species extracts was determined, using APACT-4004 aggregometer (LABiTec, Ahrensburg, Germany).

**Plant material**

The herbs were selected based on two factors: Iranian traditional source of medicine and scientific papers. Having surveyed the flora of Iran and the diversity of *Allium* species, different regions were chosen as target. After collecting specimens, voucher specimens were confirmed and deposited at the herbarium (Table 1).

**Extraction**

Aerial parts of herbs were carefully washed under tap water and left to dry in controlled temperature (22°C) without exposure to the light and moisture. They were chopped and then passed through sieve size 60 (25/0mm). The bulbous onions of plants, after collecting and cleaning up, were kept in nitrogen tanks in order
to preserve the volatile compounds. They were then transferred to Freeze Dryer and dried. In each case, 500g of powdered plant was extracted with 1500ml methanol by maceration method. Extraction was conducted for 48h and repeated four times under stirring at 80r/ minute rate. The extracts were then filtered and concentrated under vacuum at 40°C using a rotary evaporator.

Preparation of samples for Platelet aggregation test

The extracts were dissolved in DMSO at the concentration of 250mg/ml. In fact, the initial stock solution contained the maximum amount of the extract which was soluble in 1 ml. The entire stock was passed through a Syringe Filter PVDF 0.22µm which is compatible with the organic solvent DMSO. One microliter of the solution of extract in DMSO was added to 200µl of plasma so that, the final concentrations of extracts in plasma were from 1.25 to 0.3125mg/ml. Inspecting the filtrate for particles showed that despite the filtration, some particulate matters formed in the solution a few minutes after filtration. Therefore, in order to overcome the problem, polyvinyl alcohol (PVA) was added to the solutions at the concentration of 1% prior to the filtration process. No particulate matter formed in the solution even after one hour of the addition of PVA. A blank mixture of DMSO and PVA in plasma showed no effect on platelet aggregation process (Figure 1). Therefore a series of solutions of the plant extract was prepared in DMSO/PVA 1%.

Blood collection

Blood was obtained from healthy volunteers who did not take any medication for 14 days

Table 1. Scientific names, part of herb, place of collection, voucher number and place of deposition of plants which have been used in the present study.

<table>
<thead>
<tr>
<th>Scientific Name</th>
<th>Part of herb</th>
<th>Place of collection</th>
<th>Voucher number</th>
<th>Place of specimen deposition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Allium ampeloprasum L.</td>
<td>Aerial part</td>
<td>Mount Rig, lordegan- Shahr-e-kord</td>
<td>8012</td>
<td>1</td>
</tr>
<tr>
<td>Allium hirtifolium Boiss.</td>
<td>Bulb</td>
<td>Mount Sabz-e-koh, Shahr-e-kord</td>
<td>8010</td>
<td>1</td>
</tr>
<tr>
<td>Allium hirtifolium Boiss.</td>
<td>Aerial part</td>
<td>Mount Sabz-e-koh, Shahr-e-kord</td>
<td>8010</td>
<td>1</td>
</tr>
<tr>
<td>Allium haemanthoides Boiss. &amp; Reut.</td>
<td>Aerial part</td>
<td>Izeh, Alvaz</td>
<td>8008</td>
<td>1</td>
</tr>
<tr>
<td>Allium vavillovi M.Pop &amp; Vved.</td>
<td>Aerial part</td>
<td>Shahr-e-reza, Isfehan</td>
<td>8009</td>
<td>1</td>
</tr>
<tr>
<td>Allium atrovioacum Boiss</td>
<td>Aerial part</td>
<td>Mount Rig, lordegan- Shahr-e-kord</td>
<td>8013</td>
<td>1</td>
</tr>
<tr>
<td>Allium jesdianum Boiss. &amp; Buhse</td>
<td>Bulb</td>
<td>Tezerjan-Yazd</td>
<td>2252</td>
<td>2</td>
</tr>
<tr>
<td>Allium shelkovnikovii Grossh.</td>
<td>Bulb</td>
<td>Tabriz</td>
<td>3033</td>
<td>3</td>
</tr>
</tbody>
</table>

1Herbarium of department of Pharmacognosy of faculty of Pharmacy of Shahid Beheshti University of medical sciences
2Herbarium of Yazd Research Center for Agriculture and Natural Resources
3Herbarium of East Azarbaijan Research Center for Agriculture and Natural Resources
and were fasting overnight prior to the study. Blood was collected in falcon tube containing 0.1 volume of 2.2% sodium citrate. Platelet rich plasma (PRP) was prepared by the centrifugation of citrated blood at 100g for 10 min. The residual blood was centrifuged at speed of 1500g for 15 min to give platelet poor plasma (PPP). Platelets were counted under microscope and the platelet count was adjusted to (250 ± 25) ×10⁹/L. Supernatant PRP was diluted with PPP.

**Platelet aggregation studies**

Platelet aggregation responses were monitored with a turbidometric method using an optical aggregometer. Platelet aggregation was expressed as an increase in light transmission. The levels of transmission were calibrated by PPP and PRP. Aliquots of 200μl of PRP were distributed in the test cuvettes and placed in incubation chamber of aggregometer at 37°C. Platelet aggregation was measured using PRP after activation by the addition of ADP or AA according to Born method (12). 1μl of the prepared solution of methanolic extracts in DMSO/PVA was added to the PRP, 5 min prior to the activation with AA (1.35mM) and ADP (5µM). The extent of aggregation was quantified by determining the maximum height of the curve. The platelet aggregation inhibitory activity was expressed as percent inhibition compared with that measured for the vehicle (DMSO/PVA1%) alone (13). Finally, percent inhibition for extract is calculated according to the following formula:

\[
100 \times \left(1 - \frac{D}{S}\right) = \text{percent inhibition}
\]

where:

- \(D\) = Aggregation in the sample
- \(S\) = Aggregation in the presence of solvent (DMSO/PVA1%)

**Statistical analysis**

The anti-aggregation value of each extract was expressed as either % inhibition or IC₅₀ values (the concentration of the compound causing 50% inhibitory effects). The IC₅₀ values were estimated by non linear curve-fitting and presented as their respective 95% confidence limits. One-way analysis of variance (ANOVA) followed by Tukey’s post test was used to assess the significant differences (p < 0.05) between the responses caused by the extracts. All the statistical analyses were accomplished using the computer software GraphPad Prism 3.02 for Windows (GraphPad Software, USA).

**Results and discussion**

Aerial parts of the plant species *Allium* were extracted by maceration method with methanol. After concentrating under the vacuum at 40°C with rotary evaporator, the yield of extraction was calculated (Table 2). The highest % yield is related to leaves of *A. atroviolaceum*. While the bulb of *A. jesdianum* Boiss. & Buhse has lowest % yield of extraction.

PV A (polyvinyl alcohol) was used to prevent the formation of particulate matters in the plant extract solutions in DMSO. Investigating the effect of blank DMSO/PV A solution on platelet aggregation confirmed that the presence of PV A at 1% concentration has no effect on platelet behavior. To the best of our knowledge, this is the first report for the application of PV A in determining anti-platelet aggregation activity of solutions which tend to form insoluble particles in DMSO.

Anti-platelet aggregation activity is not uncommon from plants extracts. Gady’s *et al* (2009) have reported a significant platelet aggregation inhibitory activity from the total aqueous extract of leaves of parsley (IC₅₀: 5.6mg/ml) against the aggregation induced by ADP (14). Although there are citations of antiplatelet aggregation activity of some *Allium* species (15-17), there are no previous reports, at least to our knowledge, on the activity of the genus on *in-vitro* platelet aggregation induced by AA and ADP. *Allium* extracts which were added to PRP 5min prior to their stimulation by AA (1.35mM) and ADP (5µM) inhibited platelets aggregation

<table>
<thead>
<tr>
<th>Yield of extraction</th>
<th>Scientific name</th>
</tr>
</thead>
<tbody>
<tr>
<td>7.3%</td>
<td>Allium ampeloprasum L.</td>
</tr>
<tr>
<td>5%</td>
<td>Allium hirtifolium Boiss.</td>
</tr>
<tr>
<td>3%</td>
<td>Allium hirtifolium Boiss.</td>
</tr>
<tr>
<td>6%</td>
<td>Allium haemanthoides Boiss. &amp; Reut</td>
</tr>
<tr>
<td>7%</td>
<td>Allium vavillivi M.Pop &amp; Vved.</td>
</tr>
<tr>
<td>8%</td>
<td>Allium atroviolaceum Boiss.</td>
</tr>
<tr>
<td>2.5%</td>
<td>Allium jesdianum Boiss. &amp; Buhse</td>
</tr>
<tr>
<td>2.7%</td>
<td>Allium shelkownikovii Grossh.</td>
</tr>
</tbody>
</table>

Table 2. Yields of methanolic extraction by maceration method.
in a concentration-dependent manner. The effects of extracts at different concentrations were measured and compared to each other. The results were expressed as IC₅₀ values with their respective 95% confidence limits (Table 3).

In the present study, among the eight Allium species tested, seven extracts (A. ampeloprasum, A. hirtifolium [Bulb], A. haemanthoides, A. vavillovi, A. atroviolaceum, A. jesdianum and A. shelkovnikovii) were found to possess satisfactory inhibitory effect on platelet aggregation against ADP. The IC₅₀ values for A. ampeloprasum, A. hirtifolium (Bulb), A. haemanthoides, A. vavillovi, A. atroviolaceum, A. jesdianum and A. shelkovnikovii extracts are presented in Table 3. According to Tukey’s multiple comparison test, the mentioned extracts showed significant differences in their IC₅₀ values (p < 0.05) except A. shelkovnikovii versus A. hirtifolium (Bulb). The IC₅₀ values of extracts increased in the following order: A. atroviolaceum < A. jesdianum < A. hirtifolium (Aerial part) < A. shelkovnikovii ≤ A. hirtifolium (Bulb). Based on the data obtained the A. atroviolaceum extract with IC₅₀ value of 0.4881 mg/ml (0.4826-0.4937) has maximum antiplatelet aggregation capacity induced by ADP.

However, A. hirtifolium (Aerial part) extract produced a weak activity on in-vitro platelet aggregation induced by ADP (5μM) while A. ampeloprasum (Aerial part), A. haemanthoides (Aerial part) and A. vavillovi (Aerial part) extracts caused a weak effect on in-vitro platelet aggregation induced by AA (1.35mM). They did not achieve ≥ 50% inhibition of platelet aggregation activity. The maximum inhibition (%) was 41.32% ± 3.15 for A. hirtifolium (Aerial part), 37.01% ± 4.34 for A. ampeloprasum (Aerial part), 31.85% ± 5.21 for A. haemanthoides (Aerial part) and 41% ± 3.67 for A. vavillovi (Aerial part) at maximum concentration of 1.25 mg/ml.

### Table 3. Effect of methanolic extract of Allium species on in-vitro platelet aggregation induced by AA (1.35mM) and ADP (5μM).

<table>
<thead>
<tr>
<th>Plant species</th>
<th>IC₅₀ (ADP) (mg/ml)</th>
<th>IC₅₀ (AA) (mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A. ampeloprasum (Aerial part)</td>
<td>0.6982 (0.6089-0.8005)</td>
<td>-</td>
</tr>
<tr>
<td>A. hirtifolium (Bulb)</td>
<td>0.8693 (0.7806-0.9681)</td>
<td>0.8765 (0.8765-0.8766)</td>
</tr>
<tr>
<td>A. hirtifolium (Aerial part)</td>
<td>-</td>
<td>0.6843 (0.5665-0.8266)</td>
</tr>
<tr>
<td>A. haemanthoides (Aerial part)</td>
<td>0.7366 (0.6856-0.7913)</td>
<td>-</td>
</tr>
<tr>
<td>A. vavillovi (Aerial part)</td>
<td>0.5524 (0.4997-0.5525)</td>
<td>-</td>
</tr>
<tr>
<td>A. atroviolaceum (Aerial part)</td>
<td>0.4945 (0.4137-0.5911)</td>
<td>0.4881 (0.4826-0.4937)</td>
</tr>
<tr>
<td>A. jesdianum (Bulb)</td>
<td>0.6607 (0.5140-0.8491)</td>
<td>0.5228 (0.5057-0.5404)</td>
</tr>
<tr>
<td>A. shelkovnikovii (Bulb)</td>
<td>0.6905 (0.4448-1.072)</td>
<td>0.8365 (0.7495-0.9336)</td>
</tr>
</tbody>
</table>

*Note: The IC₅₀ value of the positive control, quercetin, was measured as 0.1197 (0.1047-0.1362) mg/ml against AA and 0.1982 (0.1405-0.2131) mg/ml against ADP.

* Letters (a-e) denote homogenous subsets at p < 0.05 (Tukey’s post test).
function, play a critical role in the pathogenesis of some cardiovascular diseases such as arterial hypertension, atherosclerosis and subsequent ischemic events. This study highlights *Allium* anti-platelet properties. This finding is in agreement with other previous published data (15-17). We have shown that *in-vitro*, methanolic *Allium* extract inhibited in concentration-dependent manner, AA and ADP-induced platelet aggregations. Our results indicate that the maximum effect of anti-platelet aggregation was related to *A. atroviolaceum*. Therefore, *A. atroviolaceum* with an IC$_{50}$ value 0.4881 (0.4826-0.4937) mg/ml inhibits platelet aggregation induced by AA and to a lesser extent aggregation induced by ADP (p value < 0.05). However Hiyasat et al (2009) have reported that *A. ursinum* and *A. sativum* inhibit platelet aggregation induced via the ADP pathway and to a lesser extent the aggregation induced by epinephrine, whereas Aggregate Resources Act (ARA)-, collagen- and A23187-induced aggregations were not affected (15).

Medicinal plants are potential sources of lead compounds which can be used for further study and optimization as new drugs (18). Phytochemical analysis of Allium species has revealed the presence of polar compounds such as sulfur compounds and flavonoids, saponins and sapogenins (8). It is very unlikely that only one compound in the extract has been responsible for the observed anti platelet activity and it is more likely that a combination of compounds are involved in the exerted inhibitory effect, such as polyphenolic compounds that could prevent platelet functions. Additionally numerous *in-vitro* and *in-vivo* studies have shown that flavonoids inhibited primary homeostasis and many pathways associated with platelet activation and aggregation (14).

In conclusion, the present work demonstrated that *Allium* extracts could inhibit *in-vitro* platelet aggregation induced by AA and ADP. These results support the hypothesis that the dietary intake of *Allium* may be beneficial in normalizing platelet hyperactivation, in nutritional prevention of cardiovascular diseases which are potentially interesting in the development of new prevention strategies. Therefore, they are good candidates for further *in-vitro* and *in-vivo* studies to find potential lead compound for antiplatelet aggregation.

**Acknowledgements**

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**References**


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